The Genus *Hafnia*: from Soup to Nuts

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INTRODUCTION

The genus *Hafnia* is one of more than 40 genera that currently comprise the family *Enterobacteriaceae*. Although Møller originally described this genus in 1954 (89), the legitimacy of this group was constantly challenged over the next two decades, often being referred to by such synonyms as “*Enterobacter alvei*”, “*Enterobacter aerogenes* subsp. *hafniae*” and “*Enterobacter hafniae*” (www.bacterio.cict.fr/h/hafnia.html). Subsequent DNA-DNA relatedness investigations conducted by Don Brenner and the Centers for Disease Control and Prevention (CDC) conclusively established that these bacteria represented a unique taxon within the family and that at least two or more DNA groups existed within the genus (20, 21, 131).

Although this genus is over 50 years old, very little is known about these organisms in regard to the role(s) they may play as both human and veterinary pathogens. In one sense, these bacteria appear to be “orphan enterobacteria” in search of a medical and public health identity. *Hafnia* is fairly often isolated from clinical material, yet with the exception of its doc-
umented role as a rare cause of bacteremia (31, 104, 105), its relationship to other clinical infections is questionable. In fact, data from the early 1990s suggesting that hafniae were true enteropathogens (4, 5) now turn out to have been incorrectly attributed to hafniae rather than to the actual pathogen, *Escherichia albertii* (63, 69). The possible role of *Hafnia* in bacterial gastroenteritis is presently unknown, although one recent case report suggests a possible role in enteritis (see Addendum).

Over the past quarter of a century, there have been few studies that have systematically looked at the role of these bacteria in clinical microbiology. Greipsson and Priest (56) published the only taxonomic study on this genus in 1983, and Günthard and Pennekamp (59) reported on the clinical significance of the only large series of extraintestinal isolates. Recently there have been a number of studies describing and characterizing multiple genomospecies within the genus, phylogenetic relatedness, clinical infections, pathogenicity, and biochemical characteristics of genomovars. This review attempts to systematically review the pertinent literature on this genus, discuss current controversies surrounding these bacteria, and shed some light on the potential role these “orphan enterobacteria” may play in human disease.

**HISTORICAL PERSPECTIVES**

It is the subject of some speculation and debate as to when organisms currently in the genus *Hafnia* were first isolated. In 1919, L. Bahr worked on a bacterium that he designated “*Bacillus paratyphi-alvei,*” an organism reputedly pathogenic for bees but not mice or guinea pigs (121). One of Bahr’s apparently authentic “*Bacillus paratyphi-alvei*” strains (referred to as “*Paratyphus alvei*”) was subsequently characterized in 1954 as belonging to a new group of enteric bacteria for which Möller coined the name “*Hafnia* group” during a systematic investigation of amino acid decarboxylase patterns among members of the family *Enterobacteriaceae* (89). Some groups subsequently questioned the legitimacy of this name in light of the fact that Bahr’s strains differed in some biochemical characteristics from those described by Möller. However, Möller considered that Bahr’s strain should be regarded as the type species of *Hafnia,* and he suggested the name *Hafnia alvei.*

There are no extant cultures of Bahr’s strains. However, in the early 1940s Stuart and colleagues (135) biochemically characterized a large group of paracolon bacteria. Within the group named “*Paracolon Aerobacter,*” two divisions were recognized based upon the results of indole–methyl red–Voges-Proskauer–citrate assays. Division 2 contained seven distinct biotypes, the largest of which was referred to as type 32011 (35 cultures). Type 32011 and related types had properties generally consistent with inclusion in the genus *Hafnia,* being indole, lactose, and sucrose negative; aerogenic; and Voges-Proskauer (VP) positive. Type 32011 strains were subsequently characterized by several independent groups, including Edwards and Ewing (29), who confirmed that the biochemical characteristics of type 32011 were identical or nearly identical to those reported for the “*Hafnia* group” by Möller and others.

From the 1940s until the mid- to late 1970s, when nomenclature and classification issues surrounding this taxa were resolved, *H. alvei* was known by a number of different names, including “*Paracolon Aerobacter,*” “*Paracolobactrum aero- noides,*” *Bacillus cadaveris,* the Hafnia group, *Enterobacter alvei,* and *Enterobacter hafniae* (16, 120, 121). For one brief period of time it was even relegated to subspecies rank within *Enterobacter aerogenes* (32). However, subsequent taxonomic investigations demonstrated that it belonged to a unique lineage within the *Enterobacteriaceae* and deserved genus (species) rank (see “NOMENCLATURE, TAXONOMY, AND CLASSIFICATION” below). The name *Hafnia alvei* appears in the Approved Lists of Bacterial Names, and the type strain is ATCC 13337T (129). Three of the oldest extant cultures of *H. alvei* are ATCC 13337T (Stuart’s type 32011 = NCTC 8105 = NCD 434-68), ATCC 11604 (“*Paracolobactrum aerogenoides*” = 3565 [Conn 3565], Stuart’s type 32011) of Borman et al. (16), and NCTC 6578 of Gale and Epps (43).

**NOMENCLATURE, TAXONOMY, AND CLASSIFICATION**

**Nomenclature**

The specific epithet in the name *Hafnia alvei* is derived from the Latin noun alveus, meaning beehive, with “alvei” meaning “of a beehive.” Ewing (33) questioned the epithet “*alvei,*” stating that the name implied that these bacteria had something to do with bees or beehives although they did not. However, *H. alvei* has been recovered on occasion from the intestines of honeybees (*Apis mellifera*) as well as from honey, and several of these strains are included in the BCCM (Brussels, Belgium) collection (125). The genus name *Hafnia* is the historical name (Havn) for the city of Copenhagen, Denmark (122).

**Biochemical and Numerical Taxonomy Studies**

Prior to the advent of molecular biology, bacterial nomenclature and taxonomy were dependent primarily upon characters or phenotypes of cultures expressed under defined conditions. In their most sophisticated form, such analyses employed the principles of Adansonian taxonomy (numerical taxonomy or numerical phenetic analysis), but often studies simply compared large data sets of biochemical characteristics between two apparently different groups. In that vein, members of the family *Enterobacteriaceae* (17–19, 60, 75, 119) were grouped into tribes consisting of genera with similar characteristics. The tribe *Klebsiellae* contained four such genera (*Klebsiella, Enterobacter, Hafnia,* and *Serratia,*), with most but not all of the tribe’s members sharing a number of phenotypic features. These features included formation of acetyl methylcarbinol (VP), growth on Simmon’s citrate agar and in potassium cyanide broth, urea negativity (48 h), and failure to produce hydrogen sulfide (33). Few studies have dealt with the genus *Hafnia* by employing numerical taxonomy principles. Bascomb and coinvestigators (12) studied members of the then-tribe *Klebsiellae,* including 15 strains of *H. alvei.* By using three different methods of numerical classification, all *H. alvei* strains were found to form a separate and distinct branch with a rank similar to that of the tribe *Klebsiellae.* Johnson et al. (73) studied 384 strains in the family *Enterobacteriaceae* for 216 independent characters, including morphological, biochemical, and physiologic traits; 10 of these enteric isolates were *H. alvei.* Based upon simple matching (*S*ₐ) and Jaccard (*S*ₐ) coefficients, *H. alvei* strains were found to cluster independently of other groups, joining
members of the tribe *Klebsiellae* at 78% similarity. Those authors concluded that this justified the status of this taxon as an independent genus rather than being placed within the genus *Enterobacter*. The only study to specifically look at *Hafnia* by use of numerical taxonomic principles was that of Greipsson and Priest (56), who studied 47 strains from the United Kingdom, Japan, France, and the United States by using 101 different characters. All 47 strains exhibited high similarity values (>78%) by $S_{ae}$ coefficient when average-linkage cluster analysis was used with the type strain NCTC 8105$^T$ (ATCC 13337$^T$), located near the center of the phenon. These results support previous conclusions reached by Johnson et al. (73) that all *H. alvei* isolates reside within a single phenon or cluster.

**DNA Relatedness**

Despite the general conclusions drawn from several phenotypic studies regarding the homogeneity of *Hafnia* isolates, other studies provided indirect evidence that genetic diversity exists within this nosemespieces. A study by Gavini and colleagues (45) identified two distinct *Hafnia* phenons, designated I$_1$ and I$_2$, in their analysis of 32 strains originating from feces and different water sources. These two clusters could not be unambiguously separated from one another, although they differed in the frequency of certain characters, including production of acetylmethylcarbinol at 37°C and utilization of maltose and D-tartrate. Another clue suggesting diversity within *H. alvei* involves the moles percent G+C content of hafniae (66). Two very different G+C contents for *Hafnia* have been reported; the more common estimate is 48.7 mol%, while a higher value of 52.6 to 57.7 mol% has also been published (56).

The first direct evidence proving the existence of more than one DNA group within *Hafnia alvei* was reported by Steigerwalt and coinvestigators (131) as part of a study of DNA relatedness between *Enterobacter* and *Serratia* species. In that study, using *H. alvei* ("E. hafniae") CDC 4360-67 as the source of labeled DNA, two distinct DNA groups could be discerned, one highly (85% to 100%) related to CDC 4360-87 and one less (51% to 55%) related; the latter contained the type strain of *H. alvei*, ATCC 13337$^T$. *H. alvei* was found to be only 16% to 24% related to other members of the *Enterobacteriaceae* by DNA-DNA hybridization in that study. In subsequent publications, Brenner (20, 21) reported the existence of two or more hybridization groups (HGs) within *H. alvei*, and this has been confirmed in subsequent studies (47). Furthermore, Brenner (21) found that the type strain of *Ohesambacterium proteus* (ATCC 12841$^T$) is 75% related to DNA group 2 (in some studies called DNA group 1) of *H. alvei* and is 52% related to the type strain of *H. alvei*, ATCC 13337$^T$. Thus, a valid type strain for *O. proteus* does not exist. Table 1 lists reference strains and their vernacular designations from surveys that studied both *H. alvei* DNA groups (HGs). By convention, the type strain for the nosemespieces (ATCC 13337$^T$) and related reference strains should be classified as DNA group 1 (HG1), since this group represents *H. alvei* sensu stricto. DNA group 2 (HG2, unnamed) is represented by several reference strains, including ATCC 29927. CDC 4360-67 (DNA group 2) was not retained in the permanent culture collection of CDC and is no longer available as a reference strain for DNA group 2 (HG2) (C. O’Hara, personal communication).

Other recent studies employing different techniques also support the division of the genus *Hafnia* into two or more groups. 16S rRNA sequencing of 22 *H. alvei* isolates revealed two clusters, one containing 5 strains (including ATCC 29926) that exhibited 0% to 0.19% sequence divergence from ATCC 13337$^T$ (DNA group 1) and a second cluster (DNA group 2) of 17 strains that exhibited 0% to 0.09% sequence divergence from ATCC 29927 (0.95% to 1.04% divergence from ATCC 13337$^T$) (71). A subsequent study of 52 *Hafnia* strains by the same group again found two DNA groups that were clearly distinguishable using 16S rRNA gene sequencing (70). An Australian study in 2003 investigated the genetic structures of 161 *Hafnia* isolates by multilocus enzyme electrophoresis at 12 different loci (98). As in the previous study, two populations were detected, one consisting of 83 strains (including ATCC 13337$^T$ and ATCC 29926) and a second group of 81 isolates (including ATCC 29927). These studies indicate that 16S rRNA sequencing and multilocus enzyme electrophoresis also detect HGs that were previously recognizable only through DNA pairing studies.

**Phylogenetic and Related Studies**

Only a limited number of phylogenetic investigations have included one or more strains of *H. alvei*. The utility of 16S rRNA sequencing to study the family *Enterobacteriaceae* has been questioned due to the high intrafamily DNA relatedness values and limitations on the resolution of 16S rRNA for this group.

<table>
<thead>
<tr>
<th>Reference</th>
<th>No. of groups detected</th>
<th>Proposed DNA group nomenclature for <em>H. alvei</em></th>
</tr>
</thead>
<tbody>
<tr>
<td>131</td>
<td>3$^e$</td>
<td>DNA group 2 (ATCC 13337 = NCTC 8105)</td>
</tr>
<tr>
<td>20</td>
<td>3$^e$</td>
<td>DNA group 1 (ATCC 5632-72 = ATCC 29926)</td>
</tr>
<tr>
<td>21</td>
<td>3$^d$</td>
<td>NG$^a$</td>
</tr>
<tr>
<td>33</td>
<td>2</td>
<td>DNA group 2 (ATCC 13337)</td>
</tr>
<tr>
<td>47</td>
<td>2</td>
<td>HG1 (CDC 5632-72)</td>
</tr>
</tbody>
</table>

$^a$ Based upon DNA relatedness studies.

$^b$ Based upon ATCC 1337$^T$ representing DNA group 1.

$^c$ DNA group 3, represented by strain CDC 329-73, was subsequently determined to be *Yokenella regensburgei* (20).

$^d$ HG3 was resistant to *Hafnia*-specific bacteriophage and was more closely related to *Salmonella-Escherichia-Citrobacter-Enterobacter* than to *Hafnia*.
based upon the number of informative sites (27, 130). Spörrer et al. (130) found that the type strain of *H. alvei* exhibited 99.5% sequence similarity to the type strain for *O. proteus* (ATCC 12841T), supporting previous observations by Brenner (21) regarding the DNA relatedness of these two species. In that study, the nearest neighbors to *H. alvei* were two distinct *Serratia* clusters containing nine species. Dauga (27) also looked at the phylogenetic position of *H. alvei* ATCC 13337T relative to other *Enterobacteriaceae*. By using the neighbor-joining method, *H. alvei* was rooted by the tribe *Proteaceae* (containing the genera *Proteus*, *Providencia*, and *Morganella*); by maximum-likelihood analysis, *H. alvei* clustered with *Serratia*, while by the distance method its nearest neighbors were the *Escherichia-Salmonella-Citrobacter* group. A 2003 study employing only a few enteric genera found the closest relative to *H. alvei* to be *Klebsiella pneumoniae* based upon 16S rRNA gene sequencing (139).

Hedegaard and colleagues (62) investigated the utility of partial sequencing of the *infB* gene (encoding translation initiation factor 2) as a phylogenetic tool for studying 39 *Enterobacteriaceae* strains. By using a clinical strain of *H. alvei* and the distance matrix method, the closest neighbors to hafniae were found to be members of the tribe *Proteaceae*. Phylogenetic investigations using *gyrB* gene (encoding the ATPase domain of DNA gyrase) and protein sequence data yielded mixed results (27). Dauga (27) noted that there was atypical codon usage (third-codon G+C content) in the *gyrB* gene of *H. alvei* relative to other *Enterobacteriaceae*, which suggested foreign acquisition of this gene. If this is true, analysis of the *gyrB* sequences would be indicative only of the origin of the gene and not the species. Wertz et al. (139) studied five housekeeping gene and 16S rRNA gene sequences in an analysis of a limited series of species in the *Enterobacteriaceae*. Cladograms generated by maximum-likelihood analysis found that *H. alvei*’s nearest neighbor was *Serratia plymuthica* for four genes (*groEL*, *gyrA*, *pgi*, and *ompA*), while by *gapA* sequencing it was *Enterobacter cloacae* and by 16S rRNA analysis *K. pneumoniae*. Phylogenetic data to date are inconclusive regarding where the genus belongs. One study on the evolution of aromatic amino acid biosynthesis genes and enzymes suggests clustering of *H. alvei* with *Edwardsiella*, *Yersinia enterocolitica*, and *Proteus vulgaris* (2). However, to date phylogenetic data regarding where the genus should be rooted are inconclusive.

**EPIDEMIOLOGY**

**Environmental Distribution**

Most standard microbiology texts list mammals, birds, reptiles, fish, soil, water, sewage, and foods as sources from which hafniae can be recovered (35, 36). However, there are surprisingly little hard data in the literature on this topic, and no systematic environmental surveys on the genus *Hafnia* have been published. Most of the data regarding the environmental distribution of hafniae come from investigations on the frequency, incidence, concentration, and distribution of members of the family *Enterobacteriaceae*. Host and geographical factors appeared to influence the thermal niche of *H. alvei* isolated from Australian mammals in one study (97).

The gastrointestinal tracts of animals, and in particular mammals, appear to be a very common ecologic habitat for hafniae. Paleomicrobiology investigations have identified *H. alvei* originating from intestinal mass samples and sediment collected from 12,000-year-old mastodon remains in Michigan and Ohio (110). In a study of 642 Australian mammals, Gordon and FitzGibbon (53) found that *H. alvei* was the third most common enteric species identified, following *Escherichia coli* and *E. cloacae*. The isolation of *H. alvei* was significantly associated with recovery from marsupial carnivores and murid rodents. Hafniae have also been recovered sporadically from pack animal manure samples collected from national park trails (28) and from 7% of grizzly and black bear specimens tested (51). Among avian species, *H. alvei* has been frequently isolated from birds of prey, including falcons, owls, and turkey vultures; even high-altitude alpine accentors that have virtually no contact with humans have had *Hafnia* isolated from them, at frequencies ranging from 3% to 16% (9, 78, 142). Other sources for *H. alvei* include reptiles (snakes and skinks), invertebrates, insects, fish, and bats (24, 52, 98).

Food products commonly yield hafniae. Meats, including minced and vacuum-packed beef and pork products, ground beef chubs, and retail packages, harbor *H. alvei* (82, 113). One Finnish study found almost 50% of all enteric isolates recovered from refrigerated meats to be *H. alvei* (113). A subsequent study identified 34% of minced meat samples, 14% of milk and cream products, and 12% of freshwater fish as containing hafniae (44). Counts of *H. alvei* in these samples were as high as 7.5 × 10⁶ to 8.0 × 10⁷ CFU/g (82). Some isolates of *H. alvei* recovered from albacore produced histamine, although the relevance of this finding to scrombroid poisoning is unknown (76). Other consumables that are reported to yield *H. alvei* include cheese and honey (90). Vegetables do not appear to be a frequent reservoir for these bacteria, although *H. alvei* was recovered from nine Finnish and imported vegetable samples in one study (100). There are only a few anecdotal reports describing the isolation of hafniae from water (98).

**Disease States Associated with *H. alvei***

There are only a limited amount of data presently available on disease states associated with *H. alvei*. There are two well-described outbreaks of *H. alvei* infection associated with the poultry industry. Real et al. (108) described an outbreak of disease in commercial laying hens in Spain, characterized by loss of appetite, decreased egg productivity, catarhral enteritis, and fulminating septicaemia. Prominent histopathologic features of the disease included hepatosplenomegaly, multifocal necrotizing hepatitis, and splenitis. *H. alvei* was recovered in pure culture from infected tissues at necropsy. A very similar disease presentation could be reproduced using wild-type *H. alvei* isolated from ill hens experimentally infected by either the intraperitoneal or oral route. In 2004, a large outbreak of *H. alvei* disease in 12-week-old pullets was reported from Italy (103). These pullets initially suffered from anorexia, depression, ruffled feathers, and diarrhea. The overall mortality rate was 20.7%. Autopsy findings were remarkably similar to those observed by Real and colleagues (108), including catarhral enteritis and hepatosplenomegaly. Again, the disease could be
experimentally reproduced in healthy pullets challenged with the infective strain.

*H. alvei* has been implicated in a variety of other animal infections. *H. alvei* has been reported as the cause of an epizootic outbreak of septicemia in rainbow trout in Bulgaria (47). *Hafnia* was recovered in pure culture from parenchymal organs during the acute phase of disease and was also isolated from necrotic tissues later in the course of infection. A 1963 French publication describes an epizootic outbreak of disease in white laboratory rats 3 to 8 weeks old (8). A number of the rats were lethargic and died unexpectedly. Autopsy demonstrated splenic hypertrophy (four to five times normal size) with congestion in visceral organs, including the brain and cerebellum. *H. alvei* was recovered from samples of organ tissues and heart blood. This strain possessed phenotypic properties that are consistent with inclusion in the genus, except that these were VP negative (≥85% of *H. alvei* isolates are positive) and produced acid from dulcitol (<1% of *H. alvei* isolates are positive). Other animal diseases linked to *Hafnia* infection include caprine pneumonia (127) and equine abortion (91).

**Ecologic Structure**

The first evidence suggesting that there may be a nonrandom association of distinct genetic populations of *H. alvei* with specific ecologic niches comes from a recent study by Okada and Gordon (98). In that investigation, genetic group 2 strains (corresponding to DNA group 2 in Table 1) were predominately associated with freshwater fish, while strains from genetic group 1 (corresponding to DNA group 1 in Table 1) were associated with birds, frogs, invertebrates, and water isolates. Both groups were equally represented in mammals and reptiles.

**Clinical Frequency, Outbreaks, and Nosocomial Infections**

*H. alvei* is an uncommon human pathogen, although this species has received increased attention from the medical community over the past decade due to its possible association with gastroenteritis (Fig. 1) (see “CLINICAL SIGNIFICANCE” below). Hafniae are commonly recovered from oropharyngeal specimens, from the gastrointestinal tract, less often from blood, and from a variety of other anatomic sites, including tissue, urine, and catheters (59, 138). In a study of 17 *Hafnia*
alvei ("Enterobacter hafniae") isolates recovered by the Mayo Clinic from 1968 to 1970, only 5 isolates (29%) were judged to be clinically significant (138). In all five of these cases, *H. alvei* was determined to be a secondary pathogen (respiratory, *n* = 2; abscess, *n* = 3). Overall, the mean age of persons infected/colonized with *H. alvei* was 52.9 years, with a male/female ratio of 1:1.1. Slightly over 50% of all isolates were acquired via a nosocomial route, including all five associated with infection (138). In only two instances was *H. alvei* recovered in pure culture, and in neither case was the strain clinically significant. A 1996 study by Günthard and Pennekamp (59) reviewed 80 *H. alvei* isolates isolated from 61 patients over a 30-month period (1992 to 1995). As in the previous report, the mean age of all patients was 53.5 years; however, almost two-thirds of the patients were male. In only 6 of these 61 patients (9.8%) was *H. alvei* thought to be the etiologic agent of the disease, three of which were proven cases (septicemia, *n* = 2; peritonitis, *n* = 1) and three of which were probable cases (septicemia, *n* = 2; pneumonia, *n* = 1).

There are no well-described outbreaks of disease caused by *H. alvei* in which the epidemiologic, clinical, and laboratory correlates are overwhelmingly in support of an unquestionable role for hafniae in these illnesses. Table 2 lists five outbreaks of gastrointestinal disease, affecting between 2 and 72 individuals, in which *H. alvei* was suspected as the possible etiologic agent. Two Canadian outbreaks of gastroenteritis that occurred 1 month apart in the same institution in inpatients suggest a possible role for hafniae in the disease process. However, only 32% of the patients had *H. alvei* recovered in large numbers from their stools, and one symptomatic person continued to grow large numbers of hafniae from feces even after the person became asymptomatic (107). A report from Russia describes the possible association of *H. alvei* with diarrhea in 72 hospitalized persons and provides some microbiologic and immunologic data to support this hypothesis; however, a number of questions remain (54). These include a short incubation period, unusual symptoms (possibly neurotoxicosis), and the presence of rotavirus in at least some of these samples (54). Clearly, more definitive work in this area needs to be performed.

Since *H. alvei* is presently an uncommon human pathogen, incidence data such as those derived from the National Nosocomial Infection Surveillance System for many enteric pathogens are not available (112). From three separate studies, the percentage of documented *H. alvei* nosocomial infections ranged from 38% to 59%, although the absolute number of cases was relatively small (8 to 80) and they spanned a number of years (59, 105, 138).

### CLINICAL SIGNIFICANCE

**Historical Anecdotes Regarding the Association of Hafniae with Human Disease**

There are numerous narrative comments in the medical and scientific literature regarding the potential pathogenicity of *H. alvei*. In most instances these comments center on presumed associations between this bacterium and diarrheal disease. In 1943, Stuart and Rustigian (134) reported on one outbreak of mild to acute gastroenteritis that involved a fairly sizable number of persons in Rhode Island, with the common thread of consumption of milk or milk-related products from the same supplier. The implicated organism was biotype 32011 ("*H. alvei*") (Table 2). In the same study, the authors also described two apparent laboratory-acquired gastrointestinal infections, in a research student and in a glassware technician, both of whom handled biotype 32011. Other reports similar to this have surfaced from time to time, such as the case reports of Harada et al. (61) in 1957 describing the isolation of *H. alvei* from two siblings, who developed diarrhea 3 weeks apart. However, most of these data are extremely sketchy, and Matsumoto (85) in 1963 stated that the pathogenicity of *Hafnia*, at least for humans, was not well clarified.

The first credible report linking *H. alvei* to human illness came from a 1967 case study by Jenss and McCarthy (72). A 58-year-old man who underwent multiple medical procedures for a duodenal ulcer developed *Hafnia* bacteremia with fever (40.5°C) and hypotension (80/50) 6 days after surgery. Two blood culture specimens were positive for *H. alvei*, and he was

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**TABLE 2. Reputed outbreaks of disease caused by *H. alvei***

<table>
<thead>
<tr>
<th>Outbreak no. (reference)</th>
<th>Yr</th>
<th>Location</th>
<th>No. ill</th>
<th>Symptoms</th>
<th>Persons affected</th>
<th>Comments</th>
</tr>
</thead>
<tbody>
<tr>
<td>1 (134)</td>
<td>1943</td>
<td>United States</td>
<td>28</td>
<td>GE</td>
<td>Inpatients and staff</td>
<td>Biotype 32011 (&quot;<em>H. alvei</em>&quot;) isolated from a patient, cream-filled pastry, and milk; milk was the implicated vehicle</td>
</tr>
<tr>
<td>2 (61)</td>
<td>1957</td>
<td>Japan</td>
<td>2</td>
<td>GE</td>
<td>9-yr-old girl and 6-yr-old brother</td>
<td>Strains with identical biochemical and serologic properties</td>
</tr>
<tr>
<td>3 (106, 107)</td>
<td>1979</td>
<td>Canada</td>
<td>15</td>
<td>GE</td>
<td>Inpatients</td>
<td><em>H. alvei</em> (n = 5) as pure or predominant growth; convalescent phase, 4/5 negative for hafniae; all strains biotype 1, serotype O3:H−</td>
</tr>
<tr>
<td>4 (106, 107)</td>
<td>1979</td>
<td>Canada</td>
<td>25</td>
<td>GE</td>
<td>Inpatients</td>
<td><em>H. alvei</em> (n = 8) as pure or predominant growth; strains biotype 1, serotype O3:H+</td>
</tr>
<tr>
<td>5 (54)b</td>
<td>2003</td>
<td>Russia</td>
<td>72</td>
<td>GE, neurotoxicosis</td>
<td>Inpatients</td>
<td><em>H. alvei</em> St− isolated; present in high concn in acute phase; disappearance in convalescent phase; rise in antibody titers to <em>H. alvei</em>; rotavirus (n = 8)</td>
</tr>
</tbody>
</table>

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*a* GE, gastroenteritis.

*b* Article in Russian, only English abstract translation available.

*c* St−, heat-stable enterotoxin.
treated with oxytetracycline, to which he responded favorably. Two years later, a second case of H. alvei bacteremia was described; this was in another 58-year-old male with suspected myocardial infarction (31). H. alvei was repeatedly isolated from blood cultures for over a month despite multiple antibiotic regimens, leading to the suspicion of a hidden nidus of infection. No such focus was ever identified, and it was postulated that the bacteremia continually seeded from the genitourinary tract and retention catheter. These two publications provided strong evidence that H. alvei could cause extraintestinal infections. A study from the Mayo Clinic in 1971 confirmed that hafniae could cause secondary illnesses associated with wounds/abscesses and respiratory problems, including bronchopneumonia (138).

To date there have been three reports describing a series of cases involving infections or asymptomatic colonization with H. alvei; in each of these studies the number of patients ranged from 8 to 61 (59, 105, 138). In one investigation of 17 patients where most hafniae isolated were judged to be commensals, 59% of all isolates were determined to have been transmitted nosocomially (138), while in a smaller study of 8 patients, 63% of the infections were found to be community acquired (105). Most individuals (63% to 93%) colonized or infected with hafniae have had underlying diseases, the most common of which are cancer (in particular hematologic malignancies), surgery, trauma, chronic or acute pulmonary disease, cirrhosis/hepatitis, and pancreatitis. In the largest series of cases, 66% of the patients were intubated, including all those from whom H. alvei was recovered from the respiratory tree (59). Two studies found that 37% to 55% of persons colonized/infected with Hafnia had received antibiotics prior to the isolation of this microorganism. No deaths attributable to H. alvei were recorded in any of these three studies.

From 12% to 75% of clinical specimens yield H. alvei in pure culture (59, 105, 138). Sites most likely to yield hafniae as the sole pathogens include urine and blood. When found as part of mixed microbial flora, they are often associated with other enteric bacteria, staphylococci, streptococci, and yeast (59). When hafniae have been isolated in pure growth or as a predominant flora, there is a better correlation with disease status (e.g., bronchopneumonia) than when H. alvei is recovered in small numbers (138).

### Bacteremia

There are no national figures on the incidence or number of cases of H. alvei bacteremia in the United States. A 2004 publication from the United Kingdom reports the number of cases of Hafnia bacteremia varying from 19 to 42 per year between 2001 and 2003 (6). If one compares the 2002 population of the United Kingdom (roughly 59 million) to that of the United States (287 million), this translates into somewhere between 92 and 204 cases of Hafnia bacteremia occurring in the United States annually, and this no doubt is an underestimate given the lack of an active surveillance system for this uncommon pathogen.

There have been relatively few cases of Hafnia bacteremia reported in the medical literature. Of those cases, 18 were published between 1967 and 2004 with enough data to characterize the series to some extent, although many of these reports lack laboratory information, including methods of identification of hafniae and biochemical characteristics of the infective strain. Of the 18 cases reported in Table 3, 12 occurred in adults, 4 in pediatric patients (2 to 13 years of age), and 2 in neonates (8 to 20 days old). Neonatal infections may be linked to frequent carriage of hafniae by women as part of the vaginal flora prior to birth (92). The symptoms associated with Hafnia sepsis are typical of gram-negative septicemias in general and include fever (38.6 to 40.5°C), occasional chills, and abdominal pain (14, 23, 26, 31, 37, 57, 72, 87, 105, 145). Diarrhea, either during or preceding the septic crisis, is not a consistent feature of H. alvei bacteremia but has been reported on several occasions. Fecal samples have been described as being bloody, melenic, or tarry in consistency (26, 31, 48, 104,

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### TABLE 3. Reported cases of Hafnia bacteremia in the literature

<table>
<thead>
<tr>
<th>Case no.</th>
<th>Yr</th>
<th>Age (yr)/sex</th>
<th>Underlying condition(s)</th>
<th>Immunocompromised</th>
<th>Acquisition</th>
<th>Reference</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>1967</td>
<td>58/M</td>
<td>None</td>
<td>No</td>
<td>N</td>
<td>72</td>
</tr>
<tr>
<td>2</td>
<td>1969</td>
<td>58/M</td>
<td>None</td>
<td>No</td>
<td>NN</td>
<td>31</td>
</tr>
<tr>
<td>3</td>
<td>1971</td>
<td>69/M</td>
<td>Diabetes, chronic pyelonephritis</td>
<td>No</td>
<td>N</td>
<td>87</td>
</tr>
<tr>
<td>4</td>
<td>1975</td>
<td>13/F</td>
<td>None</td>
<td>No</td>
<td>N</td>
<td>55</td>
</tr>
<tr>
<td>5</td>
<td>1977</td>
<td>20/M</td>
<td>Trauma (gunshot)</td>
<td>No</td>
<td>NN</td>
<td>14</td>
</tr>
<tr>
<td>6</td>
<td>1987</td>
<td>6/M</td>
<td>Nonlymphocytic leukemia</td>
<td>Yes</td>
<td>NN</td>
<td>145</td>
</tr>
<tr>
<td>7</td>
<td>1988</td>
<td>20 days/M</td>
<td>Low birth weight, premature</td>
<td>Yes</td>
<td>NN</td>
<td>48</td>
</tr>
<tr>
<td>8</td>
<td>1991</td>
<td>63/M</td>
<td>Lung cancer</td>
<td>Yes</td>
<td>N</td>
<td>84</td>
</tr>
<tr>
<td>9</td>
<td>1996</td>
<td>8/M</td>
<td>HIV+</td>
<td>Yes</td>
<td>NN</td>
<td>26</td>
</tr>
<tr>
<td>10</td>
<td>1996</td>
<td>37/M</td>
<td>Acute pancreatitis</td>
<td>No</td>
<td>N</td>
<td>59</td>
</tr>
<tr>
<td>11</td>
<td>1996</td>
<td>49/F</td>
<td>Sigmoid diverticulitis</td>
<td>No</td>
<td>N</td>
<td>59</td>
</tr>
<tr>
<td>12</td>
<td>1997</td>
<td>61/F</td>
<td>Biliary cirrhosis</td>
<td>Yes</td>
<td>N</td>
<td>10</td>
</tr>
<tr>
<td>13</td>
<td>1997</td>
<td>2/M</td>
<td>Autoimmune hepatitis</td>
<td>Yes</td>
<td>NN</td>
<td>10</td>
</tr>
<tr>
<td>14</td>
<td>1997</td>
<td>54/F</td>
<td>HIV+</td>
<td>Yes</td>
<td>NN</td>
<td>37</td>
</tr>
<tr>
<td>15</td>
<td>2000</td>
<td>79/F</td>
<td>Liver cirrhosis, cholangiocarcinoma</td>
<td>Yes</td>
<td>NN</td>
<td>105</td>
</tr>
<tr>
<td>16</td>
<td>2000</td>
<td>73/F</td>
<td>Recklinghausen syndrome</td>
<td>No</td>
<td>NN</td>
<td>57</td>
</tr>
<tr>
<td>17</td>
<td>2004</td>
<td>69/M</td>
<td>Stromal tumor</td>
<td>No</td>
<td>NN</td>
<td>104</td>
</tr>
<tr>
<td>18</td>
<td>2004</td>
<td>8 days/M</td>
<td>None</td>
<td>Yes</td>
<td>NN</td>
<td>23</td>
</tr>
</tbody>
</table>

a M, male; F, female.

b Known at time of admission or diagnosis. HIV+, HIV positive.

c N, nosocomial; NN, nonnosocomial.

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**Underlying condition(s)**

- Diabetes, chronic pyelonephritis
- Low birth weight, premature
- Lung cancer
- HIV+ (known at time of admission or diagnosis)
- Acute pancreatitis
- Sigmoid diverticulitis
- Biliary cirrhosis
- Autoimmune hepatitis
- HIV+ (known at time of admission or diagnosis)
- Liver cirrhosis, cholangiocarcinoma
- Recklinghausen syndrome
- Stromal tumor

---

**Immunocompromised**

- No
- Yes

---

**Acquisition**

- N (nosocomial)
- NN (nonnosocomial)

---

**Reference**

- 72
- 31
- 87
- 55
- 14
- 145
- 48
- 84
- 26
- 59
- 59
- 10
- 10
- 10
- 37
- 105
- 57
- 104
- 23
Additional laboratory studies, when reported, include an elevated level of C-reactive protein (6.8 to 37.3 mg/dl) and leukocytosis (14,200 to 32,600 mm³) (23, 26, 37, 57, 105). A majority of these blood-borne infections occur in males (71%) and are community acquired. The times of first positive Hafnia blood cultures in hospitalized patients have ranged from 1 to 41 days after admission. Half of all bacteremic patients were immunocompromised, with malignancies, liver disease, or human immunodeficiency virus (HIV) infection being the most common underlying conditions responsible for their impaired immune status. Several persons have also developed Hafnia bacteremia subsequent to liver transplants (10), and in one case Hafnia bacteremia occurred in a soldier following a gunshot wound and surgery (14). Two cases of pneumonia intestinalis accompanying H. alvei bacteremia have also been described, one in a 6-year-old boy with nonlymphocytic leukemia (145) and another in a 20-day-old infant with necrotizing enterocolitis (48).

Although all 18 H. alvei septicemia cases listed in Table 3 appear to have resolved successfully with medical intervention, including one case with septic shock and disseminated intra-vascular coagulation (84), an unusual finding in several reports was the chronic nature of the bacteremia. In addition to the report of Englund (31), cited above, Grajwer and associates (55) reported a second case of chronic polymicrobic bacteremia involving H. alvei in a healthy 13-year-old girl. H. alvei was isolated from seven blood culture bottles (six pure culture) 6 weeks after clearance of her initial septic episode. Her infection resolved, but only after a prolonged course of treatment that included aminoglycosides and an antipseudomonal penicillin. A third case of chronic septicemia has been reported by Conte et al. (26); this was in an HIV-positive 8-year-old boy who had two separate bacteremic episodes 18 days apart that were apparently caused by the same strain. The authors could find no apparent reason for the septic relapse after appropriate antimicrobial chemotherapy for the initial episode.

Most cases of H. alvei bacteremia are monomicrobial, although rare instances of polymicrobial septicemia involving Pantoea agglomerans (55) and Enterococcus faecalis (10) have been reported; the number of positive blood cultures in each case report ranged from one to seven. Besides blood, H. alvei has been recovered from other anatomic sites in approximately 33% of cases, including hepatic abscesses, pancreatic pseudocyst fluid, sputum, pleural fluid, feces, and a central venous catheter (10, 37, 48, 59). Although the source of H. alvei bacteremias is unknown in most instances, the presumed origin of infections is the gastrointestinal or respiratory tract.

Hafnia alvei (“Enterobacter hafnia”) meningitis in a 1-year-old Iranian child with intermittent fevers of 1 month in duration has been reported (88). A cerebrospinal fluid tap grew H. alvei (no blood culture results were reported), and she was treated with gentamicin, chloramphenicol, and tetracycline. Although she started to improve, she died of nosocomially acquired Pseudomonas aeruginosa sepsis.

Gastroenteritis

A controversial issue. H. alvei has a checkered past regarding its possible or reputed association with diarrheal disease. This association is chronicled in early publications from the onset of the discovery of type 32011 in the mid-1940s through the 1950s, including anecdotal stories linking this genus to cases of gastrointestinal disease (61, 134). By the 1960s, interest in hafniae as enteric pathogens had essentially waned, although sporadic mention of their possible role in diarrhea occasionally surfaced, such as in a report by Ratnam (107) describing two Canadian outbreaks of gastroenteritis associated with H. alvei.

In 1991, renewed interest in hafniae as bona fide enteropathogens was reawakened by a report of Albert et al. (4) describing diarrhea thought to be caused by H. alvei in a 9-month-old girl. This strain elicited diarrhea in rabbits and was found to contain the attaching-and-effacing gene (eae) of enteropathogenic E. coli. Other reports by the same authors followed, describing similar H. alvei strains linked to diarrhea and possessing the eae gene (5). These publications further triggered a series of case histories (109, 141) documenting Hafnia as a cause of acute gastroenteritis, with each of these studies citing the prior work of Albert et al. (4, 5), and a number of other studies and reviews cited accumulating evidence regarding the reputed pathogenicity of H. alvei (59).

Although the strains described by Albert et al. were initially identified as H. alvei by using the API 20E strip, subsequent studies by independent research teams suggested that these strains might not in fact belong to this genus. These conclusions were reached on the basis of 16S rRNA gene sequencing, the failure of the isolates described by Albert et al. to be lysed by a Hafnia-specific bacteriophage, and other atypical biochemical properties (see “LABORATORY ISOLATION AND IDENTIFICATION” below) (65, 69, 114). Further study indicated that the strains described by Albert et al. belonged to the genus Escherichia (69), and a proposal to include them in the genus Escherichia as a new species, E. albertii, was formally made by Huys and associates in 2003 (63).

Is H. alvei an enteropathogen? Given the fact that the most credible recent information supporting the case for Hafnia as an enteric pathogen was actually based upon studies performed on strains that are not hafniae but rather members of the genus Escherichia, where does this leave us with isolates of H. alvei sensu stricto?

The single best indicator of the enteropathogenicity of any organism is the isolation of only that bacterium from multiple patient samples during the acute phase of an outbreak. When such isolates are related to one another, and hopefully to strains recovered from a vehicle of infection such as consumable products, the incriminating evidence becomes even stronger. In the case of H. alvei, several outbreaks attributed to this microorganism have been described (Table 2), but none of these completely meet the definition listed above. For the outbreaks reported to date, the best and most detailed report is that of Ratnam and others (106, 107). In support of the pathogenicity of these strains were the recovery of H. alvei as predominant or heavy pure growth from 13 of 23 stools (57%) involved in these two outbreaks, the fact that strains from both outbreaks possessed the same biotype (biotype 1) and serotype (O3:HNM), and the fact that of over 1,000 stools subsequently tested after these outbreaks, only one yielded H. alvei. However, in this study, no virulence factor was identified and there was no human immune response recorded or pathological data provided to corroborate culture results, and subsequent studies...
produced (see “PATHOGENICITY” below). Thus, at present, there
are very few epidemiologic, clinical, and laboratory data to
mean that hafniae are not enteric pathogens, just that much
more work and evidence need to be accumulated to support
that hafniae are not enteric pathogens, just that much
more work and evidence need to be accumulated to support
such a position.

Respiratory Tract Infections

Next to the gastrointestinal tract, hafniae are most com-
monly recovered from respiratory secretions such as sputum,
tracheal and bronchial aspirates, nasal smears, and broncho-
alveolar lavage fluid and from the mouth and throat (59, 138).
The vast majority of these isolates are not clinically significant.
Washington et al. (138) isolated eight strains of H. alvei in a
2-year period at the Mayo Clinic. Of these eight isolates, only
two (25%), from sputum and trachea, were determined to be
clinically relevant as secondary pathogens; both were associ-
ated with pneumonia, with one of these accompanied
with a fiberoptic bronchoscopy performed with a protected sheath
brush. She was treated with aztreonam but subsequently died,
and it is unclear how much hafniae contributed to her eventual
demise. Collectively, these seven patients had a mean age of 60
years and had a variety of underlying diseases, including
chronic obstructive pulmonary disease, lung cancer, and HIV
infection. A case of nosocomial pneumonia due to Hafnia in a
person on mechanical ventilation for 12 days duration has also
been reported from Switzerland; H. alvei was again recovered
in pure culture from a bronchoscopy specimen (40).

Miscellaneous Infections

H. alvei has been reported to be a rare cause of a variety of
other extraintestinal infections. The analysis of these case re-
ports is complicated by two facts. First, hafniae are often re-
covered from such syndromes as part of a mixed infection, and
it is difficult to assess how much H. alvei contributes to the
overall disease process. Second, in other instances there is only
circumstantial evidence at best that Hafnia caused the illness
or syndrome in question. Often this circumstantial evidence
involves the recovery of these bacteria in large concentrations
from gastrointestinal samples. Whether such a circumpect
analysis is appropriate is unclear.

Although wounds would be thought to be common sites
from which to recover hafniae, there are surprisingly few pub-
lications in the literature on this topic (40). Berger et al. (14)
recovered H. alvei along with P. aeruginosa from a nosocomial
wound infection of a 19-year-old soldier who had sustained a
gunshot wound. What if any role hafniae played in the infec-
tious process is unclear. H. alvei has also been isolated from a
psosas abscess, but this gram-negative bacterium was isolated
simultaneously with Streptococcus bovis (105). In a series of 17
cases of Hafnia colonization/infection, 3 were associated with
wounds or abscesses (138). In all three cases they were thought
to be secondary pathogens, although again they were recov-
ered as part of a mixed microbial flora. Detailed information
on these cases was not provided. The only convincing case of a
wound infection attributed to H. alvei was reported by Augustin
and Cunha (7) and involved a 44-year-old male who sustained
a penetrating trauma to his right buttock by carpet nails. A
subsequent abscess developed, and 50 ml of serosanguinous
fluid grew a pure culture of H. alvei. H. alvei has infrequently
been linked to hepatobiliary disease, including two cases of
hepatic abscesses (mixed cultures and bacteremia after liver
transplantation) (10). Two cases of cholecystitis associated
with Hafnia infection have also been published (105, 144). In
one of these cases, that of a 67-year-old woman who apparently
developed biliary disease after handling fish, hafniae were recov-
ered from the infected gall bladder in small numbers in
comparison to the predominant Aeromonas. Most recently, a
single case of spontaneous bacterial peritonitis caused by H.
alvei in a 60-year-old Saudi man with underlying mesothelioma
has been reported (3).

The urinary tract occasionally harbors hafniae. Ramos and
Dámaso (105) isolated H. alvei in pure culture from urine
specimens of three different patients and felt they were clini-
cally significant, while Washington et al. isolated hafniae in
small numbers from a similar number of persons without signs
of disease and determined that they were commensals (138).
Septic arthritis associated with the recovery of both H. alvei
and *Klebsiella oxytoca* from joint fluid of a 29-year-old woman after anterior cruciate ligament reconstruction has been reported (83). A case of reactive arthritis of greater than 10 weeks in duration has been linked to *H. alvei* enteritis (93). Although this pathogen was recovered in large numbers from feces, the joint fluid, blood, and urine were culture negative; the role of hafniae in this illness is debatable. *Hafnia* endophthalmitis has been reported in two instances. One was as part of a coinfection with *Salmonella enterica* subsp. *arizonae* (22). In this specific case, involving a 55-year-old Hispanic woman, the source of infection appeared to be snake powder from Mexico used to season food. A second case has recently been reported from Spain (118). In both instances, visual perception was lost despite aggressive medical intervention.

**PATHOGENICITY**

Understanding of which virulence factors play a role in the pathogenicity of *Hafnia* in regard to both human and animal infections is presently in its infancy. The limited amount of research in this area is most likely attributable to the low frequency of occurrence of this species in human illnesses and the fact that there are no clear-cut disease syndromes specifically associated with *H. alvei*. A further obstacle has been the unfortunate association of the *eae* gene with enteritis-producing strains that were thought to be *H. alvei* (4) but that were determined years later to actually be members of a new species, *E. albertii* (63).

What is presently known regarding *Hafnia* pathogenicity can be looked at from two perspectives, that is, virulence factors potentially operative in extraintestinal infections and those restricted primarily to the intestine. With all of these data there are numerous “gaps” in information, and many of the original findings date back 2 decades or more, where the description of strains and techniques is often incomplete.

Lethality studies, such as determinations of 50% lethal dose, to compare the overt pathogenicity of *H. alvei* to those of other members of the family, including *E. coli*, *Shigella*, and *Salmonella*, have not been performed. There is some sketchy and anecdotal information that hafniae can produce morbidity and mortality in certain animal models. One study found that 60% to 100% of rainbow trout injected subcutaneously and white mice and guinea pigs injected intravenously with aliquots of broth cultures containing hafniae died within 24 h (mice) to 10 days (fish); mice injected subcutaneously or orally challenged did not die (46). Unfortunately, the inoculum used in these experiments was not quantified, although the challenge dose was likely very high. One other study mentions that mice injected intraperitoneally with hafniae did not succumb to infection (91).

Approximately 35% to 55% of *H. alvei* strains are resistant to the bactericidal activity of human or bovine serum (67, 102). The classical pathway appears to be the major route by which serum-susceptible strains are killed; preincubation of bovine serum with *H. alvei* O-specific polysaccharide reduces or eliminates the bactericidal activity (67). Iron-scavenging mechanisms can also play important roles in pathogenesis. *H. alvei* strains produce siderophores as detected by a universal chemical assay (71, 102). One study found that the *H. alvei* siderophore was of neither the hydroxamate (aerobactin) nor the catechol (enterobactin) type (67). The siderophores of *H. alvei* can, however, take up ferric enterobactin and ferrichrome via an *E. coli* FepA-like outer membrane protein (119). Other factors described for hafniae, which may or may not play a role in their pathogenicity, include mannose-sensitive and mannose-resistant (MR/K, type 3 fimbria) hemagglutinins (102). Enzymatic properties associated with some pathogenic bacteria, including hemolysin, protease, elastase, and lecithinase activities, have not been detected in hafniae (71).

As early as the 1950s some strains of *H. alvei* were thought to be enteropathogenic, based upon results in animal studies. Sakazaki (120) found that 15 *H. alvei* strains produced enteritis in the ligated rabbit intestine model. However, no credible data evolved over the next 30 years. Numerous studies in the 1990s confirmed the fact that true *H. alvei* did not contain the *eae* gene as had been previously reported (65, 69), and in fact one Spanish study found all 102 *H. alvei* strains tested to be *eae* negative by both PCR and dot blot analysis (126). Other fecal isolates associated with outbreaks of gastroenteritis have uniformly been negative for heat-stable and heat-labile enterotoxins, invasiveness, and production of Vero cell cytotoxins (106).

Approximately 70% of *Hafnia* strains carry one or no plasmids (128). However, plasmid-bearing strains often harbor very large extrachromosomal elements ranging from 128 to 256 kb (128). Not much is known regarding what genes or functions are encoded by these plasmids, although atypical properties, such as lactose fermentation, have been attributed to plasmid carriage by rare strains (81). Recently, two small plasmids designated pAhVA and pAhVB were found to encode bacteriocins, or alveicins (140). These bacteriocin-carrying plasmids are very small (5.1 to 5.2 kb), and the protein products produced from these genes range from 358 to 408 amino acids in size. The alveicins are active only against their own species; about 15% of *H. alvei* strains screened appear to produce alveicins.

**LABORATORY ISOLATION AND IDENTIFICATION**

**Cultural Characteristics**

Not much attention has been given to optimizing cultural conditions for the specific recovery of hafniae. This can be attributed mostly to two facts: the lack of any substantial evidence that *H. alvei* is indeed a bona fide enteropathogen and the infrequent occurrence of hafniae as primary or secondary pathogens in other highly contaminated specimens, such as the respiratory tract and wounds. Even classic reference texts in the field of microbiology deal little with the isolation and cultural characteristics of *H. alvei* (86, 121, 122). *Hafnia* grows in media containing 2% to 5% NaCl, over a pH range of 4.9 to 8.25, and over thermal gradients of 4°C to 44°C (56); the optimum temperature for growth has been reported as 35°C (89). Some investigations have found growth at 44°C to be variable (73). There is general agreement that almost 100% of *Hafnia* strains grow on MacConkey, Hektoen enteric, eosin-methylene blue, and xylose-lysine-deoxycholate agars, all of which are differential to moderately selective media (56, 123, 124). For other media, such as desoxycholate-citrate agar (DCA), there are conflicting data, in that Greipson and Priest (56) reported that all 47 strains they tested grew on DCA, while Sakazaki (123) found that DCA was inhibitory for many...
hafniae. Regarding more inhibitory selective media, from 25% to 60% of strains fail to grow on Salmonella-Shigella (SS) agar, while 75% to 100% of isolates are inhibited on brilliant green medium (68, 73).

Classic strains of H. alvei are lactose and sucrose negative and as such appear as nonfermenting colonies on enteric isolation media. On moderately selective agars, they typically appear as large, smooth, convex, translucent colonies of 2 to 3 mm in diameter with an entire edge (68, 123); some may exhibit an irregular border. Several media have been employed on rare occasions to recover hafniae from fecal specimens. Using SS agar, Matsumoto (85) was able to isolate H. alvei from 13% of the 1,913 stools tested in 1961; this number probably represents a minimum value, since SS agar is known to be inhibitory to many Hafnia strains. Sakazaki (121) took advantage of the inability of hafniae to ferment β-sorbitol to recover H. alvei from 42% of 598 stools by using deoxycholate-lactose-sucrose-β-sorbitol agar, while comparable recovery rates on MacConkey agar were only 2%. H. alvei has also been isolated on sorbitol-MacConkey agar, a medium designed for the isolation E. coli O157:H7 (38). For environmental samples such as food, both violet red bile lactose and fecal coliform agars have been successfully used to isolate hafniae from consumables, including cheese and sausage (56, 80). Enrichment broths in which some but not all hafniae grow include selenite and tetrathionate broths (123).

Several additional cultural characteristics of H. alvei bear mentioning. Because H. alvei strains are H2S negative, they can occasionally be mistaken for H2S-negative Salmonella on media such as enteric agar (123). Similarly, some hafniae also produce red or pink colonies on xylose-lysine-deoxycholate agar (121). Some Hafnia strains or variants are lactose positive, a trait that in some instances has been determined to be plasmid mediated (81). Atypical H. alvei colonies on violet red bile lactose and fecal coliform agars have also been described (80).

Identification

The genus Hafnia consists of strains that conform to general characteristics for inclusion in the family Enterobacteriaceae, being gram-negative, peritrichously flagellated rods that are oxidase negative, nonsporulating, produce nitrate reductase, and are positive for enterobacterial common antigen. They are facultatively anaerobic, producing acid with or without gas from the metabolism of d-glucose and other carbohydrate or carbohydrate-like compounds (35). Major distinguishing phenotypic features of the Hafnia complex include being indole negative, lysine and ornithine decarboxylase positive, arginine dihydrolase negative, and H2S and D-sorbitol negative, with production of acid from the fermentation of d-glucose, l-αrabinose, and l-rhamnose. Most strains are also VP positive and are motile (35). Strains that deviate from this ideal phenotype do exist but are uncommon (70, 120). They can be mistaken for H2S-negative Salmonella on primary plating media or can be confused with misidentified as either Enterobacter or Serratia. On occasion, members of the genus Hafnia can also be mistaken for E. coli or Yokenella regensburgei (123). Tests useful in distinguishing H. alvei from Y. regensburgei include VP and glycerol fermentation (both of which are positive for hafniae) and acid production from melibiose (which is negative for hafniae) (79, 133).

Temperature can play a prominent role in phenotype expression in H. alvei. Strains typically incubated at lower (22°C) rather than higher (35°C) temperatures are more likely to produce acetylthioarabinofuranosyl, form gas, and be more actively motile (89, 120, 123), while conversely, the methyl red test is positive at elevated temperatures. Early studies suggested that Hafnia strains were H2S positive (89), although these strains are invariably H2S negative on traditional screening media such as TSI (55). As with many Enterobacteriaceae that are H2S negative on TSI, H. alvei strains are moderately to weakly H2S positive on thiosulfate-containing media such as gelatin-cysteine-thiosulfate medium at 48 h (unpublished data).

Commercial systems. Most recent published studies involving the identification of Enterobacteriaceae on commercial systems have had little trouble identifying H. alvei. These systems include both MicroScan conventional and rapid panels (11, 42, 94, 116) and bioMérieux Vitek GNI+ and Vitek 2 systems (41, 42, 95). In five of these studies, 61 of 62 isolates (98%) were correctly identified as H. alvei (11, 41, 42, 94, 116). The single exception was an atypical strain that was esculin positive and l-rhamnose negative and that was identified as a rare biotype on MicroScan conventional panels (116). A sixth study conducted by the CDC found that 6 of 10 H. alvei strains yielded excellent identifications on Vitek ID-GNB, 3 of 10 strains produced a probability of a good identification, and 1 isolate was unidentified (95). An earlier study by the CDC found that four of nine H. alvei strains were identified as inactive E. coli by Vitek GNI+ (96). The converse situation can also occur, where species such as Salmonella enterica subsp. arizonae and Y. regensburgei have been misidentified as hafniae (96). A more recent observation concerns the recently described species E. albertii. Originally identified as H. alvei on the API 20E system by Albert et al. (4), this observation has subsequently been confirmed by other groups, and such misidentifications occur not only on the API 20E but also on Vitek and MicroScan rapid panels (1). Conventional biochemical tests are needed to distinguish between these two taxa at present (see “Conventional methods” below).

Agreement between commercially available systems and conventional biochemical formats varies with each test. Rodriguez et al. (116) found that there was generally good agreement between MicroScan Combo Negative type IS panels, conventional biochemical test results, and recorded character traits in Bergey’s Manual of Systematic Bacteriology (122) for most phenotypic results. However major differences were noted in test correlation between commercial and conventional formats for o-nitrophenyl-β-D-galactopyranoside (47.6%), urease (23.8%), VP (14.2%), and citrate (4.7%). For arginine dihydrolase, correlation of assays improved from 80.9% to 100% when test results were read manually rather than by using the WalkAway.

Conventional methods. Despite the fact that most strains of H. alvei can be easily identified using commercial systems, conventional biochemical test results can be necessary under a number of circumstances. These include when probability of a correct identification is low, when H. alvei strains with two or more atypical properties (e.g., VP negative and urease positive) are identified, when hafniae are isolated from an anatomic site not traditionally associated with this species (e.g., blood), for unusual disease syndromes linked to Hafnia infection, for chronic or recurring illnesses, and for publication
purposes. One study assessing the reproducibility of results from 22 laboratories in the United Kingdom to identify a series of enteric bacteria in a multipoint identification scheme found H. alvei to be one of the most difficult organisms to identify, with 37% of strains reported with either an incorrect identification or no identification at all (143).

Specialized tests. (i) Hafnia-specific phage. In 1967, Guinée and Valkenburg (58) described a lytic bacteriophage that was specific for the genus Hafnia. Phage 1672 lysed all 100 Hafnia strains tested against it and none of over 200 other enteric bacteria, including Salmonella, Citrobacter, Enterobacter, Klebsiella, and Serratia strains. The test is performed by inoculating a 24-h broth culture onto the surface of a nutrient agar plate and, after removal of excess fluid, adding a drop of bacteriophage suspension to the plate. After overnight incubation at 37°C, a clear lytic zone surrounded by growth is recorded as positive (123). This is an excellent test to definitively identify members of the genus Hafnia and has been used extensively in strain characterization (71, 108). Phage 1672 is available from the American Type Culture Collection (Manassas, Va.) (catalogue no. 51873-B1). The propagating host strain for this phage is H. alvei ATCC 51873 (HER 1272 [1672]).

(ii) GAD. The assay for glutamate decarboxylase (GAD) activity can be performed in a single-tube format, as described by Rice and colleagues (111). A change in color from blue to yellow is considered to be a positive test. Escherichia species, including E. coli and E. albertii, are GAD positive, while hafniae are GAD negative (71, 111).

(iii) LPA. Among enteric bacteria, strong L-prolineamino-peptidase (LPA) activity is detectable only in H. alvei, Serratia marcescens, and Serratia liquefaciens (25, 34). The assay can be performed in tube or microtiter format or using commercially available kits (Aminopeptidase Wee-Tab; Key Scientific Products). Performance in tube or microtiter format or using commercially available kits (Aminopeptidase Wee-Tab; Key Scientific Products) can be performed in a single-tube format, as described by Rice and colleagues (111). A change in color from blue to yellow is considered to be a positive test. Escherichia species, including E. coli and E. albertii, are GAD positive, while hafniae are GAD negative (71, 111).

Biotypes. (i) Biotype 1 strains. Farmer and colleagues at CDC defined biotype 1 strains in 1985 as “H. alvei biogroup 1” or “H. alvei—brewery biogroups” (36). These strains are also synonymous with the previous epithet name Obesumbacterium proteus and the H. alvei biogroup 1 strains listed in Table 1 of reference 35 (J. J. Farmer, personal communication). Biogroup 1 strains have not been recovered from clinical specimens (36). Major phenotypic differences distinguishing “brewery strains” from typical H. alvei (Table 1 of reference 35), which includes both DNA groups, are listed in Table 4.

(ii) Barbe biotypes. In 1969, Barbe described two biotypes of H. alvei based upon fermentation of D-arabinose and salcin and on esculin and arbutin hydrolysis (doctoral thesis cited in reference 21). These two biotypes did not correlate completely with the two known DNA hybridization groups within the H. alvei complex. The number of Barbe biotypes has subsequently been expanded to four by other investigators in a phenotypic and genotypic analysis of 73 H. alvei strains (71). Types 1 and 2 predominate (Table 5).

Distinguishing H. alvei complex from Escherichia albertii. There has been considerable controversy in the literature regarding the enteropathogenicity of H. alvei, based upon the initial misidentification of E. albertii strains as H. alvei (see “Gastroenteritis” in “CLINICAL SIGNIFICANCE” above). The two groups share a number of phenotypic features, including lack of indole production, inability to ferment lactose or α-sorbitol, and similar antimicrobial susceptibility profiles (69). This controversy took over 10 years to resolve with the publication of a new species, E. albertii (63). Several groups have investigated these and similar strains either prior to or after the naming of E. albertii. These phenotypic and molecular tests are presented in Table 6.

Phenotypic and genotypic separation of H. alvei DNA groups 1 and 2. One of the difficult challenges that microbiologists face is trying to develop biochemical tests that can readily distinguish most strains of DNA group 1 (H. alvei sensu stricto) from DNA group 2 (unnamed Hafnia species) isolates. While the initial studies by Steigerwalt et al. (131) in 1976 detected two distinct hybridization groups within the genus, no differential tests to separate these genomospecies were noted. The first true breakthrough in this area involves the defining publication by Don Brenner (21), where a number of biochemical characteristics were found to be preferentially associated with either DNA group 1 or 2. These tests included motility at 24 h, hydrolysis of arbutin, fermentation of salcin or inulin, and utilization of sodium acetate. In that study, Brenner noted that these two groups could be distinguished from one another by a series of tests but that no single test was completely discriminatory, although motility at 24 h was the best predictor of DNA group (DNA group 1, 9% positive; DNA group 2, 100% positive). Furthermore, Barbe’s biotypes (see above) did not correlate perfectly with these two DNA relatedness groups (21).

**TABLE 4. Distinguishing features of H. alvei and H. alvei biogroup 1 strains**

<table>
<thead>
<tr>
<th>Test</th>
<th>H. alvei</th>
<th>H. alvei biogroup 1</th>
</tr>
</thead>
<tbody>
<tr>
<td>Methyl red</td>
<td>Variable</td>
<td>+ (85)</td>
</tr>
<tr>
<td>Ornithine decarboxylase</td>
<td>+ (98)</td>
<td>Variable + (45)</td>
</tr>
<tr>
<td>Motility</td>
<td>+ (85)</td>
<td>− (0)</td>
</tr>
<tr>
<td>Growth in KCN</td>
<td>+ (95)</td>
<td>− (0)</td>
</tr>
<tr>
<td>D-Glucose (gas)</td>
<td>+ (98)</td>
<td>− (0)</td>
</tr>
<tr>
<td>Acid from:</td>
<td></td>
<td></td>
</tr>
<tr>
<td>l-Arabinose</td>
<td>+ (95)</td>
<td>− (0)</td>
</tr>
<tr>
<td>l-Rhamnose</td>
<td>+ (97)</td>
<td>− (0)</td>
</tr>
<tr>
<td>Glycerol</td>
<td>+ (95)</td>
<td>− (0)</td>
</tr>
</tbody>
</table>

* Data are from reference 35. Numbers in parentheses indicate percent positive for indicated character. +, positive; −, negative.

**TABLE 5. Modified Barbe biotypes**

<table>
<thead>
<tr>
<th>Test</th>
<th>1</th>
<th>2</th>
<th>3</th>
<th>4</th>
</tr>
</thead>
<tbody>
<tr>
<td>Escluin</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>−</td>
</tr>
<tr>
<td>Arbutin</td>
<td>−</td>
<td>+</td>
<td>+</td>
<td>−</td>
</tr>
<tr>
<td>Salcin</td>
<td>−</td>
<td>+</td>
<td>+</td>
<td>−</td>
</tr>
<tr>
<td>D-Arabinose</td>
<td>−</td>
<td>+</td>
<td>−</td>
<td>+</td>
</tr>
</tbody>
</table>

* Adapted from reference 71.
TABLE 6. Phenotypic and genotypic features useful in distinguishing H. alvei sensu stricto and E. albertii

<table>
<thead>
<tr>
<th>Property type</th>
<th>Test*</th>
<th>Result fora:</th>
<th>H. alvei</th>
<th>E. albertii</th>
</tr>
</thead>
<tbody>
<tr>
<td>Phage lysis</td>
<td>φ1672</td>
<td>+ (100)</td>
<td>– (0)</td>
<td></td>
</tr>
<tr>
<td>Biochemical</td>
<td>KCN</td>
<td>+ (98)</td>
<td>– (0)</td>
<td></td>
</tr>
<tr>
<td></td>
<td>VP</td>
<td>+ (100)</td>
<td>– (0)</td>
<td></td>
</tr>
<tr>
<td></td>
<td>GAD</td>
<td>– (0)</td>
<td>+ (100)</td>
<td></td>
</tr>
<tr>
<td></td>
<td>Glycero (24 h)</td>
<td>+ (96)</td>
<td>– (0)</td>
<td></td>
</tr>
<tr>
<td></td>
<td>L-Rhamnose (24 h)</td>
<td>+ (51)</td>
<td>– (0)</td>
<td></td>
</tr>
<tr>
<td></td>
<td>D-Xylose</td>
<td>– (100)</td>
<td>+ (0)</td>
<td></td>
</tr>
<tr>
<td>Enzymatic</td>
<td>Chitinase (6 h)</td>
<td>+ (99)</td>
<td>– (5)</td>
<td></td>
</tr>
<tr>
<td></td>
<td>Hydroxypoline amidase</td>
<td>+ (100)</td>
<td>– (0)</td>
<td></td>
</tr>
<tr>
<td></td>
<td>Tripeptidase</td>
<td>+ (100)</td>
<td>– (0)</td>
<td></td>
</tr>
<tr>
<td></td>
<td>Proline deaminase</td>
<td>+ (100)</td>
<td>+ (0)</td>
<td></td>
</tr>
<tr>
<td>Molecular</td>
<td>pheO</td>
<td>– (0)</td>
<td>+ (100)</td>
<td></td>
</tr>
<tr>
<td></td>
<td>eae</td>
<td>– (0)</td>
<td>+ (100)</td>
<td></td>
</tr>
<tr>
<td></td>
<td>cdt</td>
<td>– (0)</td>
<td>+ (100)</td>
<td></td>
</tr>
<tr>
<td></td>
<td>PFGE typical ladder (XbaI)</td>
<td>– (0)</td>
<td>+ (100)</td>
<td></td>
</tr>
</tbody>
</table>

* cdt, cytotoxle distending toxin gene; eae, E. coli attaching-and-effacing gene; pheO, outer membrane protein gene; PFGE, pulsed-field gel electrophoresis.

** Composite data from references 64, 69, 70, 71, and 132. The number of strains of each species analyzed for the indicated tests ranged from a minimum of 5 isolates to a maximum of 76 strains. Numbers in parentheses indicate percent positive.

There were no major studies on this subject for over 20 years. In 2002, a study conducted by the Microbial Diseases Laboratory found that 22 Hafnia isolates could be assigned to their correct DNA relatedness groups by 16S rRNA gene sequencing (71). Biochemical characterization of these strains showed that while motility did tend to correlate with genomospecies (DNA group 1, 20% positive; DNA group 2, 71% positive), as in the study by Brenner (21), a better differential test appeared to be malonate utilization (DNA group 1, 100% positive; DNA group 2, 17% positive). Okada and Gordon (97) subsequently confirmed the discriminatory value of the malonate test in their ecologic study of hafniae in Australia. These collective data were reinforced by a 2005 study in which malonate and several other tests were found to be useful in separating these two relatedness groups (70). The proposal was also made in later study to recognize four biogroups with H. alvei sensu stricto (DNA group 1), based upon the fermentation of d-arabinose and salicin as well as esculin hydrolysis. Table 7 provides cumulative data from several studies on the best tests currently available for separation of H. alvei DNA groups 1 and 2.

**Molecular identification.** No specific probes or assays have been developed to detect the genus Hafnia. H. alvei strains do react with oligonucleotide probes developed to identify members of the family Enterobacteriaceae. These probes target specific regions of the small-subunit (16S) or large-subunit (23S) rRNA gene (15, 99).

**Serology**

The immunochemistry of Hafnia lipopolysaccharide (LPS) is extremely complicated. All H. alvei LPS appears to contain glucose, glucosamine, heptose, and 3-deoxyoctulosonic acid (117). Some LPS also contains other amino sugars or carbohydrates such as mannose, galactose, galactosamine, and mannosamine. The core oligosaccharide structure of some strains consists of an identical hexasaccharide structure composed of two d-glucose residues, three L- d-heptose residues, and one 3-deoxyoctulosonic acid residue. Other strains have an additional core fragment (117). There is extensive serologic and immunologic diversity in this genus, and active research continues in this arena (74).

Two formal serologic typing schemes for H. alvei have been published. Riichi Sakazaki and colleagues at the National Institute of Health in Tokyo, Japan, pioneered the older of these two schemes. This scheme officially recognized 68 somatic (“O”) and 34 flagellar (“H”) types (123, 124). Unfortunately, this typing system is no longer available because of the loss of many test strains for specific O- and H-antigen groups (123). A second, independent scheme proposed by the Laboratory of Opportunistic Enteric Pathogens at the Mechnikov Research Institute for Vaccines and Sera in Moscow, Russia, recognizes 39 O and 36 H types (13, 117). The availability of this second typing scheme is unknown, and we are unaware of any recently published studies reporting Hafnia serotypes or serogroups. Some Hafnia antigens are shared with other enteric groups, potentially producing serologic cross-reactions. These groups include Salmonella, Shigella, E. coli, Citrobacter freundii, and E. cloacae (123, 124, 137). Some H. alvei strains have been reported to agglutinate in O157 antisera (123).

Several other antigens in hafniae have been described. Several studies have reported that some H. alvei strains produce K...
antigens which may actually be a slime antigen (124). Other strains produce the alpha antigen described by Stamp and Stone (30). This antigen consists of a complex of three factors, and Hafnia strains that carry this antigen either contain all three or two of these three factors. Other enteric groups, such as Providencia, occasionally carry the alpha antigen as well.

ANTIMICROBIAL SUSCEPTIBILITY

Analogous to the situation described above regarding pathogenicity, there are virtually no studies available in which a large collection of Hafnia isolates have been studied for their susceptibility to different classes of antimicrobial compounds. The lone exception to this is a 2005 study by Stock et al. (132), in which 76 H. alvei isolates were investigated for their susceptibility to 69 antibiotics or drugs. The general pattern that emerges from this study is that hafniae are typically susceptible to carbapenems, monobactams, chloramphenicol, quinolones, aminoglycosides, and antifolates (e.g., trimethoprim-sulfamethoxazole) and resistant to penicillin, oxacillin, and amoxicillin plus clavulanic acid. Susceptibility to tetracyclines and cephalosporins is variable, with most H. alvei strains being susceptible to broad-spectrum and “fourth-generation” cephalosporins (cefipime) but often resistant to narrow- and expanded-spectrum compounds. A 2000 Spanish study involving enteric pathogens also found that 32 strains of H. alvei were universally susceptible to all quinolones (including gemifloxacin and grepafloxacin), cefotaxime, gentamicin, co-trimoxazole, and naladixic acid (39); 78% of the strains in that study were susceptible to doxycycline.

Some H. alvei strains produce both low-level inducible cephalosporinases (cefazidime susceptible) and high-level constitutive cephalosporinase activity that is resistant to cefazidime (50, 101, 136). These activities are chromosomally constitutive cephalosporinase activity that is resistant to cefazidime and high-level cephalosporinases (ceftazidime susceptible) and high-level constitutive cephalosporinase activity that is resistant to cefazidime. Unrelated H. alvei isolates have been studied for their susceptibility to 69 antibiotics or drugs. The general pattern that emerges from this study is that hafniae are typically susceptible to carbapenems, monobactams, chloramphenicol, quinolones, aminoglycosides, and antifolates (e.g., trimethoprim-sulfamethoxazole) and resistant to penicillin, oxacillin, and amoxicillin plus clavulanic acid. Susceptibility to tetracyclines and cephalosporins is variable, with most H. alvei strains being susceptible to broad-spectrum and “fourth-generation” cephalosporins (cefipime) but often resistant to narrow- and expanded-spectrum compounds. A 2000 Spanish study involving enteric pathogens also found that 32 strains of H. alvei were universally susceptible to all quinolones (including gemifloxacin and grepafloxacin), cefotaxime, gentamicin, co-trimoxazole, and naladixic acid (39); 78% of the strains in that study were susceptible to doxycycline.

Some H. alvei strains produce both low-level inducible cephalosporinases (cefazidime susceptible) and high-level constitutive cephalosporinase activity that is resistant to cefazidime (50, 101, 136). These activities are chromosomally encoded cephalosporinases belonging to Bush group 1 β-lactamases (Ambler class C). The ampC gene of one H. alvei strain exhibits 94% amino acid sequence homology to a plasmidborne cephalosporinase of K. pneumoniae (49). Unrelated H. alvei isolates show 85% to 100% amino acid identity in their AmpC sequences.

CONCLUSIONS

There are many unanswered questions regarding the genus Hafnia. Clearly, the most important issue concerns the frequency and type of human infections associated with this genus. If at a future date credible evidence linking this species to diarrheal disease surfaces, and given the frequency of occurrence of H. alvei in gastrointestinal specimens, its relative importance as a human pathogen may be significantly enhanced. Another unanswered question concerns the taxonomy of this genus. The naming of DNA group 2 and identification of useful tests to separate this genospecies from H. alvei sensu stricto (DNA group 1) should lead to a better appreciation of whether there are major differences between these two species in disease presentation, environmental distribution, and pathogenicity (70). We presently know very little regarding the distribution of hafniae in environmental samples, their ecologic niches other than the gastrointestinal tract, and the ability of the H. alvei complex to incite disease in animals, fish, and other wildlife. Recent work from Australia, however, suggests that significant differences in environmental distribution do occur (98). Finally, a variety of antimicrobial resistance mechanisms have been recently identified in sporadic H. alvei isolates (50, 101). At present the frequency of these resistotypes is low, but if widespread dissemination should occur, treatment of systemic H. alvei infections could become problematic.

ADDENDUM

Since the submission of this review, we have recovered a strain of H. alvei from two stool specimens of an 11-year-old girl with probable hemolytic uremic syndrome (C. Crandall, S. L. Abbott, Y. Q. Zhao, W. Probert, and J. M. Janda, Infection, in press). This strain produces a cytolytic toxin that is active on Vero cells and is morphologically indistinguishable from Shiga toxin (Stx). However, this toxin appears to be less potent and antigenically distinct, as its activity is not neutralizable when the strain is preincubated with antisera directed against both Stx1 and Stx2.

REFERENCES