Atopic Dermatitis and Fungi
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INTRODUCTION
Atopic dermatitis (AD) is a chronic, itching, inflammatory skin disease which is associated with asthma and/or hay fever and a familial occurrence of these conditions (53, 159). AD is the most common chronic skin disease in children (145, 160, 161). Approximately 18% of 7-year-old children have or have had AD. In 90% of patients the disease starts before 7 years of age, and usually it starts before 1 year of age. The majority of the patients improve before or around puberty, and it is estimated that approximately 2% of the adult population has AD (160, 161). However, AD may start in adult life, although this is rare. There is a documented increase in the incidence of AD in all countries with a Western life-style, but the exact hereditary pathway is still unknown (26a, 27, 161, 164). If both parents have had AD, the risk for the children is approximately 70%. Infiltrates of activated T-helper cells are found in the skin of patients with AD (92). There is an imbalance between Th1 and Th2 cells toward an increase in cells of the Th2 type. Th2 cells have a cytokine profile with an increased production of interleukin 4 (IL-4), which stimulates the plasma cells to an increased production of immunoglobulin E (IgE) (8). Another interesting observation in the skin of AD patients is the presence of a dendritic epidermal cell population (Langerhans’ cells and mast cells) that carry IgE receptors and specific IgE (22, 80, 92). Antigen presented by these cells may stimulate Th2 cells to synthesize IgE and hence contribute to the inflammatory changes in the skin (22, 80, 92).

When the skin of patients with AD is examined, it is often drier than the skin of an individual without AD (146, 164). It also reacts differently in response to stimulation. The white dermography appears when the skin is lightly scratched and is characterized by blood vessel constriction instead of dilation (72). The threshold for itch is also lower than in healthy individuals. Dry skin and the weakened barrier function in patients with AD are very important in the reactions of the patients to irritants and other external trigger factors including microor-
ganisms (72, 146, 152). The pathophysiology and etiology of AD have been reviewed and discussed in a supplement of the American Journal of Dermatology (54).

*Staphylococcus aureus* can be cultured from both involved and uninvolved skin in 85 to 90% of patients with AD (1, 21, 40, 44, 95, 110). *S. aureus* may aggravate AD due to a secondary infection (3, 84, 93, 94, 118). However, of greater importance is probably the presence of exotoxins and other substances from *S. aureus* that may act as allergens or, more importantly, as superantigens (59, 60, 89, 90, 139, 142, 168). Viral infections, especially with herpes simplex virus, may also aggravate AD (84).

The yeasts *Malassezia* and *Candida* may aggravate AD due to an allergic reaction (18, 40, 41, 135). In particular, the role of the *Malassezia* yeasts is now documented in several papers (18, 40, 41). *Malassezia* species are members of the normal human cutaneous flora, and a defect in the skin barrier may facilitate the contact of these yeasts with the immune system (41). *Candida* yeasts are members of the normal flora of mucous membranes, and the major contact with the immune system is through the gastrointestinal tract and, in women, even through the vagina (111, 135). Chronic dermatophyte infections are more common in patients with AD, and dermatophytes, especially *Trichophyton rubrum*, may act as allergens (79, 139). The role of other hyphomycetes in AD has not been studied in detail (79).

Other important factors are food, house dust mites, topically applied irritants, psychological factors, and climate (130).

The diagnosis of AD is usually based on many variables, including history and clinical and laboratory findings. Rajka was the first to make a diagnostic list consisting of major and minor criteria (116). These criteria were later revised and are known as the Hanifin and Rajka criteria (53). These criteria have been scientifically evaluated and found to work fairly well in the clinical situation even though many of the criteria have no precise definition, some are nonspecific, and some are uncommon. However, Williams et al. developed a minimum list of reliable criteria for the diagnosis of AD that could be used in clinical as well as in epidemiological studies. The obligatory criteria are an itching skin condition or report from parents that a young child itches or scratches as if itching. Thereafter, at least three of the following criteria should be fulfilled: history of involvement of the skin folds on the flexor sides of the elbows and knees, anteriorly on the ankles, on the throat, and on the cheeks in children younger than 10 years; history of asthma or hay fever in the patient or of atopic disease in the immediate family of children younger than 4 years; history of general skin dryness during the last year; visible eczema present on the flexor sides of the elbows and knees, anteriorly on the ankles, and on the throat (or cheeks, forehead, and extensor side of the extremities in children younger than 4 years); and onset before the age of 2 years (this criterion should not be used if the child is younger than 4 years) (159).

The clinical picture varies with age and may be described in 3 different phases: infant phase (younger than 2 years), childhood phase (2 to 12 years), and adolescent/adult phase (27). Important differential diagnoses are seborrheic dermatitis, psoriasis, rosacea and perioral dermatitis, fungal infections, ichthyosis vulgaris, scabies and contact dermatitis. Accurate assessment of the extent and severity of AD is essential for quantitating the baseline clinical disease burden and the effectiveness of treatment regimens being tested (56). One assessment system that is widely used is the SCORAD method (141), and another method is the EASI (56). It is beyond the scope of this article to discuss which one may be superior.

Treatment of AD is topical with emollients, corticosteroids, or immunomodulating agents (17, 40, 55, 91). The use of topical tacrolimus is a new approach to the treatment of AD (55). In severe AD, systemic treatment with corticosteroids and immunomodulating drugs is needed (17, 40, 91). UV treatment is also an effective alternative (65, 86). Topical and systemic antibacterial treatment is used in patients with signs of bacterial infections. Antifungal therapy is used in patients with signs of a fungal infection. However, a more widespread use of antymycotics based on the role of the most yeasts as common allergens is still not commonly recommended.

The prognosis of AD is good; 60 to 70% of patients recover during childhood (160, 161). The prognosis is less positive in women, in patients who had severe AD as children, and in those with concurrent allergic rhinitis and/or asthma or a positive family history of AD.
The genus *Malassezia* has in a taxonomic revision been placed in the kingdom Fungi, division Basidiomycota, class Hymenomycetes, order Tremellales and family Filobasidium uniguttulatum (61). Molecular biology using rRNA sequence analysis and nDNA comparisons, as well as G+C content in extracted DNA, has clearly divided the genus *Malassezia* into seven different species (48, 49). Today this genus consists of *M. furfur* (10), *M. pachydermatis* (140), *M. sympodialis* (48, 49), *M. globosa*, *M. slooffiae*, *M. restricta*, and *M. obtusa* (48, 49).

**Morphology and growth requirements.** The seven *Malassezia* species exhibit typical morphological and physiological features (48, 49). They reproduce by unilateral budding, leaving prominent scars on the mother cells (61). The yeast cells may be bottle-shaped, globose, ovoid, or cylindrical. Buds may be formed on a narrow or wide base. The most interesting feature of the genus *Malassezia* is the presence of a thick multilayered cell wall whose inner surface is corrugated. The protoplasmic membrane is firmly attached to the inside of the cell wall. The major components of the cell wall are mannoproteins (75 to 80%), lipids (15 to 20%), and chitin (1 to 2%) (105). It is thought that the immunomodulatory capacity of the *Malassezia* yeasts are related to cell wall lipids.

Using various physical, chemical and metabolic characteristics, the *Malassezia* species can be identified (48, 49). We have recently developed a modification of previous identification systems by which the seven different *Malassezia* species easily can be identified (Fig. 1). The advantages of this system are that it is easier and faster and that Tween reaction, which is often difficult to read, is not necessary. We always isolate the *Malassezia* yeast using the media described by Leeming and Notman (87). This medium contains, among other ingredients, ox bile, glycerol, glycerol monostearate, Tween 60, and whole-fat cow’s milk. Compared to other culture media, this was the most effective for isolation of the *Malassezia* yeasts (15).

Various morphological and physiological criteria can be used to characterize the *Malassezia* yeasts (Fig. 1) (48, 49, 61).
**M. pachydermatis** is nonlipophilic and can be cultured on lipid-free culture media. Cells are small and ovoid with a very broad base, and they never produce filaments. **M. furfur** can vary in micromorphology, producing cells that are long cylindrical ovoid or even globose. A few strains may produce filaments spontaneously or during special culture conditions. **M. furfur** is catalase positive, does not split esculin, grows on Dixon agar at 38°C, and is capable of growing in the presence of Cremophor. **M. sympodialis** has small ovoid cells. It is catalase positive, splits esculin, grows on Dixon agar at 38°C, and is not capable of growing in the presence of Cremophor. **M. slooffiae** has short cylindrical cells appearing ovoid under the light microscope with buds formed on a broad base. It is catalase positive, does not split esculin, grows on Dixon agar at 38°C, and is not capable of growing in the presence of Cremophor. **M. obtusa** has large cylindrical cells with a broad base. It is catalase positive, splits esculin, does not grow on Dixon agar at 38°C, and is not capable of growing in the presence of Cremophor. **M. globosa** has stable spherical cells. Buds are formed on a narrow base and might elongate to form germination tubes or even short filaments. It is catalase positive, does not split esculin, does not grow on Dixon agar at 38°C, and is not capable of growing in the presence of Cremophor. **M. restricta** has small spherical or ovoid cells with a relatively narrow base. It is catalase negative, does not split esculin, does not grow on Dixon agar at 38°C, and is not capable of growing in the presence of Cremophor.

**Distribution and isolation.** The Malassezia yeasts are members of the normal human cutaneous flora (36, 41, 46). However, there is great variation in the density and presence in various skin locations (35, 88), in children compared to adults (15, 33), and in normal skin compared to diseased skin (37, 45). The Malassezia yeasts are also associated with several skin diseases such as pityriasis (tinea) versicolor, Malassezia foliculitis, seborrheic dermatitis, atopic dermatitis, and some forms of confluent and reticulate papillomatosis (Gougerot-Carteaud syndrome) (33, 41). They may even be involved in systemic diseases (33, 120).

In 1951, Gordon isolated lipophilic yeasts from both normal healthy volunteers and patients with pityriasis versicolor (45, 46). Using direct microscopy or culture, Roberts found what he called *P. ovale* on the scalp and on the chest of 97% and 92 to 100% normal healthy adults, respectively (122). He also isolated the yeast in large numbers from patients with pityriasis versicolor (123). In a quantitative culture study, using a modification of the Williamson-Kligman scrub technique, Malassezia yeasts were cultured from clinically normal skin of the on the chest, back, upper arm, lower leg, and dorsal aspect of the hand (35). The highest count was found on the back, and the lowest was found on the hand. An easy semiquantitative method is to use contact plates (13, 15). This was first described in detail in 1987 (13). However, better results have later been obtained using a modification of the culture medium developed by Leeming and Notman (15, 87). Leeming et al. were able to culture the Malassezia yeasts from 20 different body sites, with the largest numbers being found on the scalp and upper trunk (88). We found in a culture study of adults that the density of the Malassezia yeasts decreased with increasing age (13). Although the number of yeast cells is larger in adults, the yeasts can also be cultured from children (15, 35).

Several studies have demonstrated that the Malassezia yeasts can be cultured in newborns. The Malassezia yeasts can be cultured not only from healthy skin but also from patients with various skin diseases (35, 36, 41). The number of yeasts is not always increased, but other mechanisms may be important in the pathogenesis. The yeast may induce immunological changes or behave as irritants, and then a reduction even of a relatively small number of yeast cells will result in an improvement. Quantitative or semiquantitative cultures can therefore be used to monitor the effect of treatment or changes in predisposing factors.

Only a few culture studies have been done using the new classification of the Malassezia yeasts (32, 51, 52, 76, 107). However, it is important to look at the distribution of the various Malassezia species not only on healthy skin but also on the skin of patients with Malassezia-associated skin diseases.

Nakabayashi et al. isolated various Malassezia species from the skin of healthy individuals and from patients with pityriasis versicolor, seborrheic dermatitis, and atopic dermatitis in Japan (107). The yeasts were cultured qualitatively using swabs and Dixon’s agar medium, and specimens for culture were taken from the scalp, face, and trunk. Cultures were negative in 14% (pityriasis versicolor) to 50% (healthy individuals) of cases. *M. globosa* was the most commonly isolated species from healthy volunteers, patients with pityriasis versicolor, and nonlesional skin of patients with AD. *M. furfur* was the most commonly isolated species from patients with seborrheic dermatitis and lesional skin of patients with AD.

In a study by Kim and Kim, the Malassezia yeasts were isolated qualitatively from various body regions in normal healthy volunteers from South Korea (76). All six lipophilic species were cultured. However, there was a great variation in the incidence of these yeasts, with *M. restricta* being the most commonly isolated species from the scalp and forehead and *M. globosa* being the most commonly species isolated from the chest and the back.

In two reports from Canada, Gupta et al. have isolated the various Malassezia yeasts from both healthy individuals and patients with various Malassezia-related skin diseases (51, 52). In the first study, the Malassezia yeasts were isolated qualitatively, in patients with pityriasis versicolor, by scraping skin flakes into a paper bag, and later the flakes were transferred to Leeming and Notman agar and to Littman agar supplemented with olive oil (51). A total of 111 samples were obtained from patients in Ontario, Canada, and 18 were from patients in other countries (South Africa, Hong Kong, Portugal, Brazil, and Hawaii). In the patients from Canada, *M. sympodialis* was isolated in 59%, *M. globosa* was isolated in 25%, and *M. furfur* was isolated in 11%. In patients from other countries, *M. globosa* was the dominating species, being cultured in 62%. In the other study, the Malassezia yeasts were isolated semiquantitatively, using contact plates with the Leeming and Notman agar (52), from both healthy volunteers and patients with pityriasis versicolor, seborrheic dermatitis, AD, and psoriasis. All the lipophilic Malassezia species were isolated. *M. sympodialis* was the dominant organism in healthy controls and in patients with pityriasis versicolor and AD, while *M. globosa* was the dominant species in patients with seborrheic dermatitis and psoriasis. The number of yeast colonies was largest in lesional skin from patients with pityriasis versicolor but was not statis-
tically significantly larger than in nonlesional skin or in skin from healthy individuals. However, the number of colonies of Malassezia was statistically significantly larger in both healthy individuals and patients with pityriasis versicolor than in patients with seborrhoeic dermatitis, AD, and psoriasis.

Erchiga et al. cultured the various Malassezia species qualitatively using Dixon agar from both lesional and nonlesional skin of patients with pityriasis versicolor in Spain (32). Cultures were positive in all patients. M. globosa, either alone or in combination with M. sympodialis, was the dominant species, being cultured in 97% of patients; M. sympodialis was cultured in 32%, either together with M. globosa or alone (only three patients). In nonlesional skin from the shoulders, Malassezia species were cultured in 47% of the patients and M. sympodialis was the dominant species, being cultured in 42% of all cultures.

According to these five reports, M. sympodialis, M. globosa, and M. furfur were the most commonly isolated Malassezia species. However, there were differences in the species that was most commonly isolated, not only between healthy individuals and patients with various diseases but also between results from different countries (32, 51, 52, 76, 107).

The Malassezia yeasts as pathogens. As mentioned above, the Malassezia yeasts are opportunistic pathogens, and under the influence of various predisposing factors, they are involved in several diseases (36, 41).

In pityriasis versicolor, under the influence of exogenous or endogenous predisposing factors, the yeast changes from the blastospore form to the mycelial form (38, 39). Pityriasis versicolor is a chronic superficial disease that usually affects the upper trunk, neck, or upper arms. It has a worldwide distribution. In tropical areas, it has been found in 30 to 40% of the population. However, the incidence is much lower in temperate climates. The diagnosis is based primarily on a typical clinical picture in combination with bright yellow fluorescence under Wood’s light examination and direct microscopy. Direct microscopy is of major importance, and the round or oval budding cells and hyphae can easily be identified. There are numerous ways of treating pityriasis versicolor, both topically and with systemic drugs (42). However, the high rate of recurrence, affecting 60% of patients after 1 year and 80% after 2 years, is an outstanding problem. Consequently, a prophylactic treatment regimen is necessary to avoid recurrence.

Malassezia folliculitis is a chronic disease characterized by pruritic follicular papules and pustules located primarily on the upper trunk, neck, and upper arms (7, 41). Under the influence of predisposing factors, Malassezia folliculitis may be explained by an extensive growth of Malassezia in the hair follicle. The inflammation may be due both to products of the yeast and free fatty acids produced as a result of the lipase activity of the yeast (7, 41, 43). The diagnosis is based on a typical clinical picture of itchy papules and pustules as the predominant symptoms, direct microscopy, and the effect of anticycotic treatment (7, 41). The effect of anticycotic treatment is often dramatic. Most cases respond well to topical treatment, but systemic treatment with e.g. itraconazole or fluconazole has been used in difficult cases (42). Lesions and itching recur in most patients if treatment is not maintained intermittently. Therefore, a prophylactic treatment schedule, such as topical treatment once or twice a week, is recommended.

Seborrhoeic dermatitis is characterized by red scaly lesions located predominantly on the scalp, face, and upper trunk. There are now many studies indicating that the Malassezia yeasts play an important role in seborrhoeic dermatitis (6, 14, 36, 41, 42, 43). Many of these are treatment studies which describe the effectiveness of anticycotics, paralleled by a reduction in number of the Malassezia yeasts, whereas recolonization leads to a recurrence of seborrhoeic dermatitis. The increased incidence of seborrhoeic dermatitis in patients with immunosuppressive disorders suggests that the relationship between Malassezia and the immune system is important, but the exact mechanism involved is still unknown. Studies designed to investigate this mechanism in patients with seborrhoeic dermatitis fall into four areas (6, 43): the humoral response to Malassezia, lymphocyte transformation response to Malassezia extract, T helper (CD4)/T suppressor (CD8) cell ratios, and cytokine responses. However, studies in all of these areas have given inconsistent results, and the local reaction in the skin may give a better answer. In a recent study, the number and type of inflammatory cells and mediators from lesional and healthy skin in patients with seborrhoeic dermatitis were studied (43). Staining was often more intense when Malassezia yeasts cells were present. An increase in NK- and CD16-positive cells in combination with complement activation indicates that an irritant or nonimmunogenic stimulation of the immune system is important. With the interleukins, an increase in the production of inflammatory interleukins and an increase in the regulatory interleukins for both Th1 and Th2 cells were seen. The immune response described for cutaneous C. albicans infections (126) has similarities to the skin reactions found in seborrhoeic dermatitis (43).

Atopic Dermatitis and the Malassezia Yeasts

Distribution of the Malassezia yeasts. Many reports clearly indicate the role of Malassezia in AD, especially AD located to the head and neck area in adults (19, 26, 29, 30, 41, 62, 66, 69, 74, 83, 98, 109, 125, 131, 132, 135, 136, 137, 148, 149, 154). The Malassezia yeasts have been cultured from patients with AD with the same frequency as from healthy individuals (19, 41, 52). In one study of children from newborn to the age of 20 years, the Malassezia yeasts were isolated from lesional and nonlesional skin, from the same region, as well as from normal skin of the forehead of patients with AD (19). Normal skin on the forehead in age- and sex-matched individuals with rhinoconjunctivitis and/or asthma and healthy volunteers served as controls. Cultures did not differ significantly among the three groups or between lesional and nonlesional skin. There was, however, a difference between the age groups. Cultures were positive in 5 to 15% of children aged up 10 years and in 65 to 100% of 11- to 20-year-old persons. In a study of adult patients with AD involving the scalp and neck, Malassezia samples were taken from lesional skin of the forehead and cultured on Leeming and Notman agar in contact plates (20). Culture was positive in 83% of cases.

These two studies were performed before the new taxonomy for the Malassezia yeasts was introduced. In the above-mentioned study by Nakabayashi et al., they were able to classify the Malassezia yeasts (107). Samples were taken from lesional and nonlesional skin from the scalp, face, and trunk. In lesional
skin the most commonly isolated species was *M. furfur*, which was cultured in 21% of samples. *M. globosa* was cultured in 14%, *M. sympodialis* was cultured in 7%, *M. slooffiae* was cultured in 4%, and cultures were negative in 46%. In nonlesional skin, the most commonly isolated species was *M. globosa* (33%); *M. furfur* was cultured in 11% of samples; 22% were unknown species or contaminated cultures, and 33% of samples were culture negative.

Gupta et al. cultured the *Malassezia* yeasts semiquantitatively, using the Leeming and Notman culture medium in contact plates, from patients with various skin diseases and healthy controls (52). In patients with AD, the total number of isolated colonies was significantly larger from nonlesional skin than from to lesional skin. The number of colonies from both lesional and nonlesional skin was significantly smaller than the number of colonies from healthy individuals, patients with pityriasis versicolor (both lesional and nonlesional skin), and nonlesional, but not lesional, skin of patients with seborrheic dermatitis. In AD there were no differences in the percentages of the various *Malassezia* species isolated from lesional and nonlesional skin. *M. sympodialis* was the most commonly isolated species, being cultured in 51% of the patients, *M. globosa* was cultured in 18%, *M. furfur* and *M. obtusa* were each cultured in 10%, *M. restricta* was cultured in 8%, and *M. slooffiae* was cultured in 3%. The majority of *Malassezia* yeasts were cultured from the forehead and the trunk. In an English abstract from a paper otherwise written in Russian, Arzumanian stated that *M. sympodialis* was the most commonly isolated species isolated from both patients with AD and healthy individuals (5).

In a recently completed multicenter study in Sweden, involving the Departments of Dermatology in Gothenburg, Lund, and Stockholm and the Department of Clinical Immunology, Karolinska Hospital, Stockholm, we cultured the *Malassezia* yeasts from 125 patients with AD, 16 with seborrheic dermatitis, and 30 healthy controls (69a). Cultures were performed using the Leeming and Notman agar (87) in contact plates from normal-appearing skin of the upper back and, in AD and seborrheic dermatitis patients, also from lesional skin. Cultures were positive in 56% of patients (70 of 125) with AD, 88% of patients with seborrheic dermatitis, and 83% of healthy controls. In patients with AD, *M. sympodialis* was cultured in 40%, *M. obtusa* was cultured in 25%, *M. globosa* was cultured in 23%, *M. slooffiae* was cultured in 6%, *M. furfur* was cultured in 4%, and *M. restricta* was cultured in 2%. The same frequency of the *Malassezia* yeasts was seen in healthy controls. However, in patients with seborrheic dermatitis, *M. obtusa* was cultured in 30% and *M. sympodialis* was cultured in 30%.

**Immunology.** (i) Skin prick test for type I hypersensitivity. A positive skin prick test (SPT) was first reported in 1958 among a group of AD patients (65). In 1983, Clemmensen and Hjorth described a positive effect of oral ketoconazole in the treatment of adult patients with AD localized to the head and neck area (74). The SPT was positive in 45% of patients with AD but without the head and neck distribution, and patients with seborrhoeic dermatitis were negative. These percentages are much higher than those obtained by Waersted and Hjorth. However, several children were included in the report by Waersted and Hjorth. The *Malassezia* extract used in this and the two earlier studies is a protein extract prepared by ALK Laboratories, Copenhagen, Denmark. The same extract has also been used for a histamine release assay (Lucotest-HR; Lundbeck Diagnostics, Copenhagen, Denmark) measuring the release of histamine from basophils after stimulation with the antigen. The results of histamine release after stimulation with *Malassezia* extract paralleled the results obtained with the SPT (74).

Rokugo et al. found that the results of the SPT vary according to the age of the patients (125). Among AD patients younger than 10 years, 39% were positive in SPT, while 64% of patients older than 10 years were positive. Wessels et al. found positive SPTs to *Malassezia* in 84% of adult patients with AD (157). In a study by Broberg et al., children with AD, rhinitis and/or asthma without AD, and healthy controls were investigated (19). A total of 60 children in each category were included, and the children were divided into four groups: 0 to 5 years, 6 to 10 years, 11 to 15 years, and 16 to 21 years, with 15 in each group. SPTs to *Malassezia* were positive in 23% of patients with AD, 0% of patients with rhinitis and/or asthma, and 8% of healthy controls (three subjects). Among the patients with AD, 1 of 15 patients in the age group from 0 to 5 years was SPT positive compared to 6 of 15 (40%) in the age group from 16 to 21 years.

Kim et al. have performed an SPT with an *M. furfur* extract in 80 young adult patients with the head and neck distribution of AD and compared the results to those obtained for controls without AD (75). A total of 45% showed positive SPT responses, and the severity score of the head and neck lesions in the *M. furfur* SPT-positive group was significantly higher than in the *M. furfur*-negative group. In another study, Devos and van der Valk studied patients with AD, with and without the head and neck distribution, using SPT with a *Malassezia* extract and other procedures (28). In a total of 92 patients with AD (the majority were young adults), the SPT was positive in 40%. However, only 6% of the patients with the head and neck distribution of AD were positive. This is contradictory to all earlier studies, in which patients with the head and neck distribution of AD showed the largest number of positive SPTs to *Malassezia* (19, 26, 74, 154).

In the above-mentioned Swedish multicenter study, we also performed SPT with extracts of *Malassezia*, the recombinant allergens Mal f 1, Mal f 5, Mal f 6, and a mix of these recombinant allergens (96, 137) in patients with AD, patients with seborrheic dermatitis, and healthy controls; 74% of the AD patients had the head and neck distribution. We found that 57% of the AD patients were positive to the extract or recombinant allergens; 52% were positive to the *Malassezia* extract. However, 56% of patients with the head and neck distribution of AD were positive to the extract compared to only 36% of the patients without the head and neck distribution of AD. None of the patients with seborrheic dermatitis or the healthy
controls were positive. All readings were done in a controlled double-blind design.

In summary, positive SPT to Malassezia is found primarily in adult patients with AD and especially in adult patients with the head and neck distribution of AD (Table 1).

(ii) Specific IgE antibodies to the Malassezia yeasts. The first paper describing specific IgE antibodies to the Malassezia yeasts was published in 1991 (69). Specific antibodies were found especially in patients with AD and less commonly in patients with other atopic manifestations. This was later verified by other groups using various extract from Malassezia (19, 29, 30, 66, 69, 157, 166). In the study by Broberg at al., specific IgE antibodies were found in eight children with AD, five in the age group from 16 to 21 years, and only one patient with rhinitis and/or asthma (19). In another study with children (4 to 16 years old), 22% of patients with AD had IgE antibodies against Malassezia (97). In these patients, the AD was generally worse than in those without specific Malassezia IgE antibodies. The head and neck distribution of AD was also more common and the onset of AD occurred earlier in life than in specific IgE-negative children.

In the study by Kim et al., 68% of patients with the head and neck distribution of AD had specific IgE antibodies against Malassezia (75). The clinical severity and total IgE level in serum for these patients were greater in patients with Malassezia-specific IgE antibodies than in patients with a negative response. Devos and van der Valk also studied the presence of specific IgE antibodies in patients with AD (28). However, in contrast to their SPT results, mentioned above, they found that 100% of patients with the head and neck distribution of AD had Malassezia-specific IgE antibodies compared to only 14% of AD patients without the head and neck distribution.

Savolainen et al. recently described specific and nonspecific humoral cellular and cytokine responses to Malassezia and Candida in AD patients (135). Fifteen AD patients and 7 healthy controls were included. The total IgE level in serum was clearly higher in AD patients than in controls. The Malassezia- and C. albicans-specific IgE level was also significantly increased in AD patients compared to controls. Malassezia was a stronger inducer of IgE responses, and C. albicans more strongly induced IgG responses. However, there were no significant differences in Malassezia- and C. albicans-specific IgG levels between the groups. The rest of the study is discussed in the section on cellular and interleukin responses (see below).

Mayer and Gross have studied the presence of specific IgE antibodies against various Malassezia species (104). The presence of specific antibodies against M. sympodialis, M. furfur, and the commercially available Malassezia extract was studied in patients with AD, seborrhoeic dermatitis, pityriasis versicolor, and hymenoptera allergy and healthy controls. The allergens alone as well as various combinations were included. It was found that 35% of AD patients had specific IgE antibodies against one or several allergens, 12% of patients with seborrhoeic dermatitis were positive, 8% of patients with hymenoptera allergy were positive, and none of the patients with pityriasis versicolor or the healthy controls were positive. For patients with AD, the largest number of positive results was seen in patients with the head and neck distribution of AD. There was also strong correlation between specific IgE antibodies and the total IgE level. Differences in the type and extent of sensitization were also found among the Malassezia yeasts. M. sympodialis and the commercially available Malassezia extract were almost equally positive, while the frequency of sensitization to M. furfur was lower. The commercially available Malassezia was typed using the new classification and found to be M. sympodialis. M. sympodialis is more common on the skin of patients with AD and it therefore a better allergen for testing than M. furfur.

In the Swedish multicenter study, mentioned above, specific serum IgE antibodies to the Malassezia extract were found in 45% of patients with AD; again, patients with the head and neck distribution of AD had the highest number of reactions (55%); 40% of the AD patients had both specific Malassezia IgE antibodies and positive SPT. Patients with seborrhoeic dermatitis and healthy controls were negative. RMal f 1, RMal f 5, and RMal f 6 specific serum IgE antibodies were found in 19, 28, and 25%, respectively. Specific serum IgE-antibodies to the recombinant Malassezia antigens, alone or together, were found in 40% of the patients. In contrast to the extract, moderately strong correlations were found between SPT (in millimeters) and specific serum IgE level for rMal f 1, rMal f 5, and rMal f 6.

Specific Malassezia IgE antibodies were found in 20 to 100% of patients with AD. The prevalence was lower in children, higher in adults, and highest in adult patients with the head and neck distribution of AD. Table 2 summarizes the specific Malassezia serum IgE antibodies found in various studies.

(iii) Characterization of antigens. Several IgE binding components of Malassezia have been isolated (29, 30, 62, 66, 69, 70, 98, 131, 132, 137, 149, 157, 166, 167). Three major allergen components from Malassezia were later identified using monoclonal antibodies (166). Two of them, a 67-kDa component and a 37-kDa component, were proteins, and one, a 14-kDa component, was probably of carbohydrate origin. In other studies, allergen cross-reactivity between Malassezia and C. albicans has been found (29, 30, 98, 131). According to Zargari et al., the 14-kDa component may be found in both yeasts.
Malassezia al. have described protein components common to both drate components of the cell wall (66). However, Savolainen et al. found 15-kDa components that probably are nonprotein carbohy-

various components of Malassezia (166). Jensen-Jarolim et al. have also tried to characterize Johansson et al., 1991 (69) 19

day 4. On the other hand, the carbohydrate content remained period may also be of importance (167). The protein content of Malassezia was found to be the most important inducer of IgG antibodies. C. albicans

itor of the IgE-binding components. In contrast, Devos et al., 2000 (28) 100

Kim et al., 1999 (75) 68

Nissen et al., 1998 (109) 93

Back et al., 1995 (8) 44

Lindgren et al., 1995 (97) 22

Broberg et al., 1992 (19) 13

TABLE 2. Speciﬁed IgE serum antibodies to the Malassezia yeasts in patients with AD

<table>
<thead>
<tr>
<th>Study</th>
<th>% of AD patients with specific Malassezia IgE antibodies</th>
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<tr>
<td></td>
<td>Children</td>
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<tr>
<td>Johansson et al., 1991</td>
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<td>Broberg et al., 1992</td>
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<td>Lindgren et al., 1995</td>
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<td>Scalabrin et al., 1999</td>
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<td>Mayser and Gross, 2000</td>
<td>34</td>
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<tr>
<td>Savolainen et al., 2001</td>
<td>60</td>
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<tr>
<td>The Swedish multicenter study</td>
<td>45</td>
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</table>

(166). Jensen-Jarolim et al. have also tried to characterize various components of Malassezia, and they described 9- and 15-kDa components that probably are nonprotein carbohydrate components of the cell wall (66). However, Savolainen et al. have described protein components common to both Malassezia and C. albicans (131, 132). In 1997, Lintu et al. have described IgE antibodies to both protein and mannann antigens of Malassezia in sera from patients with AD (98). Lintu et al. found other major components from those found by Zargari et al. In another paper, Lintu et al. described cross-reacting IgE and IgG antibodies to both Malassezia, C. albicans, Saccharomyces cerevisiae, and, to some extent, Cryptococcus albidus (100). In the IgE response, the main cross-reacting pattern was the mannann region and Malassezia was the most potent inhibitor of the IgE-binding components. In contrast, C. albicans was found to be the most important inducer of IgG antibodies. There are several explanations for the different results obtained. The technique used is important, and the strain of Malassezia used is also important. The length of the culture period may also be of importance (167). The protein content of the extract was highest after 2 days but gradually declined after day 4. On the other hand, the carbohydrate content remained fairly constant. The stability of the extracts is also influenced by the storage period and conditions (99). The stability of Malassezia protein extracts stored, even at 4°C, for more than 1 month was poor. The 9-kDa component was more stable than the 20-kDa component and, the 96-kDa bands in particular were far more labile. In none of these reports were the Malassezia yeasts classiﬁed according to the new taxonomy.

Antigenic bands against both M. sympodialis and M. furfur were found in sera from patients with AD (103). The molecular masses of the most important bands were 15, 22, 30, 37, 40, 58, 79, 92, 99, and 124 kDa for M. sympodialis and 15, 25, 27, 43, 58, 92, 99, and 107 kDa for M. furfur. Control sera were negative. When M. furfur-positive sera were preincubated with M. sympodialis, the clear band of 15 kDa disappeared, indicating a possible cross-reaction of the antibodies. However, this study clearly demonstrates that there are differences between different strains of Malassezia and that differences in results may be due to the use of different strains.

Schmidt et al. have identiﬁed the complete cDNA sequence and expression of one major allergen protein of Malassezia (137). According to the WHO/IUS nomenclature, this recombinant allergen is named Mal f 1. Yasueda et al. have isolated two additional major recombinant allergens, named Mal f 2 and Mal f 3 (165), and Onishi et al. have isolated the Mal f 4 recombinant allergen (114). Recently, Rasool et al. isolated and characterized Mal f 7, Mal f 8, and Mal f 9 (119). However, like the allergen Mal f 1, these three recombinant allergens were prepared from the ATCC strain 42132 of M. furfur (formerly P. orbiculare). This Malassezia strain is not M. furfur but M. sympodialis (104). I was the one who donated this strain to the ATCC in 1979, and I have now retyped it according to the new taxonomy. According to this, I have also retyped it as M. sympodialis. These recombinant allergens should therefore be named Mal sy (1 etc.). The reference strain used by Yasueda et al. (165) and by Onishi et al. (114) are from another reference laboratory (strain 2782 from Teikyo Institute for Medical Mycology) and is named M. furfur. I have not personally conﬁrmed that this species is M. furfur. Recently, Koyma et al. isolated three major components of M. globosa from extracts of this yeast (82). These antigens were named Mal g 46a, Mal g 46b, and Mal g 67. Mal g 46b was found to be the major antigen, and it reacted only with sera from patients with AD and with none of the sera from patients with seborrheic dermatitis, pityriasis versicolor, or chronic urticaria or healthy controls. Mal g 46b reacted with 69% of the sera from patients with AD. Mal g 46b reacted with 83% of sera containing IgE antibodies to M. globosa. In a lectin blot with concanavalin A, both Mal g 46 a and Mal g 46 b reacted with concanavalin A, indicating that these antigens were glycoproteins containing mannose chains in the molecules. When species speciﬁcity was tested, the antibody reacted strongly with M. globosa. A slight reaction was seen with M. restricta, but no reaction was seen with M. sympodialis, M. furfur, or M. slooffiae. Also, no reaction was seen with C. albicans. In the Japanese population, this antigen may be more important that antigens against, e.g., M. sympodialis, because M. globosa is more commonly cultured from AD patients in Japan (107).

(iv) Cellular and cytokine responses. Both in vitro and in vivo studies of cellular and cytokine responses have been performed. In parallel to SPT, several skin patch test experiments have been performed for investigation of a delayed-type hypersensitivity to the Malassezia yeasts. Rokugo et al. found many patients with AD who were patch test positive to a Malassezia extract (125). However, they used the chamber scarification method, and Kieffer et al., using a standard test procedure without scarification, were not able to reproduce these large numbers of positive results (74).

Patch test reactions are often difficult to read in patients with AD. To evaluate the results of primarily aeroallergens, a special technique called the atopy patch test (APT) was developed (151). Briefly, the stratum corneum is stripped approximately 15 times with cellophane tape to achieve a slight increase in its permeability. The allergens are then applied on paper disks in Finn chambers (8 mm; Epitest Ltd Oy, Tuusula, Finland) on normal-appearing skin of the back and covered with a nonirritating tape. The disks are removed after 48 h, and the skin reaction is evaluated 24, 48, and 72 h after that. In a study by Tengvall-Linder et al., using this technique, 8 of 15 adult pa-

patients with AD were patch test positive to a Malassezia extract (151). However, none of 8 patients with seborrheic dermatitis
or 15 healthy volunteers were positive. The AD patients with positive APT reactions had significantly higher specific *Malassezia* serum IgE antibody levels than did AD patients with negative reactions. *Malassezia*-specific IgE, but not total IgE, levels correlated with the degree of APT response at 48 h in the whole group of AD patients. However, specific serum IgG levels did not differ significantly between the two groups of AD patients.

In the above-mentioned Swedish multicenter study, patients with AD, patients with seborrheic dermatitis, and healthy controls were also subjected to APT with both the crude *Malassezia* extract and extracts containing the recombinant allergens Mal f 1, Mal f 5, and Mal f 6, both alone and in combination. Positive APT reactions were found in 38% of patients with AD, 7% of patients (1 patient) with seborrheic dermatitis, and none of the healthy controls. For the AD patients, positive APT reactions to rMal f 1, rMal f 5, and rMal f 6 were found in 6, 15, and 10%, respectively. No positive reactions were seen in patients with seborrheic dermatitis or in healthy controls. The APT reactions differed among these three recombinant allergens. rMal f 5 gave the strongest reactions, and 75% of these patients were also SPT positive and had specific serum IgE levels.

Kröger et al. have measured the synthesis of various lymphokines as well as the IgE synthesis from peripheral blood mononuclear cells after stimulation with a *Malassezia* extract (83). They found that IL-4 and IL-10 synthesis was increased and IL-2 and gamma interferon (IFN-γ) synthesis decreased in patients with AD and with RAST*-specific serum IgE antibodies compared to healthy controls. IgE synthesis in the supernatants was only increased in patients with AD and with RAST*-specific serum IgE antibodies.

The proliferative response in peripheral blood mononuclear cells (PBMC) to a *Malassezia* extract has been studied in 10 AD patients with specific serum IgE antibodies to *Malassezia* and in 6 healthy controls with no specific serum *Malassezia* IgE antibodies (148). The PBMC response to *Malassezia* was significantly higher in the AD group than in the control group. A response to a *C. albicans* extract was either not measurable or very low. T cells were obtained from peripheral blood and the skin by cloning from one AD patient with severe lesions and a high titer of specific *Malassezia* serum IgE antibodies. Several T-cell clones (TCC) from lesional skin showed a strong response to a *Malassezia* extract, whereas only a few TCC from peripheral blood were positive and none from nonlesional skin were positive. The response to a *C. albicans* extract was very low.

In the same study, freshly isolated PBMC produced only low cytokine levels of IL-4, IL-5, and IFN-γ after 12 days of incubation with a *Malassezia* extract. When blood- and skin-derived *Malassezia*-reactive CD4+ TCC were tested for their production of IL-4, IL-5, and IFN-γ after stimulation with phytohemagglutinin (PHA) and an anti-CD3 antibody, a variation of cytokine profile was produced. Th1, Th2, and Th0, cytokine profiles and a Th2/Th0 cytokine profile were found. The Th2-like profile of *Malassezia*-reactive TCC derived from lesional skin indicates that *Malassezia* may play a role in maintaining IgE-mediated skin inflammation in AD. Incubation of TCC with a *Malassezia* antigen for 12 days induced measurable production of IL-5 but not IL-4 or IFN-γ. Nonstimulated and *Malassezia*-stimulated PBMC expressed similar levels of IFN-γ mRNA at all time points. Stimulation of PBMC with *Malassezia* induced a time-dependent increase in gene expression for both IL-5 and IL-13, with maximal expression at 24 h. Gene expression for these cytokines was hardly detectable in nonstimulated PBMC. Stimulation with anti-CD3 antibody induced a profound increase in the expression of genes for IFN-γ, IL-5, and IL-13 at 16 to 24 h.

The same group has compared the immune response of AD patients to a *Malassezia* extract with that of healthy controls (150). The proliferative response of PBMC was significantly higher in the AD patients than in the healthy controls. Significantly higher levels of IL-5 was produced by PBMC from AD patients compared to healthy controls. *Malassezia*-reactive T-cell lines obtained after stimulation of PBMC by *Malassezia* antigen produced significantly higher levels of IL-4 and IL-5 after stimulation with anti-CD3 antibody and showed a higher IL-4/IFN-γ ratio in AD patients than in healthy controls. These results indicate that the *Malassezia* yeasts may play a role in maintaining skin inflammation in AD.

Recently, Buentke et al. looked at the possible interaction of immature and mature antigen-presenting monocyte-derived dendritic cells (MDDCs) with *Malassezia* yeast cells and different *Malassezia* allergenic components (23). Internalization of *Malassezia* yeast cells and yeast components by immature MDDCs was found, and the results showed that a median of 94% of the immature CD1+ MDDCs were *Malassezia* whole-cell extract positive, 81% were positive to the recombinant antigen rMal f 5, and 93% were mannan positive. However, mature MDDCs were significantly less positive. The binding of the *Malassezia* extract and mannan was inhibited in a dose-dependent manner by methyl-a-d-mannopyranoside, suggesting uptake via the mannose receptor. This study demonstrated that human immature CD1+ MDDCs can efficiently take up *Malassezia* and allergenic components from the yeast in the absence of IgE, implying that sensitization of AD patients to *Malassezia* can be mediated by immature dendritic cells in the skin. In their paper, the authors claimed that the *Malassezia* species used was *M. furfur*. However, the species was the above-mentioned ATCC isolate 42132, which both by myself and by Mayser and Gross (103), has been retyped as *M. sympodialis*. Specific IgE antibodies to *M. sympodialis* are more commonly seen in AD patients than are specific IgE antibodies to *M. furfur* (101).

Savolainen et al. have recently tried to characterize the *Malassezia*- and *C. albicans*-specific and -nonspecific humoral, lymphoproliferative, and cytokine responses in AD patients compared to healthy controls (135). *Malassezia*- and *C. albicans*-specific IgE levels and *Malassezia*-induced PBMC proliferation were elevated in AD patients. Both *Malassezia*- and *C. albicans*-specific IgE antibodies were detected in AD patients but not in healthy controls. The median titer of specific IgE response was significantly higher for *Malassezia* than for *C. albicans*. The IL-4/IFN-γ ratio induced by *Malassezia* was higher than that induced by *C. albicans*. The PHA-induced IL-2 and IL-4 responses and the *C. albicans*-induced IL-5 response and IFN-γ response were elevated in AD patients. Several correlations were found between total and yeast-specific IgE levels in serum, *Malassezia*-specific lymphoproliferation, PHA-induced IL-2, IL-4, and IL-5 levels, and *C. albicans*-induced IL-5 levels. The cytokine profiles found in this study for *Malassezia* were in parallel with and close to the results...
found by Tengvall-Linder et al. (149), and they support the role of a combination Th0-, Th1-, and Th2-responding *Malassezia* cells in the pathogenesis of AD. Differences in *Malassezia* responses obtained may occur because different *Malassezia* species were studied. In the study by Savolainen et al., *Malassezia* appears to be associated more with IL-4 responses and *C. albicans* appears to be associated more with IFN-γ responses (135).

So far, the results of cytokine production of lymphocytes have been mentioned. In a paper by Watanabe et al., cytokine production by human keratinocytes after stimulation with cells of various *Malassezia* species has been described (156). The *Malassezia* species studied were *M. pachydermatis*, *M. furfur*, *M. slooffiae*, and *M. sympodialis*, and the cytokine production studied was that of IL-1β, IL-6, IL-8, monocyte chemotactic protein-1 (MCP-1), and tumor necrosis factor-alpha (TNF-α). Cytokine production by keratinocytes after coculture with *M. furfur* was low or undetectable. In the supernatants cocultured with the other *Malassezia* yeasts, IL-1β, IL-6, IL-8, and TNF-α were detected whereas the MCP-1 levels were low or undetectable. The cytokine levels were highest with *M. pachydermatis*. *M. pachydermatis* is cultured primarily from animals, and the dermatitis seen in animals is often more inflammatory than the human *Malassezia*-related diseases. There were even differences between the *Malassezia* yeasts in the number of yeast cells that were needed to start cytokine production.

Further studies are needed to explain differences in results between AD and other skin diseases as well as differences between the various *Malassezia* yeasts. A number of results for lymphocyte stimulation responses and cytokine production have been reported in the literature. Kroger et al. (83) found an increase in IL-4 and IL-10 production and a decrease in IL-2 and IFN-107 production in AD patients after stimulation with *Malassezia* extract; Tengvall-Linder et al. (150) and Savolainen et al. (135) found an increase in IL-4 and IL-5 production.

**Treatment.** In a study by Gupta et al., in the intra susceptiibility of the seven different *Malassezia* species against the azoles, ketoconazole, itraconazole and voriconazole and the allylamine terbinafine was tested (50). All of the *Malassezia* yeasts were highly sensitive to all threeazole drugs. However, only *M. pachydermatis* and *M. sympodialis* were highly sensitive to terbinafine.

A double-blind crossover study by Clemmensen and Hjorth has shown that in patients with AD, oral ketoconazole is statistically more effective than placebo (26). In an open study by Back et al. ketoconazole at 200 mg once daily for 2 months followed by 200 mg twice weekly for 3 months resulted in an improvement in clinical score in AD patients (8). There was also a reduction in the levels of both total IgE and specific IgE for *Malassezia* and *C. albicans*. There was a correlation between the reduction in the levels of total IgE and specific IgE for *Malassezia* but not between reduction in clinical score and the level of specific IgE for *Malassezia*. However, for specific *C. albicans*, there was a correlation between a reduction in the specific IgE and total IgE levels and clinical score.

Today oral ketoconazole should not be used for long-term treatment due to the risk of liver toxicity. If ketoconazole is used for more than 14 days, liver enzymes should be monitored before the start of treatment and every 14 days during treatment. The newer azoles, the triazoles fluconazole and itraconazole, have less risk of side effects; although they are more expensive, they should be the drugs of choice.

In a double-blind placebo-controlled study, Back and Bartosik have treated 29 AD patients with oral ketoconazole at 200 mg once daily or placebo for 3 months (9). In the ketoconazole group, the levels of total IgE and specific IgE to both *Malassezia* and *C. albicans* decreased significantly. The patients were allowed to use topical corticosteroids during the study period, and the clinical score decreased equally in both groups. However, the improvement was correlated with the use of topical corticosteroids in the control group but not in the ketoconazole group.

Lintu et al. have, in a double-blind, controlled study, treated 80 AD patients with oral ketoconazole at 200 mg or placebo once daily for 30 days (101). Yeast cultures from the skin and pharynx, SPT and specific IgE antibodies for *Malassezia*, *C. albicans*, and *S. cerevisiae*, the total IgE level in serum, and the clinical severity score was assessed at baseline and after 1 and 3 months. A significant improvement was seen in the clinical score after 30 days of treatment in the ketoconazole group but not in the placebo group. The number of positive *Malassezia* cultures was also reduced significantly in the ketoconazole group but not in the placebo group. The clinical response was highest in female patients with positive cultures for *Malassezia* and/or *C. albicans*. In this study there was a slight, but not statistically significant, reduction in the total IgE level after 30 days of treatment with ketoconazole. For all other parameters studied, there were no significant difference between baseline and the 2- or 3-month follow-up visits. This may be due to a shorter treatment period than in earlier trials.

In one study, the positive effect of oral itraconazole in the treatment of patients with AD has been mentioned (66). Topical treatment with bifonazole, in an unknown number of patients, has also been effective in the treatment of AD (144). In an open study, the combination of hydrocortisone and miconazole was effective in the treatment of AD (158). However, in a double-blind controlled study of the effect of hydrocortisone and miconazole compared with hydrocortisone alone in the treatment of AD, no statistically significant difference was found and both treatments were effective (20). Oral treatment may be more effective because it more effectively eradicates the *Malassezia* yeasts located deep in the follicle.

The number of treatment studies using antifungal therapy is small, and new well conducted, controlled and double-blind trials are needed to confirm if antifungal therapy is beneficial in the treatment of some patients with AD.

**Candida**

More than 50 different *Candida* species have been described (111). Many species have been isolated from human sources, but eight species are dominant these are *C. albicans*, *C. glabrata*, *C. guilliermondii*, *C. kefyr*, *C. krusei*, *C. parapsilosis*, *C. tropicalis*, and *C. doblinensis*. All the pathogenic *Candida* species multiply primarily by the production of buds from blastospores. The sizes and shapes of blastospores are sometimes characteristic for a species, but the gross microscopic appearance is more or less similar. The cell wall is multilayered, and the cell membrane is a typical eukaryotic phospholipid bilayer with frequent invaginations. The most commonly encountered pathogenic *Candida* species usually grow well in aerobic cul-
tures base on rich or poor nutrient media with a pH in the range of 2.5 to 7.5 and at a temperature in the range of 20 to 38°C.

**Distribution on normal skin.** In persons working in hospitals, in patients with skin diseases, in cancer patients, in patients with immune defects, and in individuals living in a warm and humid climate *C. albicans* may be cultured even from glabrous skin. Other *Candida* species, e.g., *C. parapsilosis*, are often cultured from the toe cleft.

**Distribution on mucous membranes.** Isolation of *Candida* from the gastrointestinal tract or, in women, the vagina does not automatically imply disease because many healthy persons are colonized with *Candida* in these areas (85, 111, 129). An prevalence of 3 to 65% in asymptomatic persons has been reported (111). Under the influence of various predisposing factors, the prevalence may increase (111, 129). The most commonly cultured *Candida* species is *C. albicans*, but *C. glabrata*, *C. tropicalis*, *C. parapsilosis*, and *C. krusei* may also be found.

**The Candida yeasts as pathogens.** Infections with *Candida* occur not only due to the presence of individual or environmental predisposing factors but also due to the virulence of the infecting organism (111, 129). Important predisposing factors are human immunodeficiency virus infection, other immunosuppressive disorders, diabetes mellitus, age, pregnancy, hormonal dysfunction, contraceptive pills, and the use of drugs, e.g., oral antibiotics, oral corticosteroids, and immunosuppressive drugs. The presence of skin diseases often predisposes to skin infections with *Candida*.

Theoretically, *Candida* may be involved in diseases due to infections or to allergic reactions (4, 111, 129). In the host defense against *Candida*, both nonspecific immune mechanisms (e.g., an intact skin barrier or mucous membrane, presence of other microorganisms, polymorphonuclear leukocytes, macrophages, complement, and NK- and/or CD16-positive cells) and specific immune mechanisms (e.g., serum antibodies and cell-mediated immunity) are involved (129). Chronic mucocutaneous candidiasis and systemic candidiasis are both associated with defects in the cell-mediated immune response (111, 129). Virulence factors in *Candida* that are important for an infection are, e.g., adhesion to epithelial or mucosal cells, proteinases, hypha formation, propensity to switch phenotype, and surface hydrophobicity.

Infections by various members of the genus *Candida* have been reported from almost all anatomical sites (47, 111, 112); they can be divided into superficial and deep infections. It is not within the scope of this article to give a detailed description of *Candida* infections. The most commonly seen skin infections are intertrigo, cheilitis, diaper rash, balanitis, paronychia, nail infections (most commonly secondary to paronychia), and folliculitis. Superficial infections on mucous membranes include vaginitis, balanitis (which may involve both skin and mucous membranes), stomatitis, glossitis, and other superficial infections of the gastrointestinal tract. In deep candidiasis, almost all organs may be involved. Many times it is impossible to make a distinction between a primary systemic *Candida* infection and disseminated candidiasis.

*C. albicans* contains several antigenic components capable of stimulating immediate hypersensitivity responses. However, although *Candida*, and especially *C. albicans*, is often mentioned in relation to various allergic reactions, the majority of these reports are anecdotal and not reports of evidence-based studies. In patients with cutaneous *Candida* infections, sterile eczematous lesions may occur in other areas of the skin (111). This type of reaction, often called “id” reaction, is also seen in dermatophyte infections, especially tinea pedis due to *Trichophyton mentagrophytes* (var. interdigitale). Although not evidence based, this type of reaction is called an allergic reaction to fungal material because it disappears when the fungal skin infection heals. The role of an allergic type I reaction to *C. albicans* has also been mentioned for other skin diseases, e.g., chronic urticaria and psoriasis.

Other reports, also of anecdotal origin, in the literature claim that *Candida* allergy may be a major cause of manifold feelings of lack of well-being, e.g., anxiety, depression, diarrhea, premenstrual tension, poor intellectual function, and many more (16, 111). These reactions should be due to the presence of *Candida* in the gastrointestinal tract or in the blood. These reports are not evidence based, but they may be harmful because they often recommend antimycotic therapy. Non-evidence-based antimycotic therapy should of course be avoided due to the risk of allergic drug reactions, drug interactions, and the risk of development of fungal drug resistance.

**Atopic Dermatitis and the Candida yeasts**

**Distribution of Candida.** There is little information about the colonization of the skin in patients with AD. The available literature describes that *Candida* species, and especially *C. albicans*, have been cultured more frequently from both normal and lesional skin in patients with AD than from healthy individuals (4, 111). However, there are no reports about any correlation between the presence of *C. albicans* on the skin, the activity of AD, and/or the effects of antifungal therapy. *Candida* has been cultured more frequently from the gastrointestinal tract in patients with AD than in healthy controls (4, 111, 129, 132). In a study by Buslau et al., *Candida* species were cultured more frequently from feces of patients with AD and those with psoriasis (24); *C. albicans* was cultured in 90% of the cases where *Candida* species were found. The authors cultured yeasts in 70% of patients with AD compared to 54% of healthy controls (24). In a paper by Savolainen et al., *C. albicans* was cultured from the nasopharynx in 31 of 52 patients (59%) with AD (132). Skin cultures for fungi were negative. A correlation between the severity of AD and the levels of specific IgE antibodies to *C. albicans* was seen only in AD patients with positive nasopharyngeal cultures.

**Immunology.** (i) Skin prick test, with *C. albicans* and other yeasts, for type I hypersensitivity. In a study of 117 adult patients with AD, 28 patients with allergic rhinitis, and 39 healthy controls, 52% of patients with AD were SPT positive to a *C. albicans* extract compared to 22% of patients with allergic rhinitis and 5% of healthy controls (132). Positive SPTs were more commonly seen in patients with severe AD. Positive SPTs to *S. cerevisiae, Malassezia*, and molds showed a specific association with the presence of specific *C. albicans* IgE antibodies.

In a study from Japan, approximately 30% of patients with AD were SPT positive to a *C. albicans* extract and approximately 40% of AD patients were SPT positive to a *Malassezia*
extract (147). There was no difference between AD patients with or without simultaneous atopic respiratory disease. However, only approximately 10% of healthy controls were SPT positive. There was a positive correlation between positive results in SPT and specific RAST for both *C. albicans* and *Malassezia*. In another study by the same group, AD patients who showed larger SPT responses to a *C. albicans* extract tended to display lower lymphocyte proliferative responses (148). In a study by Morita et al. (106), the role of *C. albicans* in AD was evaluated and various immunological parameters were compared in patients with AD (37), patients with allergic rhinitis (23), and nonatopic subjects (42). The SPT response was significantly larger in the AD group than in the non-AD group.

Nissen et al. prick tested 15 patients with the head and neck distribution of AD or secondary infected AD with both culture filtrates and cellular extracts of *C. albicans* and *Malassezia* (109). Culture filtrates contain metabolites and cell wall components, whereas cellular extracts contain predominantly cell walls and probably fewer metabolites. Of 15 patients, 4 (27%) were SPT positive to *C. albicans* cellular extract and 2 or 13% were SPT positive to the culture filtrate. Of 15 patients, 9 (60%) were SPT positive to a commercially available *Malassezia* extract. The authors also included the leukocyte histamine release test in this study. The results were more or less comparable to the SPT results.

Kortekangas-Savolainen et al. have studied the SPT results to an extract of *S. cerevisiae* in 226 patients with AD, 50 patients with allergic rhinitis and/or asthma, and 173 nonatopic controls (81). A positive SPT result was seen in 94% of patients with severe AD, 76% of patients with moderate AD, and 25% of patients with mild AD. Patients with allergic rhinitis and/or asthma and nonatopic controls showed positive results in 8 and 2% of tests, respectively. A significant correlation was found between total serum IgE levels and SPT results. There was also a parallel between SPT result with *Malassezia* and *C. albicans*, indicating a cross-reactivity.

Positive SPT results to *Candida* were found in 22 to 94% of AD patients who were lower in healthy controls. The largest number of positive reactions was seen in patients with severe AD. Previous studies have found positive SPT reactions to *C. albicans* in 52% (132), 25 to 94% (81), 30% (147), and 27% (109) of patients with AD.

(ii) Specific IgE antibodies to *C. albicans* and other yeasts. One of the first reports of production of specific IgE antibodies against *C. albicans* was presented by Okudaira et al. in 1983 (113). They found the production of specific *C. albicans* IgE antibodies in patients with AD and/or asthma but not in controls.

In the above-mentioned paper by Savolainen et al., the severity of AD was associated with the production of *C. albicans*-specific IgE antibodies (132). In patients with the head and neck distribution of AD, there was a significant correlation between specific IgE antibodies and severity of AD only in patients with gastrointestinal growth of *C. albicans*. This pattern was also the same for AD in general but not for AD located only to the body or extremities. The specific summarized IgE staining responses to all allergens (both mannan and proteins) in immunoblot analyses, as well as the total serum IgE levels, were highest in patients with severe AD.

In a study by Tanaka et al., the production of specific IgE antibodies against *C. albicans* and *Malassezia* was significantly higher in patients with AD than in healthy controls, and the levels were also higher in patients with AD (49 patients) alone than in patients with both AD and allergic respiratory disease (48 patients) (147). In another study, Tanaka et al. found that AD patients with higher total IgE levels in serum as well as higher specific *C. albicans* IgE antibody levels tended to display lower lymphocyte proliferative responses than did patients with lower levels, although the correlation was not statistically significant (148).

Back et al. found specific IgE antibodies to *C. albicans* in 21 of 69 patients with AD (33%) but none in patients with seborrheic dermatitis or in healthy controls (8). The correlation between specific IgE antibodies to *C. albicans* and *Malassezia* was strong: 20 of 28 patients with specific *Malassezia* IgE antibodies had IgE antibodies to *C. albicans*, and 20 of 21 patients with specific *C. albicans* IgE antibodies also had IgE antibodies to *Malassezia*. There was also a correlation between the total IgE level and the levels of specific IgE antibodies to both *C. albicans* and *Malassezia*.

Matsumura et al. studied total IgE and production of specific IgE antibodies to *C. albicans* in 46 patients with AD, 41 patients with allergic respiratory disease, and 42 healthy controls (102). Both patients with AD and those with allergic respiratory disease showed a significantly higher total IgE than did controls. However, specific *C. albicans* IgE antibody levels were significantly higher in patients with AD than in patients with allergic respiratory disease. Patients with AD also reacted to a larger number of allergens than did those with allergic respiratory disease.

Doekes et al. found a highly significant correlation between levels of anti-*Malassezia* IgE and IgE reacting with extracts of *C. albicans* (30). Of 128 sera from AD patients, 34 sera reacted positively with both yeast extracts, 38 reacted with *Malassezia* but not with *C. albicans* extract, and only 1 of the 56 anti-*Malassezia*-negative sera showed a very weak reaction with *C. albicans*. The correlation was due to a marked cross-reactivity, as shown by inhibition enzyme-linked immunosorbent assay. However, *Malassezia* allergens were more potent inhibitors than were the corresponding *C. albicans* components, suggesting that the IgE antibodies in AD result from sensitization to *Malassezia* and then cross-react with *C. albicans*.

In a report by Nermes et al., a RAST test for determination of IgE antibodies to *C. albicans* mannan was described (108). Of 78 patients with AD, 53 (68%) showed elevated levels compared to healthy controls, and there was a significant correlation with the severity of AD. Of 30 patients with asthma 16 (53%) had elevated IgE mannan *C. albicans* antibody levels in serum. However, 12 of these 16 patients with a positive result also had AD. Of 32 patients with allergic rhinitis but without AD, 12 (38%) had elevated mannan IgE antibody titers. This RAST method is a sensitive assay for determination of polysaccharide-specific IgE *C. albicans* antibodies in patients with AD.

As part of a more extensive immunological study of patients with AD, Kawamura et al. measured total IgE and specific *C. albicans* IgE antibody levels in serum in 15 adult patients with AD (73). All patients had elevated total IgE levels, and 12 of 14 patients showed elevated specific *C. albicans* IgE antibody levels.
Savolainen et al. studied the presence of IgE-, IgA-, and IgG-specific yeast antibodies (total) and yeast mannan-specific IgE, IgA, and IgG antibodies in patients with atopic diseases (133). Of 20 patients, 14 were C. albicans SPT positive, 1 was C. utilis SPT positive, and 1 was Rhodotorula rubra SPT positive. A total of 12 patients with AD and 2 with rhinitis had specific C. albicans IgE antibodies in immunoblot analyses, and 10 (9 with AD and 1 with rhinitis) of 14 tested patients had C. albicans-specific mannan IgE antibodies. Nine patients had specific IgE antibodies to C. utilis, 8 patients had specific IgE antibodies to R. rubra, 4 patients had specific IgE antibodies to S. cerevisiae, and 4 patients had specific IgE antibodies to Cryptococcus albidos. IgA-specific antibodies were detected in 25 to 85% of the patients; the highest level was for C. albicans, and the lowest level was for R. rubra. IgA mannan-specific antibodies were detected in 18 to 100%; the highest level was for C. albicans and the lowest level was for Cryptococcus albidos. IgG-specific antibodies were detected in 20 to 95%; the highest level was for S. cerevisiae, and the lowest level was for C. utilis. IgG mannan-specific antibodies were detected in 74 to 100%; the highest level was for R. rubra, and the lowest level was for S. cerevisiae. Cross-reactivity between C. albicans and the other yeasts may probably explain several of the positive reactions to yeasts other than C. albicans.

In a study by Nissen et al., IgE-specific antibodies to C. albicans and Malassezia were studied using immunoblotting (109). Of AD patients, 93% had specific Malassezia IgE antibodies, 47% had specific IgE antibodies to a C. albicans cellu-
lar extract, and 60% had specific IgE antibodies to a culture filtrate extract of C. albicans. Savolainen et al. studied specific IgE and IgG antibodies to a protein and mannan antigen from C. albicans in sera from 10 patients with AD and 7 healthy controls (134). Levels of both mannan- and protein-specific IgE antibodies were elevated in AD patients compared to controls. The anti-mannan IgG responses were higher than the anti-protein IgG responses. However, no significant differences between AD patients and controls were seen. Morita et al. found that the total IgE level was significantly higher in patients with AD (37 patients) than in both patients with allergic rhinitis (23 patients) and healthy controls (42 subjects) (106). C. albicans-specific IgE was found in 85% of AD patients compared to only 9% of patients with allergic rhinitis and 5% of healthy controls.

Scalabrin et al. investigated specific IgE responses to various fungi and house dust mites in 73 patients with AD (including 16 children), 156 patients (54 children) with asthma, and 212 non-asthmatic patients (102 children) (136). Patients with AD had significantly higher total IgE levels than did those in the other two groups. Specific IgE antibodies to both C. albicans and Malassezia were detected in 75 and 54%, respectively. However, for Malassezia, significantly larger numbers of adult patients (65%) than of children (13%) had specific IgE serum antibodies. There was also a statistically significant correlation between the total serum IgE level and the level of specific IgE to both C. albicans and Malassezia as well as between specific C. albicans and Malassezia IgE antibodies, indicating a cross-
reaction. Adachi et al. found that the percentage of AD patients with specific C. albicans IgE antibodies was significantly higher for patients with severe symptoms and high total serum IgE levels (2). This was also found in patients with specific Malassezia IgE antibodies. Patients with the head and neck distribution of AD had higher specific IgE titers to both C. albicans and Malassezia. Kimura et al. found high specific C. albicans IgE antibody levels in patients with AD (78); specific IgE antibodies were found in 87% of patients with AD compared to only 15% of patients with asthma.

In a study by Broberg et al. of children with AD, specific C. albicans IgE antibodies were found in only 16% (9 of 57) (19). Lindgren et al. measured the levels of total IgE and specific IgE antibodies to a variety of external allergens in 119 children with AD (97). Of these children, 47% had specific C. albicans IgE antibodies and 34% had specific Malassezia IgE antibodies. RAST positivity to all allergens increased with age. However, this was most pronounced for Malassezia, where 42% of 10-year-old children had specific Malassezia IgE antibodies. The RAST-positive children generally had worse AD than did the RAST-negative children. Children who were RAST positive to C. albicans and/or Malassezia not only had a more severe AD than did RAST-negative children but also more often had AD with a head and neck distribution.

The frequency of specific serum Candida IgE varied from study to study. In general, the number of patients with specific antibodies was larger for adults than for children, for patients with severe AD than for those with mild AD, and for patients with a high total IgE level. Another explanation for the variation in the results obtained in various studies is the difference in the antigen preparation. Table 3 summarizes the specific IgE antibody responses found in the literature.

Nermes et al. have studied the antibody response to C. albicans mannan in children younger than 5 years with and without AD (108). There was no difference in the production of C. albicans IgG- and IgA-specific antibodies between the two groups. IgM anti-mannan levels were higher in 5-year-old children with AD. The authors concluded that children are not more susceptible to bacterial infections on the basis of poorer ability to produce antibodies to polysaccharide antigens.

(iii) Characterization of antigens. Several of the papers dealing with characterization of C. albicans antigens have already been mentioned in the section on Malassezia antigens due to the cross-reactivity between these two yeasts.

Ishiguro et al. have identified various antigens from C. albicans that reacted with IgE antibodies from sera of 57 patients with allergic diseases (64). The 175-, 125-, 46-, 43-, and 37-kDa

<table>
<thead>
<tr>
<th>Study</th>
<th>% of AD patients with specific IgE</th>
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<tbody>
<tr>
<td>Savolainen et al., 1993 (132)</td>
<td>60</td>
</tr>
<tr>
<td>Doekes et al., 1993 (30)</td>
<td>28</td>
</tr>
<tr>
<td>Nermes et al., 1994 (108)</td>
<td>68</td>
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<td>Tanaka et al., 1994 (147)</td>
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<td>Matsumera et al., 1997 (103)</td>
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</tr>
<tr>
<td>Kawamura et al., 1998 (73)</td>
<td>88</td>
</tr>
<tr>
<td>Savolainen et al., 1998 (133)</td>
<td>80</td>
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<tr>
<td>Nissen et al., 1998 (109)</td>
<td>47</td>
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<tr>
<td>Morita et al., 1999 (106)</td>
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<tr>
<td>Scalabrin et al., 1999 (135)</td>
<td>75</td>
</tr>
<tr>
<td>Adachi et al., 1999 (2)</td>
<td>87</td>
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antigenic components reacted most frequently. The 46-, 43-, and 37-kDa antigens were recovered in cytoplasmic fractions and had homology to the S. cerevisiae glycolytic enzymes enolase, phosphoglycerate kinase, and aldolase, respectively. The 175- and 125-kDa antigens were not recovered in any fraction. This result suggests that the levels of IgE antibodies against only small parts of their epitopes are elevated in the allergic patients and that, e.g., enolase may be a major immunodominant protein in both allergies and fungal infections.

Savolainen et al. found the most dominant protein allergens from C. albicans to be 27- and 46-kDa proteins (132). However, these allergens changed when the same patients were studied 5 years later and the IgE responses were found to be directed mostly against allergen bands with molecular masses of 34, 29, 20, and 13 kDa. IgA and IgG responses were directed mainly against the polysaccharide mannan.

Kortekangas-Savolainen et al. have tried to characterize the IgE binding components of S. cerevisiae (81). The found that the most frequent staining was found with a 48-kDa band. When the staining pattern of the 48-kDa band, a 45-kDa band, and mannan was compared with C. albicans allergens or purified S. cerevisiae enolase, a simultaneous binding was seen with the 48-kDa band of S. cerevisiae and the 46-kDa band of C. albicans and enolase whereas the 45-kDa band was not associated with the 46-kDa band of C. albicans or the purified enolase. Nermes et al. analyzed cross-reactivity between C. albicans and S. cerevisiae mannan (108). IgE binding to all three studied mannans from both yeasts was seen simultaneously. However, the strongest response was seen to C. albicans mannan. These results indicate that in AD patients, simultaneous IgE responses to various yeast polysaccharides occur, that the major sensitizer is C. albicans, and that the S. cerevisiae mannan is cross-reacting.

Savolainen et al. studied the major protein profiles of extracts from C. albicans, C. utilis, R. rubra, and Cryptococcus albidus (133). The greatest similarity in the protein profiles was seen between C. albicans and C. utilis, which both showed a dominant protein staining of 46 kDa. In IgE immunoblotting, prominent IgE binding to the 46-kDa band was seen in one serum sample not only to C. albicans and C. utilis but also to S. cerevisiae and to a lesser extent to the corresponding weak bands of R. rubra and Cryptococcus albidus. The IgE immunoblotting also revealed the largest number of IgE binding bands in C. albicans followed by C. utilis, S. cerevisiae, R. rubra, and Cryptococcus albidus. When cross-reactivity was studied with immunoblotting inhibition, the C. albicans inhibition of C. utilis reached 80% inhibition. The enolase staining was almost as strongly inhibited with C. albicans as was the autoinhibition. S. cerevisiae enolase inhibited C. albicans staining to 55% and C. utilis staining to only 15%, while the same concentration gave an enolase autoinhibition of 98%. An important IgG and IgA response was seen with high-molecular-weight mannan-like components.

(iv) Cellular and cytokine responses. Several papers dealing with cellular and cytokine responses have already been mentioned in the Malassezia section. The delayed-type hypersensitivity response to C. albicans was found in several studies to be different from the result seen with Malassezia and other environmental allergens.

Using the chamber-scarification patch test technique, 97 patients with AD were investigated for delayed-type hypersensitivity against extracts from C. albicans, Malassezia, and other environmental allergens (147). Healthy controls had a higher incidence of positive patch test results (86% positive) against C. albicans than did patients with AD (49%). For the other environmental allergens tested, including Malassezia (approximately 10% positive), the incidence of positive results in patients with AD was equal to or higher than the reaction seen in healthy controls. A negative correlation between the patch test result for C. albicans and both total IgE and specific RAST to C. albicans was found in patients with AD. The authors conclude that the persistent exposure to C. albicans allergen in patients with AD leads to the development of immediate hypersensitivity, with a subsequent decrease or total loss of delayed-type hypersensitivity to C. albicans allergen. This is in parallel with the results of experimental dermatophytosis experiments in patients with AD (68). The persistent carriage of C. albicans in the oral cavity and intestine may induce a Th2-type T-cell response that facilitates the IgE response on the one hand but reduces the delayed-type hypersensitivity on the other (147).

In another study by the same group, the incidence of positive patch test results in response to a C. albicans allergen was again significantly lower in patients with AD than in healthy controls (148). The results of a lymphocyte stimulation test with a C. albicans allergen, using PBMC from AD patients, gave a significant lower stimulation index than for healthy controls. The results with Staphylococcus enterotoxin A and B were similar for AD patients and healthy controls.

Matsumura et al. performed patch test (scarification) experiments using various environmental allergens in 46 patients with AD (without any respiratory allergy), in 41 patients with allergic respiratory disease but without AD, and in 42 healthy controls (103). AD patients showed a significantly lower incidence of positive responses to C. albicans (34%) than did both patients with allergic respiratory disease (90%) and healthy controls (84%). When the patch test results were compared to specific RAST results the percentage of those with positive RAST results and negative patch test results was higher in the AD group, while the percentage of those with negative RAST results and positive patch test results was strikingly higher in the group with allergic respiratory disease.

In a study by Kawamura et al., PBMC from both patients with AD and healthy controls were incubated with a crude extract of C. albicans (73). PBMC from both AD patients and healthy controls showed a significant increase in proliferation. However, PBMC from healthy controls showed a significantly higher stimulation index than did PBMC from AD patients. When PBMC stimulated with C. albicans were costimulated with monoclonal antibodies to CD40, CD54, CD80, or CD86-positive cells, the proliferation was suppressed in a significantly larger number of subjects with anti-CD54 than with any of the other antibodies. This effect of anti-CD54 was seen in both AD patients and healthy controls.

Kimura et al. have studied the production of IFN-γ and IL-5 by PBMC after stimulation with an extract of C. albicans in patients with AD, patients with bronchial asthma and healthy controls (77). The amount of IFN-γ produced was significantly smaller in AD patients than in both patients with bronchial asthma and healthy controls. The production of IL-5 was again
In contrast to Savolainen et al. (134) they found a significant negative correlation between *C. albicans* IgE level, and specific extracts, was significantly higher in AD patients than in healthy controls (135). For *C. albicans*, IL-5 production and IFN-γ production was significantly higher in AD patients than in healthy controls. *C. albicans*-induced IFN-γ production was significantly higher for *Malassezia*. *Malassezia*-induced IL-4 production was significantly higher than for *C. albicans*. Total IgE and PHA-induced IL-2 and IL-4 levels in serum were elevated. A network of correlations was seen between the total IgE level in serum and the yeast-specific IgE level, *Malassezia*-specific proliferation, PHA-induced IL-2 and IL-4 levels, and the *C. albicans*-induced IL-5 level, suggesting that these are associated with AD. According to the authors, hypersensitivity to yeasts is an important pathogenic factor in some cases of AD. Both *C. albicans* and *Malassezia* can induce IgE, lymphoproliferation, IL-2, IL-4, IL-5, and IFN-γ responses. However, according to the authors, *Malassezia* is a stronger inducer of Th2-type response and *C. albicans* is a stronger inducer of Th1-type responses.

There are several explanations of the differences obtained in different studies. Kimura et al. studied children, and Savolainen et al. studied mainly adults. Differences in the preparation of extracts, the length of the culture period, and the methods used for the experiments are also important factors. The lymphocyte stimulation responses and cytokine responses found in the literature can be summarized as follows: Kawamura et al. (73) found increased lymphocyte stimulation but less than in healthy controls; Kimura et al. in 1996 (76) found decreased production of IL-5 and IFN-γ and in 2000 (78) found decreased production of IFN-γ; Sarolainen et al. in 1999 (134) found increased production of IL-2 and IFN-γ and in 2001 (140) found increased production of IL-5, IL-2, and IFN-γ.

**Treatment.** Several studies have already been mentioned in the section dealing with *Malassezia* yeasts and treatment of AD (8, 9, 101).

In the above-mentioned study by Morita et al., five patients with AD were treated orally once daily with 50 mg of ketoconazole and five patients with AD were treated orally once daily with 50 mg of fluconazole for 3 months (106). In both treatment groups, four patients were evaluated as improved after 3 months of treatment, and *C. albicans*-specific IgE levels in serum were significantly decreased in both groups.

In a study by Adachi et al. (2), the effect of oral antifungal treatment was evaluated in 140 patients with treatment refractory AD and with a positive specific RAST result to *C. albicans*. According to the authors, good or excellent results were obtained in 60% of patients receiving fluconazole, 35% receiving itraconazole, 31% receiving oral amphotericin B, and 28% receiving nystatin. The number in each group, the dosage, and the length of treatment are not mentioned in the English abstract of this Japanese paper. That oral amphotericin B and nystatin both work indicate that *C. albicans* present in the gastrointestinal tract may be important as an allergen in AD.

Although some studies may indicate that antifungal treatment of both *Malassezia* and *Candida* may be effective in the treatment of some patients with AD, we need further controlled studies to really prove this. Table 4 summarizes the results of treatment studies mentioned in the literature.

### TABLE 4. Treatment of AD patients with antifungal drugs

<table>
<thead>
<tr>
<th>Study</th>
<th>Treatment</th>
<th>Design</th>
<th>Results</th>
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<td>Clemensen and Hjorth</td>
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<td>Double-blind crossover</td>
<td>Ketoconazole &gt; placebo</td>
</tr>
<tr>
<td>Back et al., 1995 (8)</td>
<td>Oral ketoconazole</td>
<td>Open</td>
<td>Ketoconazole &gt; placebo</td>
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<tr>
<td>Broberg et al., 1995 (20)</td>
<td>Topical hydrocortisone + miconazole</td>
<td>Double blind</td>
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**Atopic Dermatitis, Dermatophytes, and Other Hyphomycetes**

There are conflicting data in the literature about the association of AD with chronic dermatophyte infections (68, 143).
However, if a patient with AD develops a chronic dermatophyte infection, it is often more severe and more difficult to eradicate. The first extensive report about a link between AD and chronic dermatophyte infections was presented by Jones et al. (67). They found that 14 (40%) of 25 patients with chronic dermatophyte infections were atopic (AD, asthma, hay fever, or all three). A comparison of the frequency of chronic dermatophyte infection showed that 41% (14 of 34) in the atopic group had chronic dermatophyte infections compared to only 14% (21 of 146) in the nonatopic group. They also found that there was no correlation between chronic dermatophyte infection and any clinical condition except atopy. Of the 14 AD patients with chronic dermatophyte infections, 11 (79%) manifested immediate sensitivity to Trichophyton.

However, the reason for the failure of immunity in persistent dermatophyte infections and its relationship to chronicity are still not well understood. There is an association between the presence of atopy and chronic dermatophytosis. A high proportion of those with persistent disease had atopic diseases (most commonly asthma or hay fever) as well as immediate-type hypersensitivity and raised total IgE levels (57, 68). It has been suggested that modulation of T-lymphocyte activity either locally (in the skin) or systemically may be possible. Possible mechanisms include activation of a Th2 pathway, which might explain the spectrum of antibody responses. It has also been found that dermatophyte antigens, including those that contain mannose residues, can reversibly suppress lymphocyte proliferation but not the expression of human leukocyte antigen (HLA-DK) (71, 104). Patients with persistent infection have detectable levels of circulating antigen. Both are possible factors in the regulation of immunity in dermatophytosis.

In a study by Rajka and Barlinn, patients with AD and chronic dermatophyte infections showed not only positive immediate reactions to Trichophyton but also to Penicillium and Cladosporium (117). Even patients with AD who showed no sign of dermatophyte infections had this cross-reactivity. However, this was absent in nonatopic patients with chronic dermatophyte infections, and healthy controls were negative to all the three extracts. These findings indicate that immediate reactivity to Trichophyton in AD patients is not necessarily a sign of contact with dermatophytes, but a cross-response based on mold reactivity.

Patients with dermatophytosis are usually otherwise healthy. However, chronic and extensive infections have been reported to be more common in patients with immunodeficiency (68). In addition, there is an increased incidence of atopy in those with chronic infections, suggesting that host factors may well determine the clinical course.

Dermatophytes may also have an impact on the severity of atopy, especially asthma (163). Platts-Mills et al. reported on a patient with severe asthma, tinea pedis, and tinea cruris who showed positive results of immediate skin test and RAST to Trichophyton (115). When the patient was treated with an effective antifungal drug, his asthma improved significantly. Ward et al. have reported 12 patients with allergic respiratory diseases with positive immediate reactions to Trichophyton, whose disease became worse when they were challenged with Trichophyton antigen (155). Eight of these patients improved when treated with specific systemic antifungal therapy.

Even AD patients with dermatophyte infections may show improvement of their AD when treated with specific antifungal therapy (163). Wilson et al. have described a patient with tinea unguium, recalcitrant AD, and specific IgE Trichophyton serum antibody who showed improvement of her AD when treated with oral antifungal treatment. Klein et al. have also described a patient with AD and tinea pedis and tinea unguium culture positive for T. rubrum (79). The patient showed improvement of her AD when treated with oral ketoconazole for her tinea infection.

Scalabrini et al. have studied the presence of total IgE and specific IgE to various fungi in AD patients (136). The majority of AD patients had both elevated total IgE and specific IgE antibodies directed against Aspergillus fumigatus and A. alternata (fungi that are characteristically inhaled) as well as against C. albicans, M. furfur, and T. rubrum (fungi that colonize the skin or mucous membranes). There were extensive correlations among specific IgE for each of the fungi. The authors conclude that fungi may be important allergens in several patients with AD and that antifungal therapy may improve AD in many patients.

Lindgren et al. found that 18% of children with AD had IgE antibodies to Trichophyton (97). These patients also had specific antibodies to other fungi, house dust mites, and other environmental allergens. Seven children with positive RAST results to both T. rubrum, Malassezia, and C. albicans had severe AD.

CONCLUSION

Do yeasts and other fungi play any role in AD? As shown in this review, there are conflicting data in the literature. It is important to remember that fungi are not the causative agents of AD. However, in some patients especially in adult patients who do not respond to traditional treatment, Malassezia and Candida may play a role. Although some studies indicate that antifungal treatment of both Malassezia and Candida may be effective in the treatment of some patients with AD, we need new, well-conducted, double-blind controlled studies to really prove this. It is also important to remember that a possible allergic or nonimmunogenic role of fungi is just an aggravating factor in AD and that traditional treatment with emollients and anti-inflammatory agents is still necessary. In the literature, conflicting results are presented for both immediate hypersensitivity (SPT and IgE), delayed hypersensitivity (APT), lymphocyte stimulation assays, and IL assays. There are several explanations for this: adult patients compared to children, the severity of AD, the presence of other atopic manifestations, differences in the preparation of extracts, the length of the culture period used, and the method of the assay.

REFERENCES


