



# Strategies for Prevention and Treatment of *Trichomonas vaginalis* Infections

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**SUMMARY** The last estimated annual incidence of *Trichomonas vaginalis* worldwide exceeds that of chlamydia and gonorrhea combined. This critical review updates the state of the art on advances in *T. vaginalis* diagnostics and strategies for treatment and prevention of trichomoniasis. In particular, new data on treatment outcomes for topical administration of formulations are reviewed and discussed.

**KEYWORDS** *Trichomonas vaginalis*

## INTRODUCTION

*Trichomonas vaginalis* is a human protistan parasite responsible for the most common nonviral sexually transmitted disease in the world. *T. vaginalis* is recognized as a cause of vaginitis. It can also infect the urethra and prostate in men. Although men are often asymptomatic carriers of *T. vaginalis*, dysuria and discharge have also been reported (1). In women, the disease may range from asymptomatic (up to 50% of women) to severe with serious sequelae. Classical symptoms include a malodorous and purulent discharge which results in local pain and irritation. *T. vaginalis* is implicated in reproductive tract postsurgical infections that usually remain localized to the lower part of the urogenital area. This parasite can also result in serious consequences, such as infertility, premature rupture of placental membranes, premature delivery, low-birth-weight infants, and neonatal death (2). Moreover, an increased predisposition to HIV infection has been reported for both men and women (3).

Oral metronidazole remains the recommended regimen for the treatment of trichomoniasis. However, treatment failure does occur, mainly due to significant gastrointestinal adverse effects, which have been found to be temporary and disappear after the cessation of treatment. Systemic delivery of metronidazole may also result in

**TABLE 1** Incidences and prevalences of *T. vaginalis* infection in women and men between the ages of 15 and 49 years in different world regions in 2008<sup>a</sup>

Region	Incidence				Prevalence			
	Women		Men		Women		Men	
	Per 1,000	Millions of individuals	Per 1,000	Millions of individuals	%	Millions of individuals	%	Millions of individuals
Africa	146	28.1	164.8	31.6	20.2	38.9	2	3.9
The Americas	177.7	42.5	180.6	43	22.0	52.7	2.2	5.2
Southeast Asia	40.3	18.5	50.1	24.3	5.6	25.7	0.6	3.0
Europe	51.7	11.6	48.4	10.9	5.8	13.0	0.6	1.3
Eastern Mediterranean	64.0	9.7	66.1	10.6	8.0	12.0	0.8	1.3
Western Pacific	45.6	21.9	47	23.8	5.7	27.2	0.6	2.9

<sup>a</sup>The data used are from the WHO (137).

allergy and drug resistance (4). In this context, intravaginal drug delivery allows better-tolerated prevention and treatment options for trichomoniasis that avoid systemic adverse effects.

This article offers an update on the progress in *T. vaginalis* diagnostics and chemotherapy for the treatment of trichomoniasis. A vaccine strategy for the prevention of *T. vaginalis* vaginal infections and formulations for the treatment of vaginal trichomoniasis are discussed. Although reviews of the literature do exist (5–14), documentation on the vaginal administration of drugs and strategies for the prevention of *T. vaginalis* infection has, until now, been inconsistent.

## EPIDEMIOLOGY AND TRANSMISSION

According to the WHO estimate, among the total number of new cases of curable sexually transmitted infections (STIs) in 2008 (498.9 million adults aged 15 to 49 years), 276.4 million resulted from *T. vaginalis*. The prevalence and incidence of vaginal *T. vaginalis* infection are higher in African regions and in the Americas than in other parts of the world (Table 1). Different studies conducted in Denmark, Great Britain, and France showed that the prevalence is declining in industrialized nations (15, 16). Iranian studies determined the prevalence to be 2 to 8% (17). However, based on cultural and social factors, this rate is underestimated and may be higher than 30% (17).

No association was found in the United States between trichomoniasis and HIV infection (9), while a study conducted in Africa demonstrated that HIV infection increased the prevalence of vaginal trichomoniasis (18). The exact mechanism by which *T. vaginalis* facilitates HIV transmission is poorly understood, but it is probably related to a higher susceptibility to bacterial vaginosis (19). *T. vaginalis* infections also involve inflammatory processes, which may facilitate vaginal infection with HIV (20, 21).

Trichomoniasis is a venereal disease that affects members of both sexes. Successful treatment of *T. vaginalis* infection reduced HIV transmission by mucosal routes (20, 22). A Rwandan study showed a difference in the prevalences of trichomoniasis in pregnant women who were infected with HIV or not (20.2% and 10.9%, respectively) (23). Another study, conducted in the United States, demonstrated that *T. vaginalis* contributes to HIV acquisition and exceeds the relative contributions of other STIs (22). Metronidazole used for the treatment of *T. vaginalis* infection significantly decreased the number of HIV (RNA)-free cells (24). Moreover, in HIV-infected women, complications of reproductive tract infection increased significantly with *T. vaginalis* and may decrease if *T. vaginalis* infection is controlled (3, 25).

A study conducted on 43,016 Norwegian women showed that trichomoniasis increased the risk for cervical neoplasia (CN) caused by human papillomavirus (HPV) (26). The same correlation between trichomoniasis and cervical cancer induced by HPV was found during studies conducted in Finland (27) and India (28). Another study showed that *T. vaginalis* infection promoted HPV infection by a factor of 6.5, increasing the risk for CN (29).

Some strains of *T. vaginalis* carry their own viruses that amplify inflammatory

responses (30). Trichomonasvirus released from infected *Trichomonas vaginalis* induced inflammation upon metronidazole treatment (30).

It is well known that infected pregnant women can transmit *T. vaginalis* to their fetuses (2). Unlike other nonviral STIs, trichomoniasis does not primarily reach young women (15 to 25 years of age). It affects women during the reproductive years, and high rates of infection are found in women between the ages of 35 and 40 (31, 32). Predisposing factors comprise older age, use of oral contraceptives, trading sex, smoking, single marital status, and low socioeconomic class (9, 33, 34). The prevalence and average duration of infection depend on the health care-seeking behaviors of populations and their access to health care (34).

### CLINICAL PRESENTATION

While it is usually isolated from the vagina, *T. vaginalis* can also infect the urethra and Skene's gland. The infection, once established, may persist for long periods in women. Asymptomatic *T. vaginalis* infections are well documented; up to 25 to 50% of infected women do not show clinical signs. However, women can also develop symptoms that may be cyclic and often become worse during menstruation. Among women with culture-proven *T. vaginalis* infection, only 11 to 17% present abnormal discharge, odor, pruritus, dysuria, or vaginal burning (35). A "strawberry cervix" is observed in only 2% of women (36).

In healthy adult women, the vaginal pH is around 4. During trichomoniasis, the vaginal pH increases to  $>7$  (37), which is favorable to parasite growth. The fact that trichomoniasis symptoms are worse during menstruation can be explained by changes in pH and hormones. It was proved that the activity of cell detaching factor is inhibited by estrogen (38). Furthermore, menstrual blood creates a rich medium with a high concentration of iron at a higher pH. Consequently, *T. vaginalis* reproduction and attachment to the vaginal epithelium are promoted, resulting in the worsening of symptoms (10).

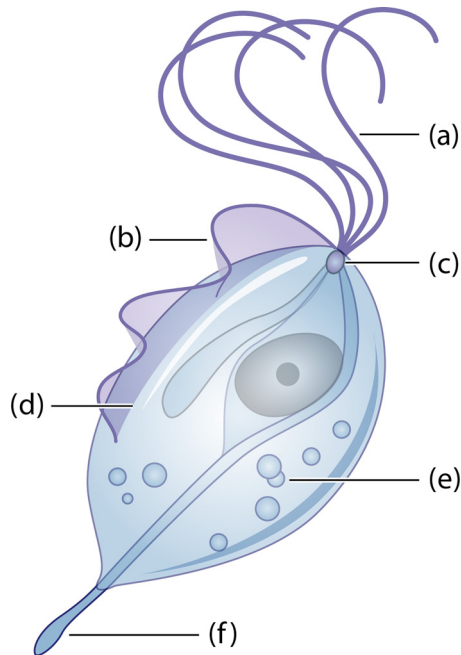
Even if trichomoniasis usually remains localized in the lower part of the urogenital area, it can occasionally provoke adnexitis or pyosalpinx and may potentially have serious sequelae in women, especially during pregnancy.

### BIOLOGY AND PATHOGENESIS OF *TRICHOMONAS VAGINALIS*

*T. vaginalis* is a parasitic protozoan that is typically pyriform, but its appearance is modified under physicochemical conditions (39) (Fig. 1). *T. vaginalis* has five flagella. Four flagella of about 7 to 18  $\mu\text{m}$  are located at its anterior end and give its characteristic twisting and wriggling movement. The fifth flagellum is incorporated within the undulating membrane, whose length is equivalent to half that of the cell, and is supported by a slender, noncontractile costa. An axial axostyle starting at the centrosome is extended by a small posterior tip end (Fig. 1). *T. vaginalis* has a large nucleus characteristic of eukaryotic cells as well as a highly developed Golgi apparatus. As it lacks mitochondria, *T. vaginalis* contains hydrogenosomes as alternative providers of energy (40).

After cytoadherence, the parasite changes its structure to an amoeboid form, which allows an increased contact surface with vaginal epithelial cells followed by adhesion to target cells (for a review, see references 11 and 41). Five *T. vaginalis* attachment proteins (AP), named adhesins, are able to mediate parasite attachment; these are AP23, AP33, AP51, AP65, and AP120. Other surface proteins are also implicated in the attachment of *T. vaginalis*, including fibronectin binding protein and glycolipids, such as lipophosphoglycan (10). Lipophosphoglycan is a pure carbolipid (no peptide component) that, similarly to prokaryotic glycoconjugates, is anchored to the *T. vaginalis* surface via inositol-phosphoceramide.

The immunoinflammatory response to infection has been investigated *in vitro* and in different animals, including murine, bovine, and nonhuman primate models (42–44). *In vitro* experiments have been performed using cervical and vaginal cells and various immune cell types.



**FIG 1** Schematic drawing of *Trichomonas vaginalis*. (a) Anterior flagellum; (b) undulating membrane; (c) pelto; (d) costa; (e) hydrogenosomes; (f) axostyle. The parasite has an average length and width of 9 to 23 and 7  $\mu\text{m}$ , respectively.

The vaginal discharge of infected women contains polymorphonuclear leukocytes. Interaction between *T. vaginalis* and cells triggers an active involvement of signaling pathways. This leads to the production of interleukin-8 (IL-8), IL-6, macrophage chemoattractant protein 1 (MCP-1), and tumor necrosis factor alpha (TNF- $\alpha$ ). Several mitogen-activated protein kinase (MAPK) signaling pathways can be activated, such as c-Jun N-terminal kinase (pJNK), p38, and extracellular signal-regulated kinase 1/2 (ERK1/2) pathways. During *T. vaginalis* infection, the signaling pathways involved are ERK1/2, p38, and NF- $\kappa$ B pathways (45). These signaling pathways also lead to increased mRNA expression of Toll-like receptors (TLRs) (30, 46).

Finally, it appears that MAPKs are also involved in the establishment of cell death in the form of apoptosis by activating Bcl-XL (a Bcl-2-like protein), but not via the Bcl-2 pathway (47), as well as NF- $\kappa$ B in macrophages (46). Implementation of this apoptosis and autophagy pathway may be demonstrated in epithelial cells to decipher more mechanisms related to the pathogenesis of the parasite. An understanding of the host-parasite interaction mechanism is still under investigation by researchers and may lead to the identification of molecular targets on *T. vaginalis* for the design of new trichomonacidal drugs.

## DIAGNOSIS

*T. vaginalis* was identified more than 150 years ago (in 1836), by Donné, when he visualized motile microorganisms in vaginal fluid from women with symptoms of infection. However, the sensitivity of this microscopic visualization technique, also called the wet mount test, is variable, with sensitivities of 38% to 82% among symptomatic women (12). Despite these statistics, the wet mount test is still used in clinical trials for evaluation of drug activity (48). This test must be performed within a few minutes after sample collection to observe viable parasites.

Broth culture of vaginal fluid requires the use of a specialized medium, such as Diamond's or Trichosel medium. Unfortunately, this diagnostic test has different drawbacks: it is limited to laboratories with access to the culture medium and an incubator and has a delay of up to 7 days for *T. vaginalis* identification (49). The culture method was improved by the development of the InPouch device, which is commercially

available. This device consists of a plastic bag with two chambers connected by a narrow passage. The collected specimen is placed in the upper chamber, while the lower chamber is for culture and further observation when necessary. Comparison of the effects of culture medium on sensitivity showed that the InPouch system is at least as sensitive as culture with Diamond's modified medium (50).

PCR-based tests can detect very few trichomonads in a sample, as well as nonviable organisms. The Affirm VP11 test (Becton Dickinson, Sparks, MD) is an amplification test for RNA that allows *T. vaginalis* detection within 30 to 60 min. The specificity and sensitivity of such tests are 99% and 90%, respectively (10).

The OSOM *Trich* rapid antigen test is an immunochromatographic capillary-flow enzyme immunoassay dipstick test that detects *T. vaginalis* membrane protein. Compared to culture, the OSOM *Trich* test has the advantage of being rapid, as the result is available in 10 min. It can be conducted on frozen samples without altering the test characteristics (51).

Nowadays, newer diagnostic options are available (49). Commercially available nucleic acid amplification tests (NAATs) have been validated for use for asymptomatic and symptomatic women and are highly sensitive tests that can be used on multiple specimen types, including urinary, urethral, vaginal, and endocervical specimens (52–54). In a study conducted during 2012 and 2013 in Jefferson County, AL, endocervical, urethral, or urinary specimens from 3,821 women and 2,514 men were collected for *T. vaginalis* detection by use of wet mount detection and a *T. vaginalis* NAAT. The results showed that the *T. vaginalis* NAAT detected infections in women at a rate that was 1.3 times higher than that for wet mount detection (53). In summary, detection of *T. vaginalis* by NAATs, even for asymptomatic patients, should result in better control and treatment of *T. vaginalis* infection (53).

Other NAATs are available, such as the AmpliVue and Solana (Quidel) tests (55). The GenXpert (Gx) assay (Cepheid) (56, 57) is the only NAAT cleared for use for men.

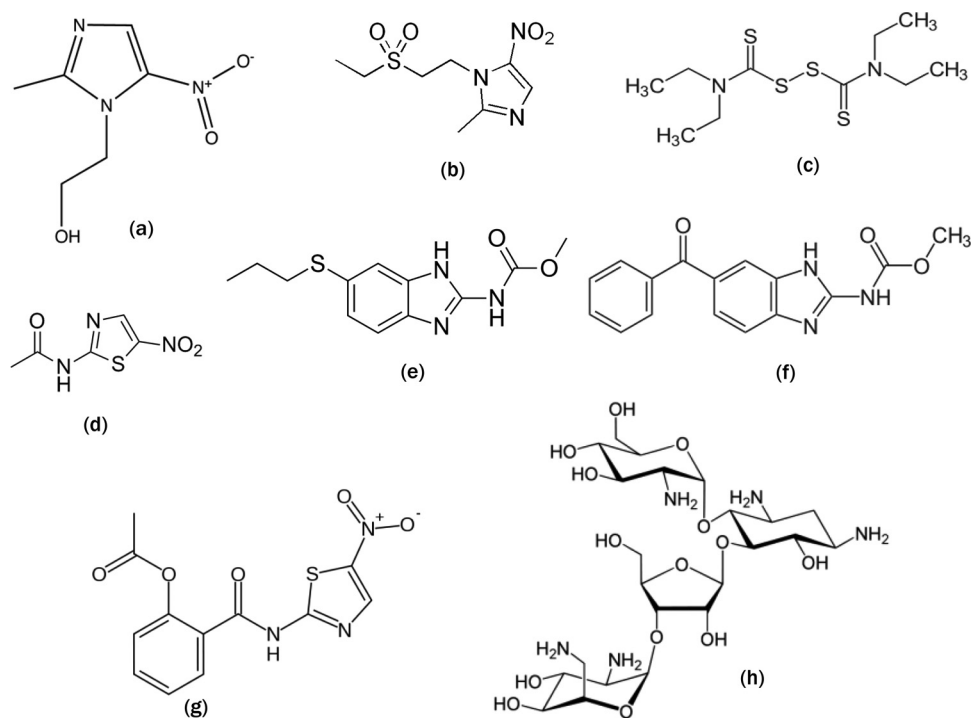
### SYSTEMIC CHEMOTHERAPY OF UROGENITAL TRICHOMONIASIS

Currently, 5-nitroimidazole drugs are commonly used for treatment of trichomoniasis by oral and parenteral routes. Among these drugs, only metronidazole and tinidazole are available in the United States and are authorized by the Food and Drug Administration (FDA) for the treatment of trichomoniasis.

Metronidazole ( $\alpha,\beta$ -hydroxyethyl-2-methyl-5-nitroimidazole; Flagyl) (Fig. 2a) was developed in 1959 and approved in the 1960s for the treatment of trichomoniasis, and it was the only drug with a high cure rate after systemic treatment. Despite the fact that the partners of patients with *T. vaginalis* infection are often infected as well, high rates of asymptomatic infection mean that they do not always seek treatment and may reinfect the partner who was treated. As a consequence, sexual partners of patients should routinely be treated (58, 59).

Metronidazole is relatively cheap, effective, and generally well tolerated. Common side effects, such as gastrointestinal disturbances, are usually mild. Occasional hematologic and neurotoxic side effects have also been reported. Concerning patients with refractory trichomoniasis infection, side effects due to metronidazole become a real problem. In fact, recurrent or resistant trichomoniasis infections are treated for longer periods with increasing doses of metronidazole. However, with higher doses, side effects are much more frequent, leading to patient discomfort and treatment failure (60). In the absence of an alternative to nitroimidazole treatment, cure can be achieved only by increasing dosages of metronidazole. However, side effects may limit the dose of metronidazole and sometimes necessitate stoppage of treatment (61). Cases that cannot be treated with higher doses of metronidazole are a real challenge for both patient and practitioner. Different treatment options with higher doses of oral metronidazole or tinidazole for longer periods have been used together with intravaginal drug delivery (62).

Tinidazole (Fig. 2b) is a nitroimidazole that was introduced in 1969 for the treatment of infections caused by *T. vaginalis*. Tinidazole is curative at lower therapeutic doses



**FIG 2** Chemical structures of anti-*T. vaginalis* drugs. (a) Metronidazole; (b) tinidazole; (c) disulfiram; (d) nithiamide; (e) albendazole; (f) mebendazole; (g) nitazoxanide; (h) paromomycin.

than those for metronidazole and results in fewer and milder side effects (63). Minimum lethal concentrations (MLCs) of tinidazole are lower than those of metronidazole for *T. vaginalis* isolates, and tinidazole resistance is not detected (64).

The mechanism of metronidazole resistance is not totally elucidated, but it is assumed that it may be due to several mutations (4). In anaerobic resistance, the activity of the key enzyme pyruvate:ferredoxin oxidoreductase (PFOR) decreases (65) or disappears in *T. vaginalis*, and the metabolism of the drug is disrupted. In aerobic resistance, transcription of the ferredoxin gene is reduced in resistant *T. vaginalis* strains (66). Aerobic and anaerobic metronidazole resistances have in common a decrease or loss of flavin reductase activity (67).

Although the 5-nitroimidazole compounds are the most effective drugs for treating *T. vaginalis* infection, disulfiram and nithiamide (Fig. 2c and d) might represent alternatives for treating patients with hypersensitivity to 5-nitroimidazole drugs. In another study, the 50% inhibitory concentrations ( $IC_{50}$ s) for albendazole (Fig. 2e) against *T. vaginalis* were 13.2  $\mu\text{g/ml}$  (50  $\mu\text{M}$ ) and 0.899  $\mu\text{g/ml}$  (3.39  $\mu\text{M}$ ) after 4 and 48 h of exposure, respectively (68, 69). Albendazole and mebendazole (Fig. 2f) have so far been found to be effective *in vitro* against *T. vaginalis* (70).

Nitazoxanide (Fig. 2g), a 5-nitrothiazolyl derivative, has shown *in vitro* activity against *T. vaginalis*, with  $IC_{50}$  and  $IC_{90}$  values of 0.034 and 2.046  $\mu\text{g/ml}$ , respectively (71). However, in 2007, there was a failure to cure trichomoniasis with nitazoxanide in three women who presented with nitroimidazole resistance (72).

Clearly, there are only a few data on alternatives to nitroimidazole derivatives, and new antitrichomonal agents or new pharmaceutical formulations are needed to treat infections with resistant *T. vaginalis* organisms.

### PREVENTION OF *T. VAGINALIS* INFECTIONS

Condom use remains the best and most reliable protection against STIs. However, due to religious or cultural reasons, condom use may be limited, particularly in some developing countries. Concurrent treatment of sexual partners is recommended to prevent reinfection. However, systemic administration of chemotherapeutics to prevent



infection results in increased incidences of nitroimidazole-refractory strains. Prevention methods using local intravaginal formulations or vaccines are thus necessary.

Vaccination against *T. vaginalis* is particularly interesting for high-risk individuals to protect themselves and their partners. This strategy would solve many of the issues that currently undermine control efforts. Vaccination against trichomonads is already commercially available against *Tritrichomonas foetus* (TrichGuard; Boehringer Ingelheim Vetmedica). This parasite is a flagellate protozoan similar to *T. vaginalis* that infects cattle. Many recent research works and reviews on vaccination against *T. foetus* have been published (73–75). These studies have shown benefits of vaccinating cattle against trichomoniasis by prevention or clearing of genital infections due to *T. foetus*. Given the similarity between *T. foetus* and *T. vaginalis*, development of a vaccine against human trichomoniasis seems achievable and has the potential for significant social and health impacts. However, the severe economic implications of *T. foetus*, which are estimated to be \$800 to \$6,030 per bull per year in the United States, stimulated research into development of a *T. foetus* vaccine. The calf crop can be reduced by up to 50% in beef enterprises, which partly explains why research has been funded for this bovine vaginal infection and not for the equivalent human infection.

Long-term immune protection is not induced by infection with *T. vaginalis* (76), and the design of an efficacious vaccine still remains a challenge. Due to the limited success of intravaginal vaccination reported in the 1960s with heat-killed *T. vaginalis* and in 1970 with Solco Trichovac, derived from heat-inactivated abnormal strains of lactobacilli, systemic administration of vaccines against *T. vaginalis* was considered. More recently, subcutaneous vaccination of mice reduced the incidence and increased the clearance of *T. vaginalis* infection (77). In the vaccination schedule, mice were vaccinated 56 and 28 days prior to vaginal infection. The vaccine consisted of whole-cell *T. vaginalis*, with aluminum hydroxide used as an adjuvant.

Male circumcision represents another means for the prevention of *T. vaginalis* transmission, since different relevant randomized trials have proven beyond a doubt that partners of circumcised men are less at risk for viral and bacterial infections than those of uncircumcised men (78–80). A clinical study demonstrated that male circumcision reduced *T. vaginalis* infection transmission from male to female partners (81). This study, conducted in South Africa, also demonstrated that woman-to-man transmission of *T. vaginalis* was reduced among circumcised men. In another clinical study, conducted in Rakai, Uganda, Gray and coworkers also observed that trichomonad infection was reduced in women with circumcised partners (82). One possible explanation for this protective effect is that the subpreputial space in uncircumcised men is moist (83), thus enhancing the survival of trichomonads.

However, different conclusions were drawn from a randomized clinical study conducted in Kenya that found no reduction of the risk of acquiring *T. vaginalis* infection (84). No protective effect of circumcision was observed for nonulcerative bacterial STIs (*Neisseria gonorrhoeae* and *Chlamydia trachomatis*) (84). Another study, conducted in Uganda, Zimbabwe, and Thailand, showed that women with circumcised partners had risks of chlamydial, gonococcal, and trichomonal infections similar to those of women with uncircumcised partners (85).

Vaginal administration of microbicide constitutes an alternative to prevent the acquisition of *T. vaginalis* infections. Microbicides are self-administered by women before intercourse, allowing more control over the acquisition of microbial infections. Microbicide administration before intercourse may limit *T. vaginalis* interaction with host cells. In a study conducted by Lushbaugh and coworkers (86), hydrogels composed of hydroxyethylcellulose (3.25% [wt/wt]) containing an antimicrobial peptide (D2A21) (0.5 or 2% [wt/vol]) or metronidazole (500  $\mu\text{g/ml}$ ) were administered to *Lactobacillus*-pretreated estrogenized young mice before inoculation of a *T. vaginalis* suspension. The results showed that the intravaginal metronidazole (500  $\mu\text{g/ml}$ ) gel completely prevented infection in all mice. Three groups of 10 mice were used for each formulation. The peptide D2A21 gel with a peptide concentration of 2% (wt/vol) was significantly more effective than the drug-free hydroxyethylcellulose gel at preventing

*T. vaginalis* infection. Indeed, 90% of mice treated with the peptide at 2% (wt/vol) did not develop infection. Interestingly, even without drug, the hydroxyethylcellulose hydrogel showed a reduction of infection, by a factor of 2. Only 53% of mice treated with hydroxyethylcellulose hydrogel developed the infection (86).

### VAGINALLY ADMINISTERED FORMULATIONS FOR TREATMENT OF *T. VAGINALIS* INFECTIONS

The major limitation related to vaginal administration of drugs for the treatment of trichomoniasis is the nonaccessibility of other infected organs (cervix, bladder, and Bartholin's, Skene's, and periurethral glands). Despite this limitation, vaginal administration of drugs represents an interesting alternative to systemic administration (i) in cases of nitroimidazole allergy, (ii) in cases of pregnancy, (iii) when desensitization is not possible, (iv) when other systemic treatment options are limited, or (v) when severe side effects due to systemic administration are observed.

#### The Vagina as a Site for Drug Administration

The vagina is composed of 26 to 29 layers of epithelial cells, depending on age and the stage of the menstrual cycle (87, 88). The state of the epithelium is highly dependent on hormonal activity. In the first part of the cycle, the epithelium has a proliferative activity accompanied by significant glycogen synthesis (89). After ovulation, glycogen synthesis slows down, the cells desquamate, and the epithelial thickness decreases.

The vaginal mucosa is lined with mucus, which is a viscoelastic gel composed of organic and inorganic salts, mucin proteins (including immunoglobulins), carbohydrates, urea, and fatty acids. Mucus protects the vaginal mucosa from the external environment and ensures lubrication. Its composition and physical characteristics vary according to the menstrual period. It is capable of capturing foreign particles and removing them, but in contrast, some particles can be directed to the uterus. This is the case, for example, for sperm during ovulation.

Vaginal fluid is composed of secretions and transudations from blood vessels, cells derived from desquamation of the vaginal epithelium, leukocytes, and secretions of the endometrium and fallopian tubes. The amount and composition of vaginal fluid change during the menstrual cycle. The amount of vaginal fluid is estimated to be between approximately 0.5 and 0.75 g (90, 91).

The normal vaginal pH is acidic (approximately 4 to 4.5) and usually varies between 3.5 and 5. This value is maintained by the resident microbiota and by lactobacilli (in particular *Lactobacillus* spp. in the vagina, also called Döderlein flora) that convert glycogen from exfoliated epithelial cells into lactic acid and produce other fatty acids. Lactobacilli also produce bactericidal substances, such as hydrogen peroxide and surfactants, that play a role in preventing infections (92). Menstrual secretions, cervical mucus, and sperm can raise the vaginal pH (93).

The vaginal route has many advantages for drug delivery. It allows drugs to avoid the hepatic primary channel, as demonstrated with propranolol (a sympatholytic nonselective beta blocker), which has a better bioavailability after administration by the vaginal route than after oral administration (94). There is also a decrease in observed side effects by vaginal administration of bromocriptine, a dopamine agonist, compared to those with oral administration (95). Hepatic side effects induced by hormone replacement therapy or birth control are also greatly reduced (96).

Pharmaceutical formulations for vaginal administration are various and include liquid solutions, emulsions, suspensions, and solids, such as pessaries, vaginal tablets, vaginal capsules, and vaginal films. There are also other pharmaceutical formulations that are semisolid, including creams, ointments, and gels. Vaginal rings, unlike semisolid formulations used to line the vaginal mucosa, are positioned at a precise place in the upper third of the vagina (which is particularly susceptible to pathogen infection), near the cervicovaginal junction. They may liberate drug in a controlled way and during a long period (up to several months) in the lumen of the vagina. Vaginal rings are



generally constituted of a polymeric matrix, such as silicone, and thermoplastic materials containing drugs.

### Vaginally Applied Formulations for Treatment of *T. vaginalis* Infections

In 1956, povidone-iodine (Betadine) was introduced as an antiseptic agent. It was shown to be an active agent against *T. vaginalis* (97–101). However, other studies observed treatment failure (102). The antiprotozoal action of povidone-iodine is dependent on the release of iodine. Thus, a 2-min douche with povidone-iodine is less effective than a 10-min douche (103). The use of povidone-iodine is counterindicated for pregnant women because of neonatal hypothyroidism reported after maternal use of povidone-iodine in pregnancy.

The use of intravaginal nonoxynol-9 administration was considered at the beginning of the 1990s for the treatment of trichomoniasis (104), but it was abandoned because of toxicity revealed during clinical trials of anti-HIV-1 microbicides (105–107). Studies have shown that nonoxynol-9 results in rupture of the vaginal epithelial barrier and accelerates HIV replication (108).

Nitroimidazoles for *T. vaginalis* treatment have been formulated for vaginal application. Metronidazole (500 mg) vaginal tablets (Tergynan) or ovules (Flagyl) are commonly used for the treatment of vaginal trichomoniasis. The classical treatment schedule consists of one application per day for 10 days (109–111).

The anti-*T. vaginalis* activity of vaginal tablets containing a lower dosage of metronidazole (100 mg) was compared to those of vaginal tablets containing placebo and 7-day oral metronidazole (500 mg twice a day). Significant differences in cure rates were observed between the placebo group and the two metronidazole groups. Oral and bioadhesive treatments did not lead to significant differences in clinical efficacy (112). Furthermore, a cure rate of 64% was obtained after administration of vaginal tablets containing 100 mg of metronidazole (113).

A pilot study compared the efficacies of 7-day treatment with oral metronidazole tablets (250 mg; three times daily) and vaginal gel treatment (0.75%; twice daily) (114). The 5-g dose of hydrogel contained only 37.5 mg of active drug. At the end of the treatments, significant reductions of genitourinary symptoms were observed with both vaginal hydrogel and tablets. However, the wet mount test showed that vaginal metronidazole administration failed to treat trichomoniasis compared to treatment with oral metronidazole. It is noteworthy that the composition of the hydrogel used in this study is unknown. Hydrogel properties can affect the distribution of the drug on the vaginal mucosa and, in turn, the anti-*T. vaginalis* activity. The cure rate was 44% with metronidazole gel, which is comparable to rates reported in previous studies of intravaginal metronidazole administration for the treatment of *T. vaginalis* infections, which ranged from 30 to 60% (109, 111). Although it failed to completely cure the infection, metronidazole gel was suitable for reducing side effects due to systemic passage. In order to facilitate treatment of resistant trichomoniasis, one strategy consisted of concomitant vaginal and oral administration of metronidazole. The success of combined oral and vaginal therapy has been known since the 1960s. A study conducted on 2,002 incarcerated women in California over 36 months compared the efficiencies of oral metronidazole (250 mg; three times daily for 3, 5, 7, or 10 days), vaginal metronidazole (500-mg vaginal inserts; once daily for 7 days), and a combination of oral and vaginal therapies (250-mg oral tablets given 3 times a day and concurrent 250-mg vaginal tablets for 5 days) (109). The results showed that the combined oral and vaginal 5-day treatment had the highest activity against *T. vaginalis* infections (109).

Combination therapy of metronidazole with other drugs is a good alternative strategy to administration of metronidazole alone. Vaginal ovules or cream containing the same metronidazole dosage (500 mg) combined with nystatin (100,000 IU) (Flagystatin) was used to treat mixed vaginal infections due to *T. vaginalis* and *Candida albicans*. Metronidazole has also been combined with other antimicrobial drugs, such as miconazole (115).

Alternative treatments include intravaginal preparations of paromomycin cream (116–118) (Fig. 2h). In 1964, paromomycin was used to treat trichomoniasis, with cure achieved in 85% of patients who received the drug topically as a vaginal pessary (119).

A case of trichomoniasis that was particularly resistant to metronidazole was successfully treated with intravaginal application of paromomycin (116). That study showed that the patient failed to respond to high-dose oral and topical metronidazole. The highest dose of metronidazole used was 7 days of oral tablets (800 mg) given three times daily, with 1 g intravaginal metronidazole given nightly. Over the next few months, the patient was treated without success with oral tinidazole, nimorazole, mebendazole, intravaginal clotrimazole, povidone-iodine, Aci-Jel, nonoxynol-9 pessaries, and hydrogen peroxide douches. This was followed by a full course of inactivated lactobacillus vaccine (Solco Trichovac) and a booster 1 year later. The patient continued to be symptomatic, and *T. vaginalis* was isolated repeatedly throughout this period. The initial MIC showed that the organism was resistant to metronidazole (116). Complete cure was achieved with 250 mg of paromomycin administered vaginally for 5 days.

In a study conducted on nine patients infected with *T. vaginalis*, among whom four patients had strains that were metronidazole resistant and five patients were allergic to metronidazole, treatment with paromomycin cream (250 mg per 4-g applicator nightly) was achieved for 2 weeks (117), and six of nine women were cured.

However, in a case study reported by Muzny and colleagues (102), intravaginal paromomycin failed to treat a patient with symptomatic *T. vaginalis* infection. This patient developed hypersensitivity to nitroimidazole drugs. Complete symptomatic cure was observed after vaginal pH acidification by intravaginal administration of boric acid for 2 months (102). Two case studies previously showed that vaginal acidification with intravaginal boric acid, for 1 and 5 months in two patients who tolerated this therapy, resulted in the treatment of recalcitrant *T. vaginalis* (120).

In a more recent study, a case report demonstrated that drug-resistant trichomoniasis can be treated successfully by concomitant administration of tinidazole (orally) and paromomycin cream nightly (121). However, frequent local vulvovaginal adverse reactions were reported for patients treated with paromomycin (62, 116, 118, 121). The side effects, such as ulcers of the vulvar and vaginal mucosal surfaces and vulvar pain, could be so severe as to require interruption of treatment (116).

During pregnancy, a daily dose of 100 mg of clotrimazole can be delivered intravaginally at bedtime for 14 days and can provide temporary relief to patients in the first trimester. With this treatment schedule, a cure rate of 50% was obtained (122).

To sum up, topically applied treatments are generally limited to adjunctive therapy or particular cases of allergy or resistance. Nevertheless, some data have shown that treatment of trichomoniasis by the vaginal route can be a real alternative to systemic treatment. All the described strategies for local delivery of anti-*T. vaginalis* formulations are based on the inclusion of a drug in a vehicle. Generally, little attention is paid to improving the formulation vehicle residence time on the mucosa infected with *T. vaginalis*.

### **General Considerations for Designing Vaginally Applied Anti-*T. vaginalis* Formulations**

Efficacious vaginally applied formulations must have different properties, such as stability in acidic medium, adhesion, nonliquefaction at body temperature, slow dissolution, lubricant properties, and nongreasiness (123). Moreover, when formulations are intended for the vagina, other elements should also be taken into account. In fact, the formulations should (i) be stable at the acidic vaginal pH, (ii) be easily applied to obtain a homogeneous distribution of the drug, (iii) be retained in the vagina as long as possible, (iv) be compatible with other coadministered substances, and (v) be nontoxic to the vaginal mucosa (124). Vaginally applied formulations can also be used as controlled-release devices for vaginally applied drugs.

pH plays a role in the amount of drug absorbed from the vaginal mucosa and must be considered in the formulation of drug delivery systems. An increase of the absorption of leuprolide (a gonadotropin-releasing hormone analog) was observed after adding organic acids which partially dissolved the cellular cement. Furthermore, the efficacy of a gel containing dinoprostone (prostaglandin E2) was significantly related to the pH of the vagina.

The apparent aqueous solubility of anti-*T. vaginalis* drugs (i.e., albendazole or clotrimazole) represents another parameter to be considered for their formulation, as it determines the choice of formulation (solid, semisolid, liquid, solution, suspension, etc.). The apparent aqueous solubility of albendazole was increased 7,600 times by using randomly methylated  $\beta$ -cyclodextrin (Rameb Me- $\beta$ -CD) at a concentration of 40% (wt/wt) (125), while the solubility of clotrimazole was improved 9,980 times by using the same cyclodextrin (126). In another study, chitosan nanoparticles exhibited strong anti-*T. vaginalis* activity *in vitro* even without addition of drugs (127).

Semisolid drug delivery forms, such as gels, are widely used in the development of topical formulations against vaginal microbial infections (128). Gels are semisolid substances generally constituted of a polymeric matrix. Pharmaceutical gels consist of natural polymers, in particular some proteins (collagen and gelatin) and polysaccharides (alginate, carrageenan, and guar gum), semisynthetic polymers (carboxymethyl cellulose and hydroxypropyl methylcellulose), and other synthetic (carbomer and Pluronic) or inorganic (aluminum hydroxide, bentonite, and laponite) substances (123). Hydrogels have many advantages: they are generally colorless, odorless, and tasteless and can be used in combination with vaginal devices or condoms (129).

In terms of their effectiveness, the most important problems presented by these forms are their low remanence at the epithelial surface and their rapid detachment from the application site (129, 130). In order to overcome these problems, mucoadhesive polymers are often added to formulations to immobilize the gel as long as possible on the vaginal mucosa (131).

For all these reasons, the use of thermosensitive and mucoadhesive gels was recently presented as an alternative strategy for treatment of *T. vaginalis* infection (127, 132, 133). These gels are liquid at low temperature, thus facilitating the spreading of the formulation on the whole vaginal area, even in difficult-to-access places, and they become semisolid at body temperature, increasing the residence time of the formulation on the mucosa (134–136).

## CONCLUSIONS

The progressive abandoning of condom use relative to discomfort and linked to the forgetting of HIV risk and the increase of poverty worldwide may partially explain the increase in the annual number of urogenital trichomoniasis cases. In addition, drug resistance emergence and intolerance to nitroimidazoles contribute to making trichomoniasis treatment a societal challenge to be addressed. Because of this, chemotherapy and vaccines are the best ways to control the expansion of this cosmopolitan disease. Besides *in vitro* screening of new compounds, all strategies that attempt to improve the biodistribution of anti-*Trichomonas* compounds provide real added value to the fight against this disease. In particular, approaches consisting of the prevention of side effects linked to parenteral treatments need to be prioritized. Thus, any investigation with the aim of developing local treatments is promising. In this regard, advances in mucoadhesive polymer formulation should particularly be supported in the future.

## REFERENCES

1. Wendel KA, Erbeling EJ, Gaydos CA, Rompalo AM. 2003. Use of urine polymerase chain reaction to define the prevalence and clinical presentation of *Trichomonas vaginalis* in men attending an STD clinic. *Sex Transm Infect* 79:151–153. <https://doi.org/10.1136/sti.79.2.151>.
2. Carlier Y, Truysen C, Deloron P, Peyron F. 2012. Congenital parasitic infections: a review. *Acta Trop* 121:55–70. <https://doi.org/10.1016/j.actatropica.2011.10.018>.
3. Fox J, Fidler S. 2010. Sexual transmission of HIV-1. *Antiviral Res* 85: 276–285. <https://doi.org/10.1016/j.antiviral.2009.10.012>.
4. Paulish-Miller TE, Augostini P, Schuyler JA, Smith WL, Mordechai E,

- Adelson ME, Gyax SE, Secor WE, Hilbert DW. 2014. *Trichomonas vaginalis* metronidazole resistance is associated with single nucleotide polymorphisms in the nitroreductase genes ntr4Tv and ntr6Tv. *Antimicrob Agents Chemother* 58:2938–2943. <https://doi.org/10.1128/AAC.02370-13>.
5. Petrin D, Delgaty K, Bhatt R, Garber G. 1998. Clinical and microbiological aspects of *Trichomonas vaginalis*. *Clin Microbiol Rev* 11:300–317.
6. Schwebke JR, Burgess D. 2004. Trichomoniasis. *Clin Microbiol Rev* 17:794–803. <https://doi.org/10.1128/CMR.17.4.794-803.2004>.
7. Soper D. 2004. Trichomoniasis: under control or undercontrolled? *Am J Obstet Gynecol* 190:281–290. <https://doi.org/10.1016/j.ajog.2003.08.023>.
8. Bachmann LH, Hobbs MM, Seña AC, Sobel JD, Schwebke JR, Krieger JN, McClelland RS, Workowski KA. 2011. *Trichomonas vaginalis* genital infections: progress and challenges. *Clin Infect Dis* 53:S160–S172. <https://doi.org/10.1093/cid/cir705>.
9. Nanda N, Michel RG, Kurdgelashvili G, Wendel KA. 2006. Trichomoniasis and its treatment. *Expert Rev Anti Infect Ther* 4:125–135. <https://doi.org/10.1586/14787210.4.1.125>.
10. Harp DF, Chowdhury I. 2011. Trichomoniasis: evaluation to execution. *Eur J Obstet Gynecol Reprod Biol* 157:3–9. <https://doi.org/10.1016/j.ejogrb.2011.02.024>.
11. Mielczarek E, Blaszkowska J. 2016. *Trichomonas vaginalis*: pathogenicity and potential role in human reproductive failure. *Infection* 44:447–458. <https://doi.org/10.1007/s15010-015-0860-0>.
12. Bhesania AH, Narayankhedkar A. 2016. Trichomoniasis—a review. *Int J Curr Microbiol Appl Sci* 5:731–741.
13. Kissinger P. 2015. *Trichomonas vaginalis*: a review of epidemiologic, clinical and treatment issues. *BMC Infect Dis* 15:307. <https://doi.org/10.1186/s12879-015-1055-0>.
14. Edwards T, Burke P, Smalley H, Hobbs G. 2016. *Trichomonas vaginalis*: clinical relevance, pathogenicity and diagnosis. *Crit Rev Microbiol* 42: 406–417. <https://doi.org/10.3109/1040841X.2015.1105782>.
15. Field N, Clifton S, Alexander S, Ison CA, Khanom R, Saunders P, Hughes G, Heath L, Beddows S, Mercer CH, Tanton C, Johnson AM, Sonnenberg P. 2016. *Trichomonas vaginalis* infection is uncommon in the British general population: implications for clinical testing and public health screening. *Sex Transm Infect* 2016:sextrans-2016-052660. <https://doi.org/10.1136/sextrans-2016-052660>.
16. Pereyre S, Nadalié CL, Bébéar C. 2016. *Mycoplasma genitalium* and *Trichomonas vaginalis* in France: a point prevalence study in people screened for sexually transmitted diseases. *Clin Microbiol Infect* 23: 122.e1–122.e7. <https://doi.org/10.1016/j.cmi.2016.10.028>.
17. Matini M, Rezaie S, Mohebbi M, Maghsood AH, Rabiee S, Fallah M, Rezaeian M. 2012. Prevalence of *Trichomonas vaginalis* infection in Hamadan City, Western Iran. *Iran J Parasitol* 7:67–72.
18. Buve A, Weiss HA, Laga M, Van Dyck E, Musonda R, Zekeng L, Kahindo M, Anagonou S, Morison L, Robinson NJ. 2001. The epidemiology of trichomoniasis in women in four African cities. *AIDS* 15:S89–S96. <https://doi.org/10.1097/00002030-200108004-00010>.
19. Moodley P, Connolly C, Sturm AW. 2002. Interrelationships among human immunodeficiency virus type 1 infection, bacterial vaginosis, trichomoniasis, and the presence of yeasts. *J Infect Dis* 185:69–73. <https://doi.org/10.1086/338027>.
20. Sorvillo F, Kerndt P. 1998. *Trichomonas vaginalis* and amplification of HIV-1 transmission. *Lancet* 351:213–214. [https://doi.org/10.1016/S0140-6736\(05\)78181-2](https://doi.org/10.1016/S0140-6736(05)78181-2).
21. Laga MA, Manoka A, Kivuvu M, Malele B, Tuliza M, Nzila N, Goeman J, Behets F, Batter V, Alary M, Heyward WL, Ryder RW, Piot P. 1993. Non-ulcerative sexually transmitted diseases as risk factors for HIV-1 transmission in women: results from a cohort study. *AIDS* 7:95–102. <https://doi.org/10.1097/00002030-199301000-00015>.
22. Sorvillo F, Smith L, Kerndt P, Ash L. 2001. *Trichomonas vaginalis*, HIV, and African-Americans. *Emerg Infect Dis* 7:927–932. <https://doi.org/10.3201/eid0706.010603>.
23. Leroy V, De Clercq A, Ladner J, Bogaerts J, Van de Perre P, Dabis F. 1995. Should screening of genital infections be part of antenatal care in areas of high HIV prevalence? A prospective cohort study from Kigali, Rwanda, 1992–1993. The Pregnancy and HIV (EGE) Group. *Genitourin Med* 71:207–211. <https://doi.org/10.1136/sti.71.4.207>.
24. Wang CC, McClelland RS, Reilly M, Overbaugh J, Emery SR, Mandaliya K, Chohan B, Ndinya-Achola J, Bwayo J, Kreiss JK. 2001. The effect of treatment of vaginal infections on shedding of human immunodeficiency virus type 1. *J Infect Dis* 183:1017–1022. <https://doi.org/10.1086/319287>.
25. Moodley P, Wilkinson D, Connolly C, Moodley J, Sturm AW. 2002. *Trichomonas vaginalis* is associated with pelvic inflammatory disease in women infected with human immunodeficiency virus. *Clin Infect Dis* 34:519–522. <https://doi.org/10.1086/338399>.
26. Gram IT, Macaluso M, Churchill J, Stalsberg H. 1992. *Trichomonas vaginalis* (TV) and human papillomavirus (HPV) infection and the incidence of cervical intraepithelial neoplasia (CIN) grade III. *Cancer Causes Control* 3:231–236. <https://doi.org/10.1007/BF00124256>.
27. Viikki M. 2000. Gynaecological infections as risk determinants of subsequent cervical neoplasia. *Acta Oncol* 39:71–75. <https://doi.org/10.1080/028418600431003>.
28. Ghosh I, Muwonge R, Mittal S, Banerjee D, Kundu P, Mandal R, Biswas J, Basu P. 2017. Association between high risk human papillomavirus infection and co-infection with *Candida* spp. and *Trichomonas vaginalis* in women with cervical premalignant and malignant lesions. *J Clin Virol* 87:43–48. <https://doi.org/10.1016/j.jcv.2016.12.007>.
29. Lazenby GB, Taylor PT, Badman BS, Mchaki E, Korte JE, Soper DE, Pierce JY. 2014. An association between *Trichomonas vaginalis* and high-risk human papillomavirus in rural Tanzanian women undergoing cervical cancer screening. *Clin Ther* 36:38–45. <https://doi.org/10.1016/j.clinthera.2013.11.009>.
30. Fichorova RN, Lee Y, Yamamoto HS, Takagi Y, Hayes GR, Goodman RP, Chepa-Lotrea X, Buck OR, Murray R, Kula T, Beach DH, Singh BN, Nibert ML. 2012. Endobiont viruses sensed by the human host—beyond conventional antiparasitic therapy. *PLoS One* 7:e48418. <https://doi.org/10.1371/journal.pone.0048418>.
31. Bowden FJ, Paterson BA, Mein J, Savage J, Fairley CK, Garland SM, Tabrizi SN. 1999. Estimating the prevalence of *Trichomonas vaginalis*, *Chlamydia trachomatis*, *Neisseria gonorrhoeae*, and human papillomavirus infection in indigenous women in northern Australia. *Sex Transm Infect* 75:431–434. <https://doi.org/10.1136/sti.75.6.431>.
32. Cudmore SL, Delgaty KL, Hayward-McClelland SF, Petrin DP, Garber GE. 2004. Treatment of infections caused by metronidazole-resistant *Trichomonas vaginalis*. *Clin Microbiol Rev* 17:783–793. <https://doi.org/10.1128/CMR.17.4.783-793.2004>.
33. Lewis DA. 2010. Trichomoniasis. *Medicine* 38:291–293. <https://doi.org/10.1016/j.mpmed.2010.03.007>.
34. Riley ED, Cohen J, Dilworth SE, Grimes B, Marquez C, Chin-Hong P, Philip SS. 2016. *Trichomonas vaginalis* infection among homeless and unstably housed adult women living in a resource-rich urban environment. *Sex Transm Infect* 92:305–308. <https://doi.org/10.1136/sextrans-2015-052143>.
35. Landers DV, Wiesenfeld HC, Heine RP, Krohn MA, Hillier SL. 2004. Predictive value of the clinical diagnosis of lower genital tract infection in women. *Am J Obstet Gynecol* 190:1004–1008. <https://doi.org/10.1016/j.ajog.2004.02.015>.
36. Fouts AC, Kraus SJ. 1980. *Trichomonas vaginalis*: reevaluation of its clinical presentation and laboratory diagnosis. *J Infect Dis* 141:137–143. <https://doi.org/10.1093/infdis/141.2.137>.
37. Anorlu RI, Fagbenro Beyioku AF, Fagorala T, Abudu OO, Galadanci HS. 2001. Prevalence of *Trichomonas vaginalis* in patients with vaginal discharge in Lagos, Nigeria. *Niger Postgrad Med J* 8:183–186.
38. Garber GE, Lemchuk-Favel LT, Rousseau G. 1991. Effect of beta-estradiol on production of the cell-detaching factor of *Trichomonas vaginalis*. *J Clin Microbiol* 29:1847–1849.
39. Pereira-Neves A, Ribeiro KC, Benchimol M. 2003. Pseudocysts in trichomonads—new insights. *Protist* 154:313–329. <https://doi.org/10.1078/143446103322454095>.
40. Hrdy I, Hirt RP, Dolezal P, Bardónová L, Foster PG, Tachezy J, Embley TM. 2004. *Trichomonas* hydrogenosomes contain the NADH dehydrogenase module of mitochondrial complex I. *Nature* 432:618–622. <https://doi.org/10.1038/nature03149>.
41. Benchimol M, Pereira-Neves A, de Souza W. 2015. Pathogenesis of *Trichomonas vaginalis* in humans, p 423–439. In Singh SK (ed), *Human emerging and re-emerging infections, vol I. Viral and parasitic infections*. John Wiley & Sons, Inc, Hoboken, NJ.
42. Cobo ER, Corbeil LB, Gershwin LJ, BonDurant RH. 2009. Preputial cellular and antibody responses of bulls vaccinated and/or challenged with *Trichomonas foetus*. *Vaccine* 28:361–370. <https://doi.org/10.1016/j.vaccine.2009.10.039>.
43. Kulda J. 1990. Employment of experimental animals in studies of *Trichomonas vaginalis* infection, p 112–154. In Honigberg BM (ed),



- Trichomonads parasitic in humans. Springer, New York, NY. [https://doi.org/10.1007/978-1-4612-3224-7\\_8](https://doi.org/10.1007/978-1-4612-3224-7_8).
44. Patton DL, Cosgrove Sweeney YT, Agnew KJ, Balkus JE, Rabe LK, Hillier SL. 2006. Development of a nonhuman primate model for *Trichomonas vaginalis* infection. *Sex Transm Dis* 33:743–746. <https://doi.org/10.1097/01.olq.0000218871.89901.61>.
  45. Singh BN, Hayes GR, Lucas JJ, Sommer U, Viseux N, Mirgorodskaya E, Trifonova RT, Sassi RRS, Costello CE, Fichorova RN. 2009. Structural details and composition of *Trichomonas vaginalis* lipophosphoglycan in relevance to the epithelial immune function. *Glycoconj J* 26:3–17. <https://doi.org/10.1007/s10719-008-9157-1>.
  46. Chang J-O, Park J-Y, Kim S-K. 2006. Dependence on p38 MAPK signaling in the up-regulation of TLR2, TLR4 and TLR9 gene expression in *Trichomonas vaginalis*-treated HeLa cells. *Immunology* 118:164–170. <https://doi.org/10.1111/j.1365-2567.2006.02347.x>.
  47. Chang J-H, Ryang Y-S, Morio T, Lee S-K, Chang E-J. 2004. *Trichomonas vaginalis* inhibits proinflammatory cytokine production in macrophages by suppressing NF-kappaB activation. *Mol Cells* 18:177–185.
  48. Schwebke JR, Lensing SY, Sobel J. 2013. Intravaginal metronidazole/miconazole for the treatment of vaginal trichomoniasis. *Sex Transm Dis* 40:710–714. <https://doi.org/10.1097/01.olq.0000431069.38601.d5>.
  49. Van Der Pol B. 2016. Clinical and laboratory testing for *Trichomonas vaginalis* infection. *J Clin Microbiol* 54:7–12. <https://doi.org/10.1128/JCM.02025-15>.
  50. Sood S, Mohanty S, Kapil A, Tolosa J, Mittal S. 2007. InPouch TV (TM) culture for detection of *Trichomonas vaginalis*. *Indian J Med Res* 125: 567.
  51. Huppert JS, Batteiger BE, Braslins P, Feldman JA, Hobbs MM, Sankey HZ, Sena AC, Wendel KA. 2005. Use of an immunochromatographic assay for rapid detection of *Trichomonas vaginalis* in vaginal specimens. *J Clin Microbiol* 43:684–687. <https://doi.org/10.1128/JCM.43.2.684-687.2005>.
  52. Schwebke JR, Hobbs MM, Taylor SN, Sena AC, Catania MG, Weinbaum BS, Johnson AD, Getman DK, Gaydos CA. 2011. Molecular testing for *Trichomonas vaginalis* in women: results from a prospective U.S. clinical trial. *J Clin Microbiol* 49:4106–4111. <https://doi.org/10.1128/JCM.01291-11>.
  53. Muzny CA, Blackburn RJ, Sinsky RJ, Austin EL, Schwebke JR. 2014. Added benefit of nucleic acid amplification testing for the diagnosis of *Trichomonas vaginalis* among men and women attending a sexually transmitted diseases clinic. *Clin Infect Dis* 59:834–841. <https://doi.org/10.1093/cid/ciu446>.
  54. Van Der Pol B, Williams JA, Taylor SN, Cammarata CL, Rivers CA, Body BA, Nye M, Fuller D, Schwebke JR, Barnes M, Gaydos CA. 2014. Detection of *Trichomonas vaginalis* DNA by use of self-obtained vaginal swabs with the BD ProbeTec Qx assay on the BD Viper system. *J Clin Microbiol* 52:885–889. <https://doi.org/10.1128/JCM.02966-13>.
  55. Gaydos CA, Schwebke J, Dombrowski J, Marrazzo J, Coleman J, Silver B, Barnes M, Crane L, Fine P. 2017. Clinical performance of the Solana™ Point-of-Care *Trichomonas* assay from clinician-collected vaginal swabs and urine specimens from symptomatic and asymptomatic women. *Expert Rev Mol Diagn* 17:303–306. <https://doi.org/10.1080/14737159.2017.1282823>.
  56. Badman SG, Causer LM, Guy R, Tabrizi SN, Francis F, Donovan B, Whitley D. 2016. A preliminary evaluation of a new GeneXpert (Gx) molecular point-of-care test for the detection of *Trichomonas vaginalis*. *Sex Transm Infect* 92:350–352. <https://doi.org/10.1136/sextrans-2015-052384>.
  57. Dize L, Agreda P, Quinn N, Barnes MR, Hsieh YH, Gaydos CA. 2013. Comparison of self-obtained penile-meatal swabs to urine for the detection of *C. trachomatis*, *N. gonorrhoeae* and *T. vaginalis*. *Sex Transm Infect* 89:305–307. <https://doi.org/10.1136/sextrans-2012-050686>.
  58. Kissinger P, Schmidt N, Mohammed H, Leichter JS, Gift TL, Meadors B, Sanders C, Farley TA. 2006. Patient-delivered partner treatment for *Trichomonas vaginalis* infection: a randomized controlled trial. *Sex Transm Dis* 33:445–450. <https://doi.org/10.1097/01.olq.0000204511.84485.4c>.
  59. Workowski KA, Berman SM. 2006. Sexually transmitted diseases treatment guidelines, 2006. *MMWR Recomm Rep* 55(RR-11):1–94.
  60. Howe K, Kissinger PJ. 2017. Single-dose compared with multidose metronidazole for the treatment of trichomoniasis in women: a meta-analysis. *Sex Transm Dis* 44:30–35. <https://doi.org/10.1097/OLQ.0000000000000537>.
  61. Cudmore SL, Garber GE. 2010. Prevention or treatment: the benefits of *Trichomonas vaginalis* vaccine. *J Infect Public Health* 3:47–53. <https://doi.org/10.1016/j.jiph.2010.01.003>.
  62. Sobel JD, Nyirjesy P, Brown W. 2001. Tinidazole therapy for metronidazole-resistant vaginal trichomoniasis. *Clin Infect Dis* 33: 1341–1346. <https://doi.org/10.1086/323034>.
  63. Raja IM, Basavareddy A, Mukherjee D, Meher BR. 2016. Randomized, double-blind, comparative study of oral metronidazole and tinidazole in treatment of bacterial vaginosis. *Indian J Pharmacol* 48:654. <https://doi.org/10.4103/0253-7613.194843>.
  64. Kirkcaldy RD, Augostini P, Asbel LE, Bernstein KT, Kerani RP, Mettenbrink CJ, Pathela P, Scwebke JR, Secor WE, Workowski KA, Davis D, Braxton J, Weinstock HS. 2012. *Trichomonas vaginalis* antimicrobial drug resistance in 6 US cities, STD surveillance network, 2009–2010. *Emerg Infect Dis* 18:939–943. <https://doi.org/10.3201/eid1806.111590>.
  65. Kulda J. 1999. Trichomonads, hydrogenosomes and drug resistance. *Int J Parasitol* 29:199–212. [https://doi.org/10.1016/S0020-7519\(98\)00155-6](https://doi.org/10.1016/S0020-7519(98)00155-6).
  66. Yarlett N, Yarlett NC, Lloyd D. 1986. Metronidazole-resistant clinical isolates of *Trichomonas vaginalis* have lowered oxygen affinities. *Mol Biochem Parasitol* 19:111–116. [https://doi.org/10.1016/0166-6851\(86\)90115-5](https://doi.org/10.1016/0166-6851(86)90115-5).
  67. Leitsch D, Janssen BD, Kolarich D, Johnson PJ, Duchêne M. 2014. *Trichomonas vaginalis* flavin reductase 1 and its role in metronidazole resistance. *Mol Microbiol* 91:198–208. <https://doi.org/10.1111/mmi.12455>.
  68. Navarrete-Vázquez G, Yépez L, Hernández-Campos A, Tapia A, Hernández-Luis F, Cedillo R, González J, Martínez-Fernández A, Martínez-Grueiro M, Castillo R. 2003. Synthesis and antiparasitic activity of albendazole and mebendazole analogues. *Bioorg Med Chem* 11: 4615–4622. [https://doi.org/10.1016/S0968-0896\(03\)00497-8](https://doi.org/10.1016/S0968-0896(03)00497-8).
  69. Oxberry ME, Thompson RCA, Reynoldson J. 1994. Evaluation of the effects of albendazole and metronidazole on the ultrastructure of *Giardia duodenalis*, *Trichomonas vaginalis* and *Spironucleus muris* using transmission electron microscopy. *Int J Parasitol* 24:695–703. [https://doi.org/10.1016/0020-7519\(94\)90123-6](https://doi.org/10.1016/0020-7519(94)90123-6).
  70. Sears SD, O'Hare J. 1988. *In vitro* susceptibility of *Trichomonas vaginalis* to 50 antimicrobial agents. *Antimicrob Agents Chemother* 32:144–146. <https://doi.org/10.1128/AAC.32.1.144>.
  71. Cedillo-Rivera R, Chávez B, González-Robles A, Tapia A, Yépez-Mulia L. 2002. *In vitro* effect of nitazoxanide against *Entamoeba histolytica*, *Giardia intestinalis* and *Trichomonas vaginalis* trophozoites. *J Eukaryot Microbiol* 49:201–208. <https://doi.org/10.1111/j.1550-7408.2002.tb00523.x>.
  72. Dan M, Sobel JD. 2007. Failure of nitazoxanide to cure trichomoniasis in three women. *Sex Transm Dis* 34:813–814.
  73. Edmondson MA, Joiner KS, Spencer JA, Riddell KP, Rodning SP, Gard JA, Givens MD. 2017. Impact of a killed *Trichomonas foetus* vaccine on clearance of the organism and subsequent fertility of heifers following experimental inoculation. *Theriogenology* 90:245–251. <https://doi.org/10.1016/j.theriogenology.2016.09.056>.
  74. Chapwanya A, Usman AY, Irons PC. 2016. Comparative aspects of immunity and vaccination in human and bovine trichomoniasis: a review. *Trop Anim Health Prod* 48:1–7. <https://doi.org/10.1007/s11250-015-0909-1>.
  75. Michi AN, Favetto PH, Kastelic J, Cobo ER. 2016. A review of sexually transmitted bovine trichomoniasis and campylobacteriosis affecting cattle reproductive health. *Theriogenology* 85:781–791. <https://doi.org/10.1016/j.theriogenology.2015.10.037>.
  76. Honigberg BM. 1987. Immunology of trichomonads, with emphasis on *Trichomonas vaginalis*: a review. *Acta Univ Carol Biol* 30:321–336.
  77. Smith JD, Garber GE. 2015. *Trichomonas vaginalis* infection induces vaginal CD4 cell infiltration in a mouse model: a vaccine strategy to reduce vaginal infection and HIV transmission. *J Infect Dis* 212:285–293. <https://doi.org/10.1093/infdis/jiv036>.
  78. Auvert B, Taljaard D, Lagarde E, Sobngwi-Tambekou J, Sitta R, Puren A. 2005. Randomized, controlled intervention trial of male circumcision for reduction of HIV infection risk: the ANRS 1265 Trial. *PLoS Med* 2:e298. <https://doi.org/10.1371/journal.pmed.0020298>.
  79. Bailey RC, Moses S, Parker CB, Agot K, Maclean I, Krieger JN, Williams CFM, Campbell RT, Ndinya-Achola JO. 2007. Male circumcision for HIV prevention in young men in Kisumu, Kenya: a randomised controlled trial. *Lancet* 369:643–656. [https://doi.org/10.1016/S0140-6736\(07\)60312-2](https://doi.org/10.1016/S0140-6736(07)60312-2).
  80. Weiss HA, Thomas SL, Munabi SK, Hayes RJ. 2006. Male circumcision and risk of syphilis, chancroid, and genital herpes: a systematic review

- and meta-analysis. *Sex Transm Infect* 82:101–109. <https://doi.org/10.1136/sti.2005.017442>.
81. Sobngwi-Tambekou J, Taljaard D, Nieuwoudt M, Lissouba P, Puren A, Auvert B. 2009. Male circumcision and *Neisseria gonorrhoeae*, *Chlamydia trachomatis* and *Trichomonas vaginalis*: observations after a randomised controlled trial for HIV prevention. *Sex Transm Infect* 85:116–120. <https://doi.org/10.1136/sti.2008.032334>.
  82. Gray RH, Kigozi G, Serwadda D, Makumbi F, Watya S, Nalugoda F, Kiwanuka N, Moulton LH, Chaudhary MA, Chen MZ, Sewankambo NK, Wabwire-Mangen F, Bacon MC, Williams CFM, Opendi P, Reynolds SJ, Laeyendecker O. 2007. Male circumcision for HIV prevention in men in Rakai, Uganda: a randomised trial. *Lancet* 369:657–666. [https://doi.org/10.1016/S0140-6736\(07\)60313-4](https://doi.org/10.1016/S0140-6736(07)60313-4).
  83. O'Farrell N, Morison L, Moodley P, Pillay K, Vanmali T, Quigley M, Hayes R, Sturm AW. 2006. Association between HIV and subpreputial penile wetness in uncircumcised men in South Africa. *J Acquir Immune Defic Syndr* 43:69–77. <https://doi.org/10.1097/01.qai.0000225014.61192.98>.
  84. Mehta SD, Moses S, Agot K, Parker C, Ndinya-Achola JO, Maclean I, Bailey RC. 2009. Adult male circumcision does not reduce the risk of incident *Neisseria gonorrhoeae*, *Chlamydia trachomatis*, or *Trichomonas vaginalis* infection: results from a randomized, controlled trial in Kenya. *J Infect Dis* 200:370–378. <https://doi.org/10.1086/600074>.
  85. Turner AN, Morrison CS, Padian NS, Kaufman JS, Behets FM, Salata RA, Mmiro FA, Mugerwa RD, Chipato T, Celentano DD, Rugpao S, Miller WC. 2008. Male circumcision and women's risk of incident chlamydial, gonococcal, and trichomonal infections. *Sex Transm Dis* 35:689–695. <https://doi.org/10.1097/OLQ.0b013e31816b1fcc>.
  86. Lushbaugh WB, Blossom AC, Shah PH, Banga AK, Jaynes JM, Cleary JD, Finley RW. 2000. Use of intravaginal microbicides to prevent acquisition of *Trichomonas vaginalis* infection in *Lactobacillus*-pretreated, estrogenized young mice. *Am J Trop Med Hyg* 63:284–289.
  87. Patton DL, Thwin SS, Meier A, Hooton TM, Stapleton AE, Eschenbach DA. 2000. Epithelial cell layer thickness and immune cell populations in the normal human vagina at different stages of the menstrual cycle. *Am J Obstet Gynecol* 183:967–973. <https://doi.org/10.1067/mob.2000.108857>.
  88. Mitchell CM, McLemore L, Westerberg K, Astronomo R, Smythe K, Gardella C, Mack M, Magaret A, Patton D, Agnew K, McElrath MJ, Hladik F, Eschenbach D. 2014. Long-term effect of depot medroxyprogesterone acetate on vaginal microbiota, epithelial thickness and HIV target cells. *J Infect Dis* 210:651–655. <https://doi.org/10.1093/infdis/jiu176>.
  89. Ayehunie S, Islam A, Cannon C, Landry T, Pudney J, Klausner M, Anderson DJ. 2015. Characterization of a hormone-responsive organotypic human vaginal tissue model: morphologic and immunologic effects. *Reprod Sci* 22:980–990. <https://doi.org/10.1177/1933719115570906>.
  90. Owen DH, Katz DF. 1999. A vaginal fluid simulant. *Contraception* 59:91–95. [https://doi.org/10.1016/S0010-7824\(99\)00010-4](https://doi.org/10.1016/S0010-7824(99)00010-4).
  91. Owen DH, Katz DF. 2005. A review of the physical and chemical properties of human semen and the formulation of a semen simulant. *J Androl* 26:459–469. <https://doi.org/10.2164/jandrol.04104>.
  92. Boris S, Barbés C. 2000. Role played by lactobacilli in controlling the population of vaginal pathogens. *Microbes Infect* 2:543–546. [https://doi.org/10.1016/S1286-4579\(00\)00313-0](https://doi.org/10.1016/S1286-4579(00)00313-0).
  93. Woolfson AD, Malcolm RK, Gallagher R. 2000. Drug delivery by the intravaginal route. *Crit Rev Ther Drug Carrier Syst* 17:509–555. <https://doi.org/10.1615/CritRevTherDrugCarrierSyst.v17.i5.30>.
  94. Patel LG, Warrington SJ, Pearson RM. 1983. Propranolol concentrations in plasma after insertion into the vagina. *Br Med J (Clin Res Ed)* 287:1247–1248. <https://doi.org/10.1136/bmj.287.6401.1247>.
  95. Vermesh M, Fossum GT, Kletzky OA. 1988. Vaginal bromocriptine: pharmacology and effect on serum prolactin in normal women. *Obstet Gynecol* 72:693–698.
  96. Ceddar M, Judd H. 1987. Nonoral routes of estrogen administration. *Obstet Gynecol Clin N Am* 14:269–298.
  97. Gershenfeld L. 1962. Povidone-iodine (PVP-I) as a trichomonicide. *Am J Pharm Sci Support Public Health* 134:324–331.
  98. Shook DM, Captain MC. 1963. Clinical study of a povidoneiodine regimen for resistant vaginitis. *Curr Ther Res* 5:256–263.
  99. Ratzan JJ. 1969. Monilial and trichomonal vaginitis—topical treatment with povidone-iodine preparations. *Calif Med* 110:24–27.
  100. Maneksha S. 1974. Comparison of povidone-iodine (Betadine) vaginal pessaries and lactic acid pessaries in the treatment of vaginitis. *J Int Med Res* 2:236–239. <https://doi.org/10.1177/030006057400200310>.
  101. Henderson JN, Tait IB. 1975. The use of povidone-iodine ('Betadine') pessaries in the treatment of candidal and trichomonal vaginitis. *Curr Med Res Opin* 3:157–162. <https://doi.org/10.1185/03007997509113664>.
  102. Muzny C, Barnes A, Mena L. 2012. Symptomatic *Trichomonas vaginalis* infection in the setting of severe nitroimidazole allergy: successful treatment with boric acid. *Sex Health* 9:389–391. <https://doi.org/10.1071/SH11114>.
  103. Wong CA, Wilson PD, Chew TA. 1990. Povidone-iodine in the treatment of metronidazole-resistant *Trichomonas vaginalis*. *Aust N Z J Obstet Gynecol* 30:169–171. <https://doi.org/10.1111/j.1479-828X.1990.tb03255.x>.
  104. Livengood CH, III, Lossick JG. 1991. Resolution of resistant vaginal trichomoniasis associated with the use of intra vaginal nonoxynol-9. *Obstet Gynecol* 78:954–955.
  105. Van Damme L, Ramjee G, Alary M, Vuylsteke B, Chandeying V, Rees H, Sirivongrangson P, Tshibaka LM, Ettiègne-Traoré V, Uaheowitchai C, Abdoul Karim SS, Mâsse B, Perriens J, Laga M. 2002. Effectiveness of COL-1492, a nonoxynol-9 vaginal gel, on HIV-1 transmission in female sex workers: a randomised controlled trial. *Lancet* 360:971–977. [https://doi.org/10.1016/S0140-6736\(02\)11079-8](https://doi.org/10.1016/S0140-6736(02)11079-8).
  106. Roddy RE, Zekeng L, Ryan KA, Tamoufé U, Tweedy KG. 2002. Effect of nonoxynol-9 gel on urogenital gonorrhoea and chlamydial infection: a randomized controlled trial. *JAMA* 287:1117–1122. <https://doi.org/10.1001/jama.287.9.1117>.
  107. Wilkinson D. 2002. Nonoxynol-9 fails to prevent STDs, but microbicide research continues. *Lancet* 360:962–963. [https://doi.org/10.1016/S0140-6736\(02\)11119-6](https://doi.org/10.1016/S0140-6736(02)11119-6).
  108. Van Damme L, Govinden R, Mirembe FM, Guédou F, Solomon S, Becker ML, Pradeep BS, Krishnan AK, Alary M, Pande B, Ramjee G, Deese J, Crucitti T, Taylor D. 2008. Lack of effectiveness of cellulose sulfate gel for the prevention of vaginal HIV transmission. *N Engl J Med* 359:463–472. <https://doi.org/10.1056/NEJMoa0707957>.
  109. Pereyra AJ, Lansing JD. 1964. Urogenital trichomoniasis: treatment with metronidazole in 2002 incarcerated women. *Obstet Gynecol* 24:499–508.
  110. Durel P, Couture J, Collart P. 1960. Flagyl (metronidazole). *Br J Vener Dis* 36:154–157.
  111. Tidwell BH, Lushbaugh WB, Laughlin MD, Cleary JD, Finley RW. 1994. A double-blind placebo-controlled trial of single-dose intravaginal versus single-dose oral metronidazole in the treatment of trichomonal vaginitis. *J Infect Dis* 170:242–246. <https://doi.org/10.1093/infdis/170.1.242>.
  112. Bouckaert S, Temmerman J, Voorspoels J, Van Kets H, Remon JP, Dhont M. 1995. Preliminary efficacy study of a bioadhesive vaginal metronidazole tablet in the treatment of bacterial vaginosis. *J Pharm Pharmacol* 47:970–971. <https://doi.org/10.1111/j.2042-7158.1995.tb03279.x>.
  113. Voorspoels J, Casteels M, Remon JP, Temmerman M. 2002. Local treatment of bacterial vaginosis with a bioadhesive metronidazole tablet. *Eur J Obstet Gynecol Reprod Biol* 105:64–66. [https://doi.org/10.1016/S0301-2115\(02\)00110-0](https://doi.org/10.1016/S0301-2115(02)00110-0).
  114. duBouchet L, McGregor JA, Ismail M, McCormack WM. 1998. A pilot study of metronidazole vaginal gel versus oral metronidazole for the treatment of *Trichomonas vaginalis* vaginitis. *Sex Transm Dis* 25:176–179. <https://doi.org/10.1097/00007435-199803000-00012>.
  115. Kükner S, Ergin T, Cicek N, Uğur M, Yeşilyurt H, Gökmen O. 1996. Treatment of vaginitis. *Int J Gynaecol Obstet* 52:43–47. [https://doi.org/10.1016/0020-7292\(95\)02531-6](https://doi.org/10.1016/0020-7292(95)02531-6).
  116. Coelho DD. 1997. Metronidazole resistant trichomoniasis successfully treated with paromomycin. *Genitourin Med* 73:397–398.
  117. Nyirjesy P, Sobel JD, Weitz MV, Leaman DJ, Gelone SP. 1998. Difficult-to-treat trichomoniasis: results with paromomycin cream. *Clin Infect Dis* 26:986–988. <https://doi.org/10.1086/513951>.
  118. Helms DJ, Mosure DJ, Secor WE, Workowski KA. 2008. Management of *Trichomonas vaginalis* in women with suspected metronidazole hypersensitivity. *Am J Obstet Gynecol* 198:370.e1–370.e7. <https://doi.org/10.1016/j.jog.2007.10.795>.
  119. Dumont M, Croizat B, Douillet P. 1964. Traitement des vaginites a trichomonas par la paromomycine. *Gynecol Prat* 15:247–251.
  120. Aggarwal A, Shier RM. 2008. Recalcitrant infections successfully treated with vaginal acidification. *J Obstet Gynaecol Can* 30:55–58. [https://doi.org/10.1016/S1701-2163\(16\)32714-1](https://doi.org/10.1016/S1701-2163(16)32714-1).
  121. Nyirjesy P, Gilbert J, Mulcahy LJ. 2011. Resistant trichomoniasis: successful treatment with combination therapy. *Sex Transm Dis* 38:962–963. <https://doi.org/10.1097/OLQ.0b013e31822037e4>.
  122. Lossick JG, Kent HL. 1991. Trichomoniasis: trends in diagnosis and



- management. *Am J Obstet Gynecol* 165:1217–1222. [https://doi.org/10.1016/S0002-9378\(12\)90730-9](https://doi.org/10.1016/S0002-9378(12)90730-9).
123. Ofner CM, Klech-Gelotte CM. 2013. Gels and jellies, p 1875–1890. In Swarbrick J (ed), *Encyclopedia of pharmaceutical technology*, 3rd ed. Taylor & Francis Group, London, United Kingdom. <http://www.tandfonline.com/doi/abs/10.1081/E-EPT3-100200006>.
  124. Neves JD, Sarmiento B (ed). 2014. Drug delivery and development of anti-HIV microbicides. Taylor & Francis Group, London, United Kingdom.
  125. Pradines B, Gallard J-F, Iorga BI, Gueutin C, Loiseau PM, Ponchel G, Bouchemal K. 2014. Investigation of the complexation of albendazole with cyclodextrins for the design of new antiparasitic formulations. *Carbohydr Res* 398:50–55. <https://doi.org/10.1016/j.carres.2014.06.008>.
  126. Pradines B, Gallard J-F, Iorga BI, Gueutin C, Ponchel G, Loiseau PM, Bouchemal K. 2015. The unexpected increase of clotrimazole apparent solubility using randomly methylated  $\beta$ -cyclodextrin. *J Mol Recognit* 28:96–102. <https://doi.org/10.1002/jmr.2432>.
  127. Pradines B, Bories C, Vauthier C, Ponchel G, Loiseau PM, Bouchemal K. 2015. Drug-free nanoparticles are active against *Trichomonas vaginalis* and non-toxic towards pig vaginal mucosa. *Pharm Res* 32:1229–1236. <https://doi.org/10.1007/s11095-014-1528-7>.
  128. Das Neves J, Bahia MF. 2006. Gels as vaginal drug delivery systems. *Int J Pharm* 318:1–14. <https://doi.org/10.1016/j.ijpharm.2006.03.012>.
  129. Garg S, Goldman D, Krumme M, Rohan LC, Smoot S, Friend DR. 2010. Advances in development, scale-up and manufacturing of microbicide gels, films, and tablets. *Antiviral Res* 88(Suppl):S19–S29. <https://doi.org/10.1016/j.antiviral.2010.09.010>.
  130. Vermani K, Garg S. 2000. The scope and potential of vaginal drug delivery. *Pharm Sci Technol Today* 3:359–364. [https://doi.org/10.1016/S1461-5347\(00\)00296-0](https://doi.org/10.1016/S1461-5347(00)00296-0).
  131. Justin-Temu M, Damian F, Kinget R, Van Den Mooter G. 2004. Intravaginal gels as drug delivery systems. *J Womens Health* 13:834–844. <https://doi.org/10.1089/jwh.2004.13.834>.
  132. Pradines B, Djabourov M, Vauthier C, Loiseau PM, Ponchel G, Bouchemal K. 2015. Gelation and micellization behaviors of pluronic F127 hydrogel containing poly(isobutylcyanoacrylate) nanoparticles specifically-designed for mucosal application. *Colloids Surf B Biointerfaces* 135:669–676. <https://doi.org/10.1016/j.colsurfb.2015.08.021>.
  133. Malli S, Bories C, Pradines B, Loiseau PM, Ponchel G, Bouchemal K. 2017. *In situ* forming pluronic F127/chitosan hydrogel limits metronidazole transmucosal absorption. *Eur J Pharm Biopharm* 112:143–147. <https://doi.org/10.1016/j.ejpb.2016.11.024>.
  134. Aka-Any-Grah A, Bouchemal K, Koffi A, Agnely F, Zhang M, Djabourov M, Ponchel G. 2010. Formulation of mucoadhesive vaginal hydrogels insensitive to dilution with vaginal fluids. *Eur J Pharm Biopharm* 76: 296–303. <https://doi.org/10.1016/j.ejpb.2010.07.004>.
  135. Grisin T, Bories C, Bombardi M, Loiseau PM, Rouffiac V, Solgadi A, Mallet J-M, Ponchel G, Bouchemal K. 2017. Supramolecular chitosan micro-platelets synergistically enhance anti-*Candida albicans* activity of amphotericin B using an immunocompetent murine model. *Pharm Res* 34:1067–1082. <https://doi.org/10.1007/s11095-017-2117-3>.
  136. Grisin T, Bories C, Loiseau PM, Bouchemal K. 2017. Cyclodextrin-mediated self-associating chitosan micro-platelets act as a drug booster against *Candida glabrata* mucosal infection in immunocompetent mice. *Int J Pharm* 519:381–389. <https://doi.org/10.1016/j.ijpharm.2017.01.048>.
  137. WHO. 2012. Global incidence and prevalence of selected curable sexually transmitted infections—2008. WHO, Geneva, Switzerland. [http://apps.who.int/iris/bitstream/10665/75181/1/9789241503839\\_eng.pdf](http://apps.who.int/iris/bitstream/10665/75181/1/9789241503839_eng.pdf).

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